

## AML12 A-FLX™ FFPE Cell Pellet

### GENERAL INFORMATION

Product Name: AML12 A-FLX™ FFPE Cell Pellet  
 Reference Number: 3500-2110 Block  
 3500-2120 Slide (4µm)  
 3500-2130 FFPE scroll (20µm)  
 Date of Manufacture: See product label  
 Lot Number: See product label  
 Intended Use: For research use only

### DESCRIPTION

Cell Line: AML12  
 Tissue of Origin: Animal  
 Culturing Condition: Refer to ATCC  
 Fixation Condition: 10% neutral buffered formalin (NBF) for 24 hours at 24-27°C  
 Product Format:  
 Block: Paraffin embedded block.  
 Pellet diameter: 5mm  
 Sections: 300+ sections at 4 µm  
 Slide: One unstained section mounted on Superfrost™ Plus slide. Section thickness: 4µm  
 FFPE Scroll: One FFPE section in DNase/RNase free tube. Section thickness: 20µm

### SCHEMATICS

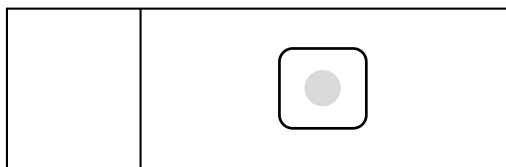


Illustration of an FFPE slide

### STORAGE CONDITION

Block & Slide: 2-8 °C with desiccation (Recommended).  
 For extended shelf life, store samples at -15 °C to -5 °C with desiccation.

### \*RECOMMENDED STABILITY RE-TESTING SCHEDULE:

Block	5 years
Scroll	1 year
Slide	1 year

\*FFPE sample stability is not universally established and is biomarker and assay dependent. More frequent re-testing may be needed for certain labile biomolecules.

### SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

### RECOMMENDED PROCEDURES

#### Staining using FFPE slides:

1. Bake slides at 60°C for 30-60 min.
2. Deparaffinize 3 times in Xylene or Xylene substitute for 5 min each time.
3. Rinse 2 times in 100% ethanol for 1 min each.
4. Rehydrate in ethanol series (95% 1 min, 70% 1 min, distilled H<sub>2</sub>O 2 times for 1 min each).
5. Proceed to staining protocol.

#### Biomolecule extraction from FFPE scrolls:

1. Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 30 sec.
2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
3. Repeat step 1 and 2 for a total of 3 times
4. Add 1ml 100% ethanol and mix by vortexing.
5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
6. Repeat step 4 and 5.
7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
8. Open the tube and dry at room temperature or 37°C for 30min. Ensure that ethanol has completely evaporated.
9. Proceed to extraction protocol.



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