

Produktinformation



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TECHNICAL DATA SHEET

THUNDER[™] Phospho-elF4E (S209) + Total eIF4E TR-FRET Cell Signaling Assay Kit

CATALOG NUMBERS KIT-EIF4EPT-500

PRODUCT DESCRIPTION

This assay kit measures intracellular levels of

phospho-eIF4E (S209) and total eIF4E protein

in cell lysates using a simple, rapid and sensitive

immunoassay based on the homogeneous (no-

wash) THUNDER™ TR-FRET technology. The kit is

compatible with both adherent and suspension cells.

400 points for phospho-eIF4E and 100 points for total eIF4E

Store at -80°C For research use only. Not for use in diagnostic procedures.

This assay kit contains two specific

and selective antibody pairs, one that

recognizes eIF4E phosphorylated at

Ser209 and another that recognizes total

(both phosphorylated and unphospho-

SPECIFICITY

rylated) eIF4E.

SPECIES REACTIVITY

Human (Swiss-Prot Acc. P06730 and Entrez-Gene Id 1977).

Other species should be tested on a case-by-case basis.

TR-FRET ASSAY PRINCIPLE

The Phospho-elF4E (S209) + Total elF4E assay kit is a homogeneous time-resolved Förster resonance energy transfer (TR-FRET) sandwich immunoassay (Figure 1). The THUNDER™ Cell Signaling assay workflow consists of 3 steps (Figure 2). Following cell treatment, cells are first lysed with the specific Lysis Buffer provided in the kit. Then Phospho-eIF4E (S209) and Total eIF4E in the cell lysates are detected in separate wells with two pairs of fluorophore-labeled antibodies in a simple "add-incubate-measure" format (single-step reagent addition; no wash steps). For detection of the phosphorylated protein, one antibody is labeled with a donor fluorophore (Europium chelate; Eu-Ab1) and the second with a far-red acceptor fluorophore (FR-Ab2). The same approach is used for the second antibody pair detecting the total protein (Eu-Ab3 and FR-Ab4). The binding of the two matched labeled antibodies to distinct epitopes on the target protein (either phospho-eIF4E or total eIF4E) takes place in solution and brings the two dyes into close proximity. Excitation of the donor Europium chelate molecules with a flash lamp (320 or 340 nm) or a laser (337 nm) triggers a FRET from the donor to the acceptor molecules, which in turn emit a TR-FRET signal at 665 nm. Residual energy from the Eu chelate generates light at 615 nm. The signal at 665 nm is proportional to the concentration of Phospho-eIF4E (S209) and Total eIF4E in the cell lysate. Data can be expressed as either the signal at 665 nm or the 665 nm/615 nm ratio.

STEP 1	STEP 2	STEP 3
Cell treatment	Cell lysis	Protein detection
 Seed adherent cells in culture plate Add media +/- compound Incubate for optimized time 	Remove media Add 1X Supplemented Lysis Buffer 2 Incubate for 30 min	 Transfer lysate (15 μL) to detection plate Add 4X Antibody Mix (5 μL) Incubate for 4 h Read TR-FRET signal

Figure 2 Assay workflow using the 2-plate (transfer) protocol

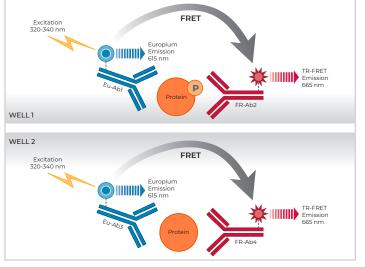


Fig. 1 Schematic representation of the TR-FRET cell signaling assay principle.

KIT COMPONENTS	500 points*
Eu-labeled phospho-eIF4E (S209) antibody (Eu-Ab1)	20 µL
Acceptor-labeled phospho-eIF4E (S209) antibody (FR-Ab2)	80 µL
Eu-labeled total-eIF4E antibody (Eu-Ab3)	5 µL
Acceptor-labeled total-eIF4E antibody (FR-Ab4)	20 µL
Lysis Buffer 2 (5X)	5 mL
Detection Buffer (10X)	250 µL
Positive control cell lysate	500 µL

* The number of assay points is based on an assay volume of 20 µL in half-area 96-well or low-volume 384-well assay plates using the kit components at the recommended concentrations (refer to the User Manual).





VALIDATION DATA

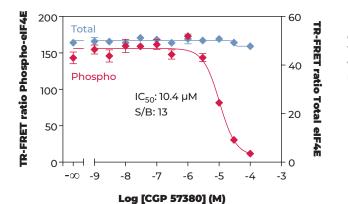
IN HEK293 CELLS

This assay kit has been validated for the relative quantification of phospho-eIF4E (S209) and total eIF4E in HEK293 cell lysates using the 2-plate assay protocol.

- Adherent cells were cultured overnight in a 96-well tissue culture plate coated with poly-L-lysine (EMEM +10% FBS).
- \cdot Following cell treatment, the media was removed and cells were lysed with the 1X Lysis Buffer 2 (50 μ L) supplemented with the phosphatase inhibitors sodium fluoride (1 mM) and sodium orthovanadate (2 mM).
- \cdot Following a 30-min incubation at room temperature (RT) on an

orbital shaker (400 rpm), lysates (15 μ L) were then transferred to a 384-well assay plate followed by addition to separate wells of either the labeled antibodies Eu-Ab1 and FR-Ab2 (5 μ L) for detection of phospho-eIF4E (S209) or Eu-Ab3 and FR-Ab4 (5 μ L) for detection of total eIF4E.

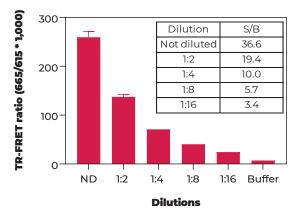
• The plate was incubated at RT for **4 hours** and the TR-FRET signal was recorded at 665 and 615 nm (EnVision®; lamp excitation).



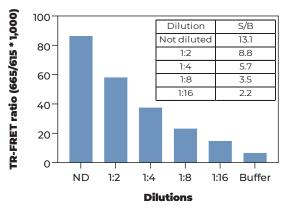
INHIBITION OF PHOSPHO-eIF4E (S209)

HEK293 cells (50,000 cells/well; in triplicate) were incubated with serial dilutions of the inhibitor CGP 57380 for 30 min at 37°C. Cells were then stimulated with 10 nM of EGF for 30 min at 37°C. Data show that treatment of HEK293 cells with CGP 57380 inhibits phosphorylation of eIF4E at S209 by EGF, but does not affect the levels of total eIF4E.

HEK293 CONTROL LYSATE TITRATION (QC TEST) PHOSPHO-eIF4E (S209)



HEK293 CONTROL LYSATE TITRATION (QC TEST) TOTAL eIF4E



Quality Control: the Phospho-eIF4E (S209) + Total eIF4E assay kit is routinely tested against EGF-treated HEK293 lysates. HEK293 cells were cultured in a TI75 flask to 85% confluence and stimulated with 30 nM EGF for 30 min at 37°C. Following cell lysis using 8 mL of 1X Lysis Buffer 2, lysates were serially diluted with 1X Lysis Buffer 2 and tested in triplicate and in separate wells for phospho-eIF4E (S209) and total eIF4E. Data show a linear relationship between lysate dilutions and TR-FRET ratio values.



FOR MORE INFORMATION ON DEVELOPING AND OPTIMIZING TR-FRET CELL SIGNALING ASSAYS, CONSULT THE USER MANUAL.