

# **Product Data Sheet**

# A498 A-FLX™ FFPE Cell Pellet

#### GENERAL INFORMATION

Product Name: A498 A-FLX™ FFPE Cell Pellet

Reference Number: 3100-0610 Block

3100-0620 Slide (4µm)

3100-0630 FFPE scroll (20µm)

Date of Manufacture: See product label
Lot Number: See product label
Intended Use: For research use only

#### **DESCRIPTION**

Cell Line: A498
Tissue of Origin: Kidney

Culturing Condition: DMEM supplemented with 10% FBS at 37°C with 5%

 $CO_2$ 

Fixation Condition: 10% neutral buffered formalin (NBF) for 24 hours at 24-

27°C

Product Format:

Block: Paraffin embedded block.

Pellet diameter: 5mm

Sections: 300+ sections at 4 µm

Slide: One unstained section mounted on Superfrost™ Plus

slide. Section thickness: 4µm

FFPE Scroll: One FFPE section in DNase/RNase free tube. Section

thickness: 20µm

# **SCHEMATICS**

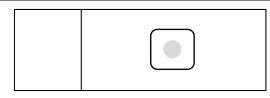


Illustration of an FFPE slide

# STORAGE CONDITION

Block & Slide: 2-8 ° C with desiccation (Recommended).

For extended shelf life, store samples at -15 °C to

-5 °C with desiccation.

# \*RECOMMENDED STABILITY RE-TESTING SCHEDULE:

Block	5 years
Scroll	1 year
Slide	1 year

<sup>\*</sup>FFPE sample stability is not universally established and is biomarker and assay dependent. More frequent re-testing may be needed for certain labile biomolecules.

#### SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

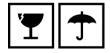
## RECOMMENDED PROCEDURES

# Staining using FFPE slides:

- 1. Bake slides at 60°C for 30-60 min.
- 2. Deparaffinize 3 times in Xylene or Xylene substitute for 5 min each time.
- 3. Rinse 2 times in 100% ethanol for 1 min each.
- 4. Rehydrate in ethanol series (95% 1 min, 70% 1 min, distilled  $H_2O$  2 times for 1 min each).
- 5. Proceed to staining protocol.

#### Biomolecule extraction from FFPE scrolls:

- 1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 30 sec.
- 2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 3. Repeat step 1 and 2 for a total of 3 times
- 4. Add 1ml 100% ethanol and mix by vortexing.
- 5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 6. Repeat step 4 and 5.
- 7. Carefully remove any residual ethanol in the tube without disturbing the
- 8. Open the tube and dry at room temperature or 37°C for 30min. Ensure that ethanol has completely evaporated.
- 9. Proceed to extraction protocol.





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