

Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

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Lieferung & Zahlungsart

siehe unsere Liefer- und Versandbedingungen

Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
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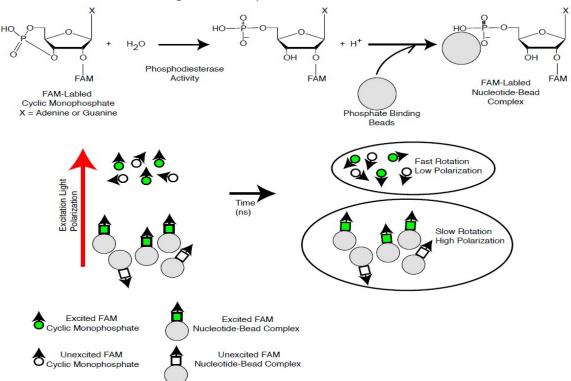


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Data Sheet PDE2A Assay Kit Catalog # 60321

BACKGROUND: Phosphodiesterases (PDEs) play an important role in the dynamic regulation of cAMP and cAMP signaling. PDE2A, also known as cAMP-stimulated phosphodiesterase, hydrolyzes cyclic nucleotides cAMP (Km = 2.4μ M) and cAMP, and is involved in the regulation of blood pressure and fluid homeostasis.

DESCRIPTION: The PDE2A Assay Kit is designed for identification of PDE2A inhibitors using fluorescence polarization. The assay is based on the binding of a fluorescent nucleotide monophosphate generated by PDE2A to the binding agent. PDE2A catalyzes the hydrolysis of the phosphodiester bond in dye-labeled cyclic adenosine monophosphate (cAMP). Nanoparticle beads selectively bind the phosphate group in the nucleotide product. This increases the size of the nucleotide relative to unreacted cAMP. Since the degree of polarization of a fluorophore is inversely related to its molecular rotation, dyes attached to the slowly-rotating nucleotide-bead complexes will not have time to reorient and therefore will emit highly polarized light. Conversely, dyes attached to the rapidly-rotating cyclic monophosphates will obtain random orientations and emit light with low polarization.



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The PDE2A inhibitor screening assay kit comes in a convenient 384-well format, including purified PDE2A1 enzyme, fluorescently labeled PDE2A substrate (cAMP), binding agent, and PDE assay buffer for 384 enzyme reactions. The key to the *PDE2A Assay Kit* is the specific binding agent. Using this kit, only two simple steps on a microtiter plate are required for PDE2A reactions. First, the fluorescently labeled cAMP is incubated with a sample containing PDE2A1 for 1 hour. Second, a binding agent is added to the reaction mix to produce a change in fluorescent polarization that can then be measured using a fluorescence reader.

COMPONENTS:

| Catalog # | Component | Amount | Storage | |
|-----------|------------------------------------|--------|---------|--------------|
| 60020 | PDE2A1 recombinant enzyme | 1 µg | -80°C | |
| 60200 | FAM-Cyclic-3', 5'-AMP: 20 µM | 50 µl | -80°C | |
| 60393 | PDE assay buffer | 25 ml | -20°C | (Avoid |
| 60390 | Binding Agent | 250 µl | +4°C | freeze/ thaw |
| 60391 | Binding Agent Diluent (cAMP) | 25 ml | +4°C | cycles!) |
| 79685 | Black, low binding, 384 microtiter | 1 | Room | |
| | plate | | temp. | |

MATERIALS OR INSTRUMENTS REQUIRED BUT NOT SUPPLIED:

Fluorescent microplate reader capable of measuring fluorescence polarization

APPLICATIONS: Great for studying enzyme kinetics and screening small molecular inhibitors for drug discovery and HTS applications.

STABILITY: 6 months from date of receipt when stored as directed.

REFERENCE(S):

Maurice DH. Front. Biosci. 2005; 10:1221-8.

ASSAY PROTOCOL:

All samples and controls should be tested in duplicate.

Step 1:

- 1) Dilute 20 µM **FAM-Cyclic-3',5'-AMP** substrate stock solution 100-fold with **PDE Assay Buffer** to make a 200 nM solution. Make only a sufficient quantity needed for the assay; store remaining stock solution in aliquots at -20°C.
- 2) Add 12.5 μl of diluted **FAM-Cyclic-3',5'-AMP** (200 nM) to each well designated "Substrate Control", "Positive Control", and "Test Inhibitor". Add 12.5 μl of **PDE Assay Buffer** to each well designated "Blank".

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- 3) Add 2.5 µl of inhibitor solution to each well designated "Test Inhibitor". Add 2.5 µl of the same solution without inhibitor (inhibitor buffer) to the wells labeled "Blank", "Substrate Control" and "Positive Control".
- 4) Add 10 µl of **PDE Assay Buffer** to the wells designated as the "Blank" and "Substrate Control".
- 5) Thaw **PDE2A1** on ice. Upon first thaw, briefly spin tube containing enzyme to recover the full contents of the tube. Aliquot **PDE2A1** enzyme into single-use aliquots. Store remaining undiluted enzyme in aliquots at -80°C immediately. Note: **PDE2A1** is very sensitive to freeze/thaw cycles. Do not re-use thawed aliquots or diluted enzyme.
- 6) Dilute **PDE2A1** in **PDE Assay Buffer** to 25 pg/μl (250 pg/reaction) in **PDE Assay Buffer***. Initiate reaction by adding 10 μl of diluted **PDE2A1** to the wells designated for the "Positive Control" and "Test Inhibitor". Discard any remaining diluted enzyme after use. *Note: optimal enzyme concentration may vary with the specific activity of the enzyme.

7) Incubate at room temperature for 1 hour.

| | Blank | Substrate Control | Positive Control | Test Inhibitor |
|---------------------------------|---------|----------------------|---------------------|-------------------|
| FAM-Cyclic-3',5'-AMP (200 nM) | _ | 12.5 µl | 12.5 µl | 12.5 µl |
| PDE assay buffer | 22.5 µl | 10 µl | _ | _ |
| Test Inhibitor | _ | _ | _ | 2.5 µl |
| Inhibitor Buffer (no inhibitor) | 2.5 µl | 2.5 µl | 2.5 µl | _ |
| PDE2A1 (25 pg/µl) | _ | _ | 10 µl | 10 µl |
| Total | 25 µl | 25 µl | 25 µl | 25 µl |

Step 2:

- 1) Shake the tube containing the **Binding Agent** to ensure it is thoroughly mixed. Mix **binding agent** thoroughly and dilute **binding agent** 1:100 with the **cAMP Binding Agent Diluent**.
- 2) Add 50 µl diluted **Binding Agent** to each well. Incubate at room temperature for 20 minutes with slow shaking.
- 3) Read the fluorescent polarization of the sample in a microtiter-plate reader capable of excitation at wavelengths ranging from 485 ± 5 nm and detection of emitted light ranging from 528 ± 10 nm. Blank value is subtracted from all other values.

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CALCULATING RESULTS:

Definition of Fluorescence Polarization:

$$P = \frac{I_{II} - I_{\perp}}{I_{II} + I_{\perp}}$$

where I_{\parallel} = Intensity with polarizers parallel and I_{\perp} = Intensity with polarizers perpendicular. Most instruments display fluorescence polarization in units of mP.

$$mP = \left(\frac{I_{II} - I_{\perp}}{I_{II} + I_{\perp}}\right) x \ 1000$$

The equation above assumes that light is transmitted equally well through both parallel and perpendicular oriented polarizers. In practice, this is generally not true and a correction must be made to measure the absolute polarization state of the molecule. This correction factor is called the "G Factor".

$$mP = \left(\frac{\mathbf{I}_{II} - G(\mathbf{I}_{\perp})}{\mathbf{I}_{II} + G(\mathbf{I}_{\perp})}\right) x \ 1000 \qquad \text{OR} \qquad mP = \left(\frac{G(\mathbf{I}_{II}) - \mathbf{I}_{\perp}}{G(\mathbf{I}_{II}) + \mathbf{I}_{\perp}}\right) x \ 1000$$

The G-factor is instrument-dependent and may vary slightly depending upon instrument and conditions. Please check the manual of your instrument to obtain the information about the establishment of the G-factor.

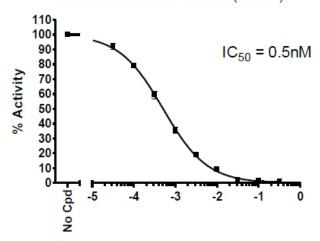


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EXAMPLE OF ASSAY RESULTS:

PDE2A1 Activity

Substrate Conc. = 100nM (cAMP)



Inhibition of PDE2A1 by Bay60-7550, measured using the *PDE2A1 Assay Kit*, BPS Bioscience # 60321. Fluorescence polarization was measured at 528 nm using a Tecan M-1000 fluorescent microplate reader. *Data shown is lot-specific. For lot-specific information, please contact BPS Bioscience, Inc. at info@bpsbioscience.com.*



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RELATED PRODUCTS:

| RELATED FRODUCTS. | | | | | | |
|-------------------|--|--|--|--|--|--|
| Catalog # | <u>Size</u> | | | | | |
| 60020 | 5 µg | | | | | |
| 60022 | 5 μg | | | | | |
| 60010 | 10 µg | | | | | |
| 60011 | 10 µg | | | | | |
| 60030 | 10 µg | | | | | |
| 60050 | 10 µg | | | | | |
| 60320 | 96 rxns. | | | | | |
| 60300 | 96 rxns. | | | | | |
| 60310 | 96 rxns. | | | | | |
| 60311 | 96 rxns. | | | | | |
| 60330 | 96 rxns. | | | | | |
| 60340 | 96 rxns. | | | | | |
| 60350 | 96 rxns. | | | | | |
| 60400 | 96 rxns. | | | | | |
| 60200 | 100 nmole | | | | | |
| 27215 | 5 mg | | | | | |
| | 60020 60022 60010 60011 60030 60050 60320 60310 60311 60330 60340 60350 60400 60200 | | | | | |