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Data sheet
IL-6:IL-6R Inhibitor Screening Assay Kit
Catalog #78027
Size: 96 reactions

DESCRIPTION: Interleukin 6 (IL-6), and its receptor complex interleukin 6 receptor (IL-6R, CD126)-gp130 (CD130), play an important role in immune response. Elevated IL-6 leads to an acute severe systemic inflammatory response known as 'cytokine storm', where an uncontrolled and excessive release of pro-inflammatory cytokines can cause multisystem organ failure and death. Mortality in COVID-19 patients has been linked to this severe response induced by the virus. Tocilizumab is a humanized monoclonal antibody against the IL-6R used as immunosuppressive drug, and it is emerging as an alternative treatment for COVID-19 patients at risk for 'cytokine storm'.

The IL-6:IL-6R Inhibitor Screening Assay Kit is designed for screening and profiling inhibitors of this receptor-ligand interaction. This kit comes in a convenient 96-well format, with purified IL-6R and IL-6-Biotin proteins, Streptavidin-HRP, and assay buffer for 100 binding reactions. The key to this kit is the high sensitivity of detection of IL-6-Biotin protein by Streptavidin-HRP. Only a few simple steps on a microtiter plate are required for the assay. First, IL-6R protein is attached to a nickel-coated 96-well plate. Next, IL-6-Biotin incubated with IL-6R on the plate. Finally, the plate is treated with Streptavidin-HRP followed by addition of an HRP substrate to produce chemiluminescence, which then can be measured using a chemiluminescence reader.

COMPONENTS:

Catalog #	Component	Amount	Storage	
	Human IL-6R alpha, Fc Tag (IL-6R)	10 µg	-80°C	Avoid multiple freeze/thaw cycles!
	Biotinylated Human IL-6 (IL-6-Biotin)	2 µg	-80°C	
79311	3x Immuno Buffer 1	50 ml	-20°C	
79728	Blocking Buffer 2	50 ml	+4°C	
79742	Streptavidin-HRP	15 µl	+4°C	
79670	ELISA ECL substrate A (transparent bottle)	6 ml	Room temp	
	ELISA ECL substrate B (brown bottle)	6 ml	Room temp	
79699	96-well white microplate	1	+4°C	

APPLICATIONS: This kit is useful for screening of blockers of the interaction between IL-6 and IL-6R.

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STABILITY: Up to 6 months from date of receipt, when stored as recommended.

REFERENCES:

Tanaka, T. *et al.* 2016. *Future Medicine* **8(8)**: 959-970
Zhang, C. *et al.* 2020. *Int.J. Antimicrob. Agents* **55(5)**: 105954.

MATERIALS OR INSTRUMENTS REQUIRED BUT NOT SUPPLIED:

PBS (Phosphate buffered saline)
Luminometer or fluorescent microplate reader capable of reading chemiluminescence
Adjustable micropipettor and sterile tips

ASSAY PROTOCOL:

All samples and controls should be tested in duplicate.

Coating the plate with IL-6R:

- 1) Thaw **IL-6R** on ice. Upon first thaw, briefly spin tube containing **IL-6R** to recover the full contents of the tube. Aliquot into single use aliquots. Immediately store remaining **IL-6R** in aliquots at -80°C. Note: **IL-6** is very sensitive to freeze/thaw cycles. Avoid multiple freeze/thaw cycles.
- 2) Dilute **IL-6R** to 2 µg/ml in PBS.
- 3) Add 50 µl of diluted **IL-6R** solution to each well and incubate overnight at 4°C.
- 4) Dilute **3x Immuno Buffer 1** to **1x Immuno Buffer 1** with water. Dilute only the amount required for the assay; store remaining **3x Immuno Buffer 1** undiluted.
- 5) Decant to remove supernatant. Wash the plate three times with 100 µl **1x Immuno Buffer 1**. Tap plate onto clean paper towels to remove liquid.

Block wells by adding 100 µl of **Blocking Buffer 2** to each well. Incubate for 10 minutes at room temperature with slow shaking. Remove supernatant as described in step 5.

Step 1:

- 1) Add 20 µl of **1x Immuno Buffer 1** to each well.
- 2) Add 10 µl of inhibitor solution to each well designated "Test Inhibitor". For the "Positive Control" and "Blank", add 10 µl of the same solution without inhibitor (inhibitor buffer). Optionally, incubate at room temperature for one hour with slow shaking.

Note: It is recommended to use PBS to dilute antibodies or other proteins acting as neutralization inhibitors. When testing small molecules dissolved in DMSO, the final DMSO concentration in the assay should be ≤1%. Inhibitor buffer should contain the same concentration of DMSO as the test inhibitor.

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- 3) Thaw **IL-6-Biotin** on ice. Upon first thaw, briefly spin tube containing enzyme to recover full contents of the tube. Aliquot **IL-6-Biotin** into single use aliquots. Immediately store remaining undiluted enzyme in aliquots at -80°C. Note: **IL-6-Biotin** is very sensitive to freeze/thaw cycles. Do not re-use thawed aliquots or diluted enzyme.
- 4) Dilute **IL-6-Biotin** to 0.5 ng/μl (approximately 25 nM) in **1x Immuno Buffer 1**. Keep diluted protein on ice until use. Discard any unused diluted protein after use.
- 5) Add another 20 μl of **1x Immuno Buffer 1** to the wells designated "Blank".

	Blank	Positive Control	Test Inhibitor
1x Immuno Buffer 1	40 μl	20 μl	20 μl
Test Inhibitor	-	-	10 μl
Inhibitor buffer (no inhibitor)	10 μl	10 μl	-
IL-6-Biotin (0.5 ng/μl)	-	20 μl	20 μl
Total	50 μl	50 μl	50 μl

- 6) Initiate reaction by adding 20 μl of diluted **IL-6-Biotin** (see Step 1-4) to wells labeled "Positive Control" and "Test Inhibitor". Incubate at room temperature for one hour with slow shaking.
- 7) Decant to remove supernatant. Wash the plate 3 times with 100 μl/well **1x Immuno Buffer 1**. Tap plate onto clean paper towels to remove liquid.
- 8) Block wells by adding 100 μl of **Blocking Buffer 2** to each well. Incubate for 10 minutes at room temperature. Remove supernatant as in Step 1-7.

Step 2:

- 1) Dilute **Streptavidin-HRP** 1000-fold with **Blocking Buffer 2**.
- 2) Add 100 μl to each well. Incubate for 1 hour at room temperature with slow shaking.
- 3) Wash plate three times with **1x Immuno Buffer 1**. Tap plate onto clean paper towel to remove liquid.
- 4) Block wells by adding 100 μl of **Blocking Buffer 2** to each well. Incubate for 10 minutes at room temperature. Decant to remove supernatant. Tap plate onto clean paper towels to remove liquid.
- 5) Just before use, mix 50 μl **ELISA ECL Substrate A** and 50 μl **ELISA ECL Substrate B**, then add 100 μl to each well. Discard any unused chemiluminescent reagent after use.
- 6) Immediately read sample in a luminometer or microtiter-plate capable of reading chemiluminescence. "Blank" value is subtracted from all readings.

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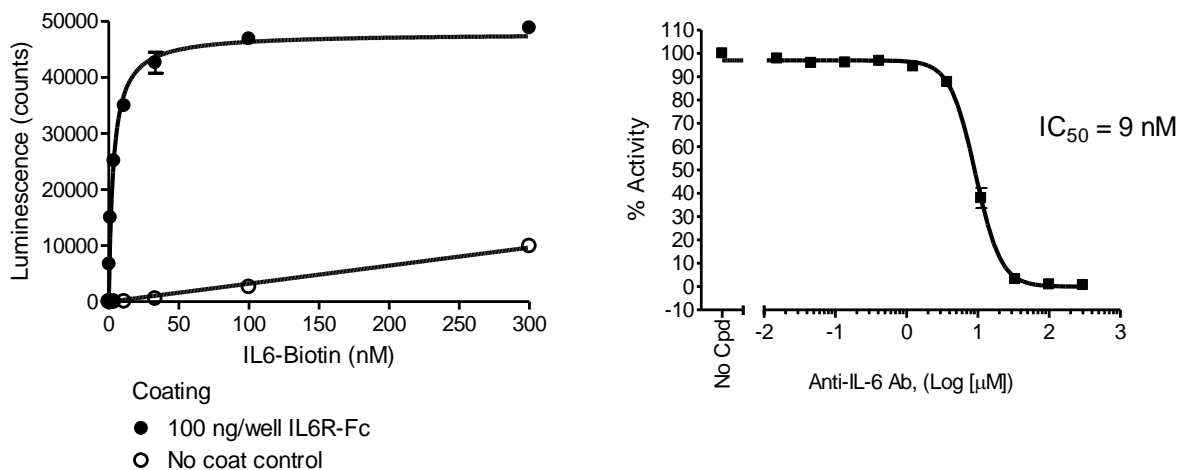
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Reading Chemiluminescence:

Chemiluminescence is the emission of light (luminescence) which results from a chemical reaction. The detection of chemiluminescence requires no wavelength selection because the method used is emission photometry and is not emission spectrophotometry.

To properly read chemiluminescence, make sure the plate reader is set for LUMINESCENCE mode. Typical integration time is 1 second; delay after plate movement is 100 milliseconds. Do not use a filter when measuring light emission. Typical settings for the Synergy 2 BioTek plate reader are: use the "hole" position on the filter wheel; Optics position: Top; Read type: endpoint. Sensitivity may be adjusted based on the luminescence of a control assay without enzyme (typically we set this value as 100).

Example of assay results:



Binding of IL-6-Biotin to immobilized IL-6R (left) and inhibition of IL-6:IL-6R binding using the Human IL-6 Antibody (R&D Systems, MAB2061) (right) in the IL-6:IL-6R Inhibitor Screening Assay Kit (BPS Bioscience, #78027). Luminescence was measured using a BioTek microplate reader. Data shown is lot-specific. For lot-specific information, please contact BPS Bioscience, Inc. at info@bpsbioscience.com.

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RELATED PRODUCTS:

<u>Product Name</u>	<u>Catalog#</u>	<u>Size</u>
IL-6 (Human) Colorimetric ELISA Detection Kit	79820	96 reactions
Human Interleukin-6	90196	20 µg
Immuno Buffer 1	79311	50 ml
Blocking Buffer 2	79728	50 ml
ELISA ECL Substrate	79760-1	200 ml

TROUBLESHOOTING GUIDE

Problem	Possible cause	Solution
Luminescence signal of positive control reaction is weak	IL-6-Biotin or IL-6R have lost activity	Proteins lose activity upon repeated freeze/thaw cycles. Use fresh IL-6R and fresh IL-6-Biotin. Store proteins in single-use aliquots. Increase time of enzyme incubation. Increase enzyme concentration.
	Incorrect settings on instruments	Refer to instrument instructions for settings to increase sensitivity of light detection.
	Chemiluminescent reagents mixed too soon	Chemiluminescent solution should be used within 15 minutes of mixing. Ensure both reagents are properly mixed.
Luminescent signal is erratic or varies widely among wells	Inaccurate pipetting/technique	Run duplicates of all reactions. Use a multichannel pipettor. Use master mixes to minimize errors.
	Bubbles in wells	Pipette slowly to avoid bubble formation. Tap plate lightly to disperse bubbles; be careful not to splash between wells.
Background (signal to noise ratio) is high	Insufficient washes	Increase number of washes. Increase wash volume.
	Sample solvent is inhibiting the enzyme	Run negative control assay including solvent. Maintain DMSO level at <1%. Increase time of enzyme incubation.
	Results are outside the linear range of the assay	Use different concentrations of IL-6-Biotin to create a standard curve

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