Product Data Sheet

K562 A-FLX™ FFPE Cell Pellet

GENERAL INFORMATION

Product Name:	K562 A-FLX™ FFPE Cell Pellet
Reference Number:	3070-0110 Block
	3070-0120 Slide (4µm)
	3070-0130 FFPE scroll (20µm)
Date of Manufacture:	See product label
Lot Number:	See product label
Intended Use:	For research use only

DESCRIPTION

Cell Line:	K562
Tissue of Origin:	Leukemia
Culturing Condition:	RPMI-1640 supplemented with 10% FBS at 37°C with 5% CO_2
Fixation Condition:	10% neutral buffered formalin (NBF) for 24 hours at 24-27°C
Product Format:	
Block:	Paraffin embedded block.
	Pellet diameter: 5mm
	Sections: 300+ sections at 4 µm
Slide:	One unstained section mounted on Superfrost™ Plus slide. Section thickness: 4µm
FFPE Scroll:	One FFPE section in DNase/RNase free tube. Section thickness: 20µm

SCHEMATICS

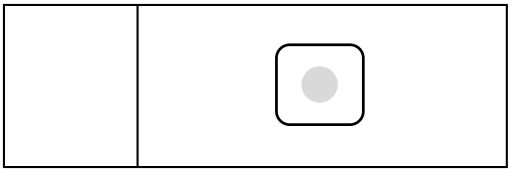


Illustration of an FFPE slide

STORAGE CONDITION

Block & Slide:	2-8 ˚C with desiccation (Recommended).
	For extended shelf life, store samples at -15 ˚C to -5 ˚C with desiccation.

*RECOMMENDED STABILITY RE-TESTING SCHEDULE:

Block	5 years
Scroll	1 year
Slide	1 year

*FFPE sample stability is not universally established and is biomarker and assay dependent. More frequent re-testing may be needed for certain labile biomolecules.

SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

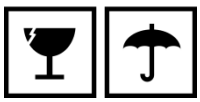
RECOMMENDED PROCEDURES

Staining using FFPE slides:

1. Bake slides at 60°C for 30-60 min.
2. Deparaffinize 3 times in Xylene or Xylene substitute for 5 min each time.
3. Rinse 2 times in 100% ethanol for 1 min each.
4. Rehydrate in ethanol series (95% 1 min, 70% 1 min, distilled H_2O 2 times for 1 min each).
5. Proceed to staining protocol.

Biomolecule extraction from FFPE scrolls:

1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 30 sec.
2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
3. Repeat step 1 and 2 for a total of 3 times
4. Add 1ml 100% ethanol and mix by vortexing.
5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
6. Repeat step 4 and 5.
7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
8. Open the tube and dry at room temperature or 37°C for 30min. Ensure that ethanol has completely evaporated.
9. Proceed to extraction protocol.



Acepix Biosciences, Inc.
32990 Alvarado-Niles Road, Suite 910 Union City· CA · 94587 · USA
Tel: 1 (888) 866-0580 · Web: www.acepixbio.com