



# SZABO SCANDIC

Part of Europa Biosite

## Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

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### Lieferung & Zahlungsart

siehe unsere [Liefer- und Versandbedingungen](#)

### Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

### SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

[mail@szabo-scandic.com](mailto:mail@szabo-scandic.com)

[www.szabo-scandic.com](http://www.szabo-scandic.com)

[linkedin.com/company/szaboscandic](https://www.linkedin.com/company/szaboscandic) 

**Description**

This cell line is a double knockout of TCR (T Cell Receptor) and B2M (Beta-2-Microglobulin). First, the TRAC (T-Cell Receptor Alpha Constant) and the TRBC1 (T-Cell Receptor Beta Constant 1) domains of the TCR $\alpha/\beta$  chains were genetically removed by CRISPR/Cas9 genome editing from Jurkat cells to generate the TCR Knockout Jurkat cell line (BPS Bioscience #78539). These TCR knockout cells were then used to generate a new cell line in which B2M was also genetically removed by CRISPR/Cas9 genome editing.

**Background**

The T Cell Receptor (TCR) is found on the surface of T-cells and is responsible for recognizing antigens bound to MHC (Major Histocompatibility Complex) molecules. Stimulation of the TCR triggers a signaling cascade that leads to the activation of transcription factors involved in the upregulation and secretion of cytokines, T cell proliferation, and cell differentiation into effector and memory cells. TCR-activated transcription factors include AP-1 (Activator Protein 1), NF- $\kappa$ B (Nuclear Factor Kappa-light-chain-enhancer of activated B cells) and NFAT (Nuclear Factor of Activated T-cells).

Beta-2-Microglobulin is a required component of Major Histocompatibility Complex (MHC) class I molecules, which present peptide fragments from within the cell to cytotoxic T cells as part of the adaptive immune system. B2M plays an essential role both in governing MHC class I molecule stability and in promoting antigen binding by presenting the antigen to CD3/TCR complex of CD8+ T cells.

Knockout of both TCR and B2M will support the manufacture of universal CAR-T cells. Knockout of TCR or B2M prevents the elimination of allogeneic T cells that express foreign HLA-I molecules, thereby enabling the generation of CAR-T cells from allogeneic healthy donors T cells in vivo.

**Application(s)**

Useful for the development of improved universal CAR-T or other effector cells.

**Materials Provided**

Components	Format
2 vials of frozen cells	Each vial contains $2 \times 10^6$ cells in 1 ml of cell freezing medium (BPS Bioscience #79796)

**Parental Cell Line**

Jurkat (clone E6-1), human T lymphoblast, suspension

**Mycoplasma Testing**

The cell line has been screened to confirm the absence of Mycoplasma species.

**Materials Required but Not Supplied**

These materials are not supplied with the cell line but are necessary for cell culture and cellular assays. BPS Bioscience's reagents are validated and optimized for use with this cell line and are highly recommended for best results. Media components are provided in the Media Formulations section below.

**Media Required for Cell Culture**

Name	Ordering Information
Thaw Medium 2	<a href="#">BPS Bioscience #60184</a>

**Storage Conditions**

Cells are shipped in dry ice and should immediately be thawed or stored in liquid nitrogen upon receipt. Do not use a -80°C freezer for long term storage. Contact technical support at [support@bpsbioscience.com](mailto:support@bpsbioscience.com) if the cells are not frozen in dry ice upon arrival.

**Media Formulations**

For best results, it is *highly recommended* to use these validated and optimized media from BPS Bioscience. Other preparations or formulations of media may result in suboptimal performance.



Note: Thaw Media do *not* contain selective antibiotics. Cells should be grown at 37°C with 5% CO<sub>2</sub>. BPS Bioscience's cell lines are stable for at least 15 passages when grown under proper conditions.

*Media Required for Cell Culture*

*Thaw Medium 2 (BPS Bioscience #60184):*

RPMI 1640 medium, supplemented with 10% FBS, 1% Penicillin/Streptomycin

**Cell Culture Protocol***Cell Thawing*

1. Swirl the vial of frozen cells for approximately 60 seconds in a 37°C water bath. As soon as the cells are thawed (it may be slightly faster or slower than 60 seconds), quickly transfer the entire contents of the vial to a tube containing 10 ml of pre-warmed Thaw Medium 2.  
**Leaving the cells in the water bath at 37°C for too long will result in rapid loss of viability.**
2. Immediately spin down the cells at 300 x g for 5 minutes, remove the medium and resuspend the cells in 5 ml of pre-warmed Thaw Medium 2.
3. Transfer the resuspended cells to a T25 flask and incubate at 37°C in a 5% CO<sub>2</sub> incubator.
4. After 24 hours of culture, check for cell viability. For a T25 flask, add 3-4 ml of Thaw Medium 2, and continue growing in a 5% CO<sub>2</sub> incubator at 37°C until the cells are ready to passage.
5. Cells should be passaged before they reach a density of 2 x 10<sup>6</sup> cells/ml. At first passage and subsequent passages, use Thaw Medium 2.

*Cell Passage*

Dilute the cell suspension into new culture vessels before they reach a density of 2 x 10<sup>6</sup> cells/ml, at no less than 0.2 x 10<sup>6</sup> cells/ml of Thaw Medium 2. The sub-cultivation ratio should maintain the cells between 0.2 x 10<sup>6</sup> cells/ml and 2 x 10<sup>6</sup> cells/ml.

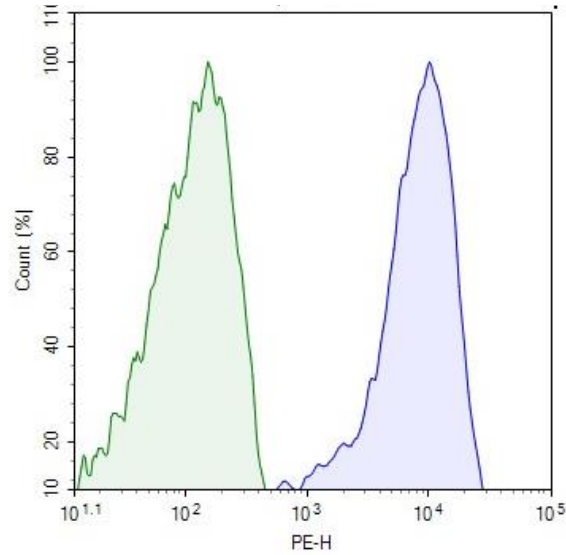
*Cell Freezing*

1. Spin down the cells at 300 x g for 5 minutes, remove the medium and resuspend the cell pellet in 4°C Freezing Medium (BPS Bioscience #79796, or 10% DMSO + 90% FBS) at a density of ~2 x 10<sup>6</sup> cells/ml.
2. Dispense 1 ml of cell aliquots into cryogenic vials. Place the vials in an insulated container for slow cooling and store at -80°C overnight.
3. Transfer the vials to liquid nitrogen the next day for storage.



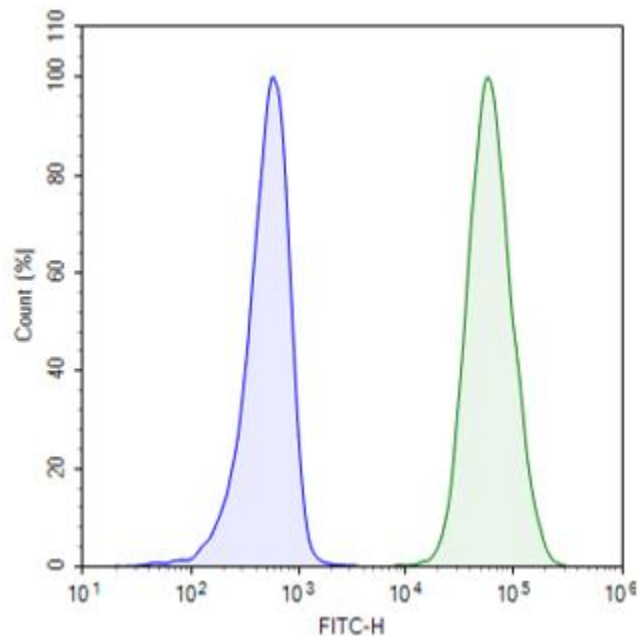
Note: It is recommended to expand the cells and freeze at least 10 vials at an early passage for future use.

### A. Validation Data



*Figure 1. Expression of TCR in the TCR/B2M Knockout Jurkat cells.*

TCR/B2M Knockout Jurkat cells (green) or parental Jurkat cells (blue) were stained with PE-labeled anti-human TCR  $\alpha/\beta$  antibody (BioLegend #306707) and analyzed by flow cytometry. Y-axis is the % cell number. X-axis is the intensity of PE.



*Figure 2. Expression of B2M in the TCR/B2M Knockout Jurkat cells.*

TCR/B2M Knockout Jurkat cells (blue) or parental Jurkat cells (green) were stained with CoraLite 488-labeled anti-human HLA-ABC antibody (BD Pharmingen #560169) and analyzed by flow cytometry. Y-axis is the % cell number. X-axis is the intensity of FITC.

## Sequences

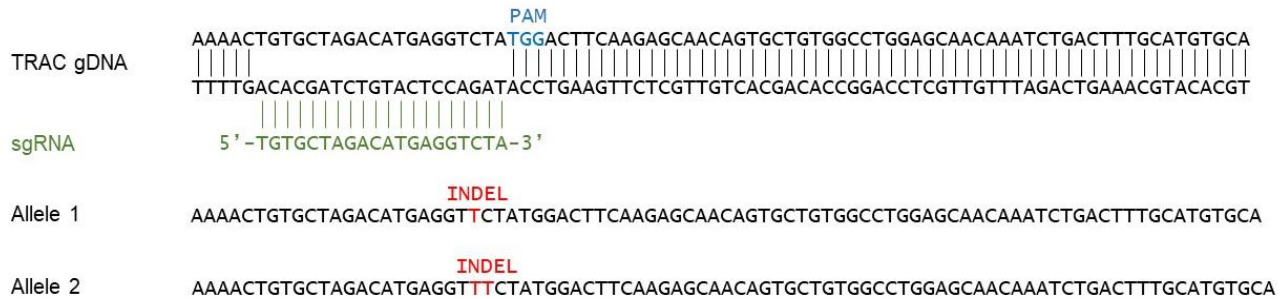


Figure 3. Genomic Sequencing of TRAC in the TCR/B2M Knockout Jurkat Cell Line.

Genomic DNA from the TCR/B2M Knockout Jurkat Cells was isolated and sequenced. The PAM (Protospacer Adjacent Motif) is shown in blue, the sgRNA (synthetic guide RNA) in green, and the Indels (Insertions/Deletions) in the two TRAC alleles are indicated in red.

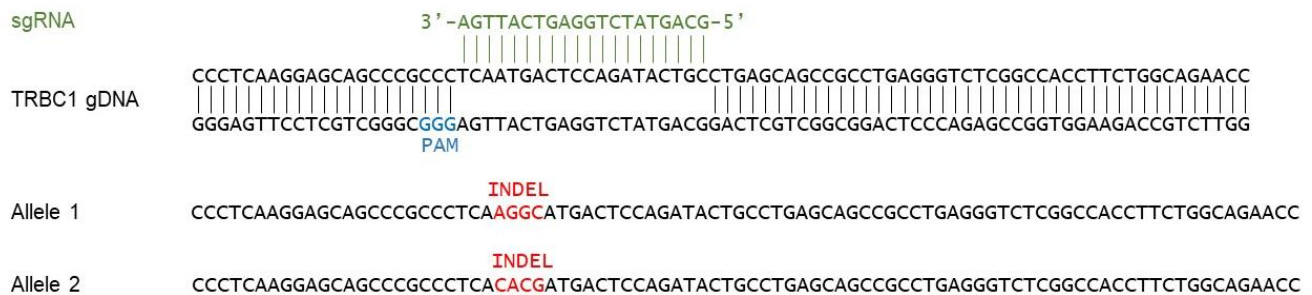


Figure 4. Genomic Sequencing of TRBC1 in the TCR/B2M Knockout Jurkat Cell Line.

Genomic DNA from the TCR/B2M Knockout Jurkat Cells was isolated and sequenced. The PAM (Protospacer Adjacent Motif) is shown in blue, the sgRNA (synthetic guide RNA) in green, and the Indels (Insertions/Deletions) in the two TRBC1 alleles are indicated in red.

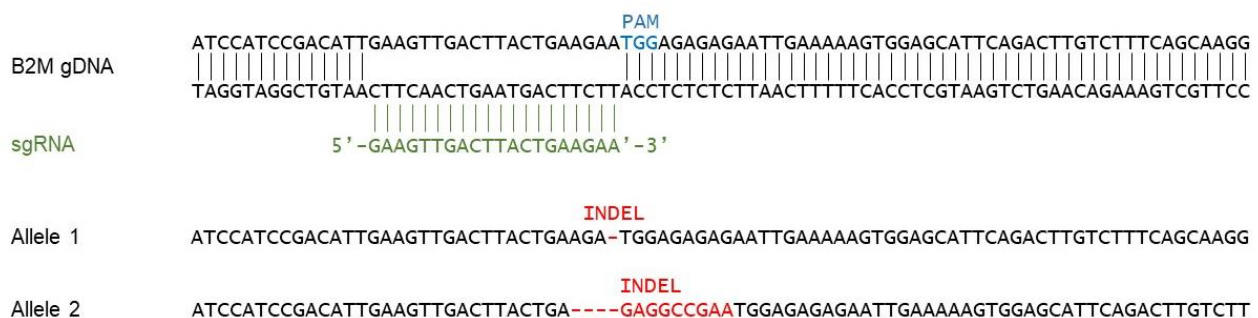


Figure 5. Genomic Sequencing of B2M in the TCR/B2M Knockout Jurkat Cell Line.

Genomic DNA from the TCR/B2M Knockout Jurkat Cells was isolated and sequenced. The PAM (Protospacer Adjacent Motif) is shown in blue, the sgRNA (synthetic guide RNA) in green, and the Indels (Insertions/Deletions) in the two B2M alleles are indicated in red.

