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Diagnostik & molekulare Diagnostik



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Data Sheet

Fluorogenic HDAC7 Assay Kit Catalog #: 50067

DESCRIPTION: The *Fluorogenic HDAC7 Assay Kit* is a complete assay system designed to measure histone deacetylase 7 (HDAC7) activity for screening and profiling applications. It comes in a convenient 96-well format, with all the reagents necessary for 100 fluorescent HDAC7 activity measurements. In addition, the kit includes purified HDAC7 enzyme and a potent HDAC inhibitor, Trichostatin A, for use as a positive and negative control. The *Fluorogenic HDAC7 Assay Kit* is based on a unique fluorogenic substrate and developer combination. This assay method eliminates dealing with the radioactivity, extraction, and chromatography aspects of traditional assays. Using this kit, only two simple steps on a microtiter plate are needed to analyze the HDAC7 activity level. First, the HDAC fluorometric substrate, containing an acetylated lysine side chain, is incubated with purified HDAC7. The deacetylation sensitizes the substrate so subsequent treatment with the Lysine Developer produces a fluorophore that can then be measured using a fluorescence reader.

HDACs regulate cellular processes by catalyzing the hydrolysis of an acetyl group from acetyllysines in modified proteins. In the HDAC assay, fluorescent-dye molecules are attached to a peptide containing acetyllysine. Attachment to the peptide quenches the fluorescence of the dye. After treatment of the peptide with an HDAC, the reaction is mixed with a development solution that is specific for nonacetylated lysines. If the acetyl group has been removed from the lysine by the HDAC, this solution will release the dye allowing for fluorescence. Fluorescence is therefore directly related to HDAC activity.

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COMPONENTS:

| Cat. # | Component | Amount | Storage | |
|--------|--|---------|---------|---------|
| 50007 | HDAC7 human recombinant enzyme | 1 µg | -80°C | |
| 50040 | Fluorogenic HDAC substrate class 2A (5 mM) | 50 µl | -80°C | |
| 50030 | 2x HDAC Developer (contains Trichostatin A) | 6 ml | -80°C | Avoid |
| | (50 μM) | | | freeze/ |
| | Trichostatin A (1 mM) in DMSO | 100 µl | -20°C | thaw |
| 50031 | HDAC Assay Buffer | 10 ml | -20°C | cycles! |
| 79685 | black, low binding NUNC black microtiter plate | 1 plate | Room | |
| | | | temp. | |

MATERIALS OR INSTRUMENTS REQUIRED BUT NOT SUPPLIED:

0.1% solution (1 mg/ml) of bovine serum albumin (BSA) in water Fluorimeter capable of excitation at 350-380 nm and detection at 440-460 nm Adjustable micropipettor and sterile tips Rotating or rocker platform

APPLICATIONS: Great for studying enzyme kinetics and screening small molecular inhibitors for drug discovery and HTS applications.

STABILITY: One year from date of receipt when stored as directed.

REFERENCE(S): Ontoria, J.M., et al., J. Med. Chem. 2009 Nov 12;52(21):6782-9.

ASSAY PROTOCOL:

Immediately prior to assay:

- 1) Dilute **Trichostatin A** 1 mM stock 10-fold with **HDAC Assay Buffer** to make a 100 μM solution. *Make only sufficient quantity needed for the assay; store remaining 1 mM Trichostatin A* stock solution in aliquots at -80°C.
- 2) Dilute **HDAC** substrate 5 mM stock 250-fold with **HDAC** Assay Buffer to make a 20 μM solution. Make only sufficient quantity needed for the assay; store remaining 5 mM stock solution in aliquots at -80°C.
- 3) Dilute **HDAC7** in **HDAC** Assay Buffer to 0.05 ng/µl (0.25 ng/reaction)*. Aliquot any remaining enzyme and store undiluted at -80°C. Keep diluted enzyme on ice. Discard any remaining diluted enzyme after use. *Note: optimal enzyme concentration may vary with the specific activity of the enzyme.



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Step 1:

In duplicate, add the reaction mixtures (below) to the microtiter black plate as follows:

- 1) Prepare the master mixture: N wells \times (5 μ l HDAC substrate (20 μ M) + 5 μ l BSA (1 mg/ml) + 30 μ l HDAC Assay Buffer). Add 40 μ l of master mixture to all wells.
- 2) Add 5 µl of inhibitor solution of each well designated "Test Inhibitor." For the "Positive Control" and "Blank," add 5 µl of the same solution without inhibitor (inhibitor buffer). Add 5 µl of diluted **Trichostatin A** (100 µM) to the wells designated "Inhibitor Control." Keep final DMSO concentration at or below 1%.
- 3) Add 5 µl of **HDAC Assay Buffer** to the wells designated "Blank."
- 4) Initiate reaction by adding 5 μl of diluted **HDAC7 enzyme** to the wells designated "Positive Control," "Test Inhibitor," and "Inhibitor Control." Incubate at 37°C for 30 min.

| | "Blank" | Positive Control | Test Inhibitor | Inhibitor Control |
|---------------------------------|---------|---------------------|-------------------|----------------------|
| HDAC substrate (20 μM) | 5 µl | 5 µl | 5 µl | 5 µl |
| BSA (1 mg/ml) | 5 µl | 5 µl | 5 µl | 5 µl |
| HDAC Assay Buffer | 35 µl | 30 µl | 30 µl | 30 µl |
| Diluted Trichostatin A (100 μM) | _ | _ | _ | 5 µl |
| Test Inhibitor | _ | _ | 5 µl | _ |
| Inhibitor buffer (no inhibitor) | 5 µl | 5 µl | _ | _ |
| Diluted HDAC7 (0.05 ng/µl) | _ | 5 µl | 5 μl | 5 µl |
| Total | 50 µl | 50 µl | 50 µl | 50 µl |

Step 2:

Add 50 µl of undiluted **HDAC Assay Developer (2x)** to each well. Incubate the plate at room temperature for 15 minutes.

Step 3:

Read sample in a microtiter plate-reading fluorimeter capable of excitation at a wavelength in the range of 350-380 nm and detection of emitted light in the range of 440-460 nm. "Blank" value is subtracted from all other values.



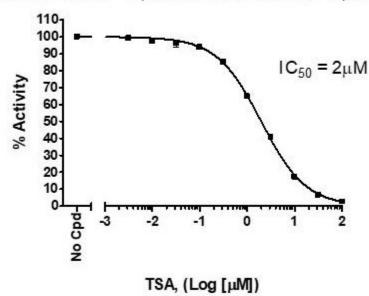
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Example of Assay Results:

HDAC7 Activity

Substrate Conc. = 2 μM Class 2a Substrate 1 (50040)



HDAC7 enzyme activity, measured using the *Fluorogenic HDAC7 Assay Kit*, BPS Bioscience Catalog #50067. Fluorescence was measured using a Bio-Tek fluorescent microplate reader. *Data shown is lot-specific. For lot-specific information, please contact BPS Bioscience, Inc. at* info@bpsbioscience.com.



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RELATED PRODUCTS:

| RELATED I RODOOTO. | | | | | | | |
|--------------------|--|--|--|--|--|--|--|
| Catalog # | <u>Size</u> | | | | | | |
| 50051 | 50 µg | | | | | | |
| 50002 | 50 µg | | | | | | |
| 50052 | 50 µg | | | | | | |
| 50003 | 50 µg | | | | | | |
| 50004 | 10 µg | | | | | | |
| 50005 | 10 µg | | | | | | |
| 50056 | 50 µg | | | | | | |
| 50006 | 50 µg | | | | | | |
| 50046 | 50 µg | | | | | | |
| 50066 | 50 µg | | | | | | |
| 50007 | 10 µg | | | | | | |
| 50008 | 50 µg | | | | | | |
| 50009 | 10 µg | | | | | | |
| 50010 | 50 µg | | | | | | |
| 50011 | 50 µg | | | | | | |
| 50033 | 96 reactions | | | | | | |
| 50034 | 96 reactions | | | | | | |
| 50041 | 96 reactions | | | | | | |
| 50062 | 96 reactions | | | | | | |
| 50073 | 96 reactions | | | | | | |
| 50064 | 96 reactions | | | | | | |
| 50065 | 96 reactions | | | | | | |
| 50076 | 96 reactions | | | | | | |
| 50068 | 96 reactions | | | | | | |
| 50069 | 96 reactions | | | | | | |
| | 50051 50002 50003 50004 50005 50056 50006 50046 50007 50008 50009 50010 50011 50033 50034 50041 50062 50073 50064 50065 50076 50068 | | | | | | |