

# **Product Data Sheet**

# SW626 A-FLX™ FFPE Cell Pellet

## GENERAL INFORMATION

Product Name:	SW626 A-FLX™ FFPE Cell Pellet	
Reference Number:	3090-0610	Block
	3090-0620	Slide (4µm)
	3090-0630	FFPE scroll (20µm)
Date of Manufacture:	See product label	
Lot Number:	See product label	
Intended Use:	For research use only	

#### DESCRIPTION

Cell Line:	SW626
Tissue of Origin:	Ovary
Culturing Condition:	RPMI-1640 supplemented with 20% FBS at 37°C with 5% $\mathrm{CO}_2$
Fixation Condition:	10% neutral buffered formalin (NBF) for 24 hours at 24- 27°C
Product Format:	
Block:	Paraffin embedded block.
	Pellet diameter: 5mm
	Sections: 300+ sections at 4 µm
Slide:	One unstained section mounted on Superfrost™ Plus slide. Section thickness: 4µm
FFPE Scroll:	One FFPE section in DNase/RNase free tube. Section thickness: 20µm

#### **SCHEMATICS**

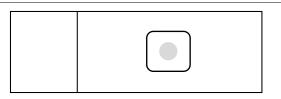


Illustration of an FFPE slide

# STORAGE CONDITION

Block & Slide: 2-8 ° C with desiccation (Recommended). For extended shelf life, store samples at -15 ° C to -5 ° C with desiccation.

#### \*RECOMMENDED STABILITY RE-TESTING SCHEDULE:

Block	5 years
Scroll	1 year
Slide	1 year

\*FFPE sample stability is not universally established and is biomarker and assay dependent. More frequent re-testing may be needed for certain labile biomolecules.

## SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

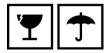
#### RECOMMENDED PROCEDURES

#### Staining using FFPE slides:

- 1. Bake slides at 60°C for 30-60 min.
- 2. Deparaffinize 3 times in Xylene or Xylene substitute for 5 min each time.
- 3. Rinse 2 times in 100% ethanol for 1 min each.
- 4. Rehydrate in ethanol series (95% 1 min, 70% 1 min, distilled  $H_2O$  2 times for 1 min each).
- 5. Proceed to staining protocol.

#### Biomolecule extraction from FFPE scrolls:

- 1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 30 sec.
- 2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 3. Repeat step 1 and 2 for a total of 3 times
- 4. Add 1ml 100% ethanol and mix by vortexing.
- 5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 6. Repeat step 4 and 5.
- 7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
- 8. Open the tube and dry at room temperature or 37°C for 30min. Ensure that ethanol has completely evaporated.
- 9. Proceed to extraction protocol.



#### Acepix Biosciences, Inc.

32990 Alvarado-Niles Road, Suite 910 Union City· CA · 94587 · USA Tel: 1 (888) 866-0580 · Web: www.acepixbio.com