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SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

mail@szabo-scandic.com

www.szabo-scandic.com

[linkedin.com/company/szaboscandic](https://www.linkedin.com/company/szaboscandic) 



6042 Cornerstone Court W, Ste B
San Diego, CA 92121
Tel: 1.858.202.1401
Fax: 1.858.481.8694
Email: info@bpsbioscience.com

Data Sheet

IL-2 Promoter Luciferase Reporter Lentivirus

Catalog #: 79825

Product Description

The IL-2 Promoter Luciferase Reporter Lentivirus are replication incompetent, HIV-based, VSV-G pseudotyped lentiviral particles that are ready to be transduced into almost all types of mammalian cells, including primary and non-dividing cells. The particles contain a firefly luciferase gene driven by the human IL-2 promoter (Figure 1). After transduction, activation of the IL-2 signaling pathway in the target cells can be monitored by measuring the luciferase activity.

Application

- Screen for activators or inhibitors of IL-2 signaling pathway in transduced target cells
- Generation of IL-2 promoter luciferase reporter stable cell line

Formulation

The lentiviruses were produced from HEK293T cells in medium containing 90% DMEM + 10% FBS.

Titer

Two vials (500 μ l x 2) of IL-2 promoter luciferase reporter lentivirus at a titer $>5 \times 10^6$ TU/ml. The titer will vary with each lot; the exact value is provided with each shipment.

Storage

Lentiviruses are shipped with dry ice. For long term storage, it is recommended to store the virus at -80°C . Avoid repeated freeze-thaw cycles. Titers can drop significantly with each freeze-thaw cycle.

Biosafety

The lentiviruses are produced with the third generation SIN (self-inactivation) lentivector which ensures self-inactivation of the lentiviral construct after transduction and integration into the genomic DNA of the target cells. None of the HIV genes (gag, pol, rev) will be expressed in the transduced cells, as they are expressed from packaging plasmids lacking the packing signal.

Although the pseudotyped lentiviruses are replication-incompetent, they require the use of a Biosafety Level 2 facility. BPS recommends following all local federal, state, and institutional regulations and using all appropriate safety precautions.

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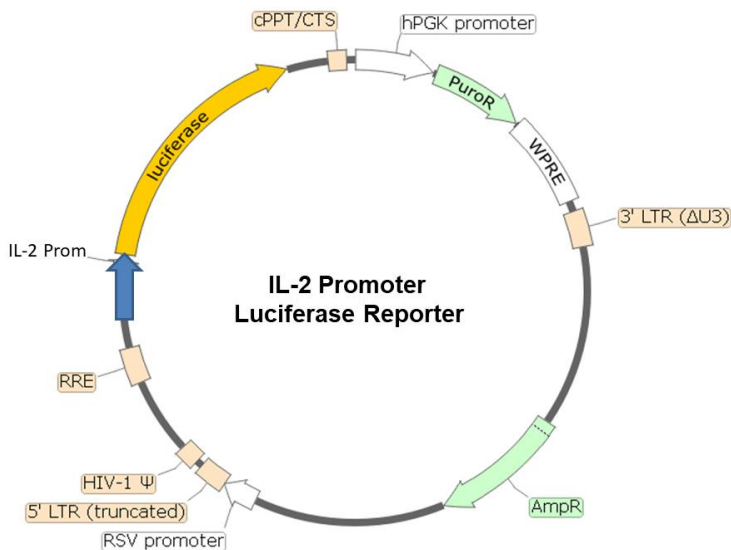


Figure 1. Schematic of the lenti-vector used to generate the IL-2 promoter luciferase reporter lentivirus

Materials Required but Not Supplied

- Jurkat cells (ATCC #TIB-152)
- Anti-CD3 agonist antibody (BPS Bioscience, #71274)
- Anti-CD28 agonist antibody (BPS Bioscience, #100182)
- Jurkat growth medium (Thaw Medium 2, BPS Bioscience, #60184)
- Polybrene (Millipore, #TR-1003-G)
- 96-well tissue culture treated white clear-bottom assay plate (Corning, #3610)
- One-Step luciferase assay system (BPS Bioscience, #60690)
- Luminometer

Assay protocol

The following protocol is a general guideline for transducing Jurkat cells using IL-2 promoter luciferase reporter lentivirus. The optimal transduction conditions (e.g. MOI, concentration of polybrene, time of assay development) should be optimized according to the cell type and the assay requirements. In most cell types, the expression of the reporter gene can be measured approximately 72 hours after transduction. For cell types with low transduction efficacy, it may be necessary to select the cells stably expressing the reporter gene with puromycin prior to carrying out the reporter assays.

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1. Precoat the 96 well plate with anti-CD3 antibody (100 μ l at 10 μ g/ml) in PBS overnight. Leave a few uncoated wells to serve as negative controls. Rinse all wells 3x with Jurkat growth medium.
2. Harvest the Jurkat cells by centrifugation and resuspend the cells in fresh growth medium. Dilute the cells to 1×10^6 /ml in growth medium. Mix 500 μ l of the Jurkat cells and 100 μ l of IL-2 promoter luciferase reporter lentivirus in a 1.5-ml Eppendorf tube. Add polybrene to a final concentration of 8 μ g/ml. Gently mix and incubate the virus with the Jurkat cells for 20 min at room temperature in the tissue culture hood.
3. Centrifuge the virus/cells mixture for 30 minutes at 800 x g at 32°C. Remove the virus containing medium and resuspend the cell pellet in 2 ml of fresh Jurkat growth medium. Transfer the cells into one well in a 6-well plate. Incubate the plate at 37°C with 5% CO₂ for 48-66 hours. The transduced Jurkat cells are ready for assay development.
4. Harvest the cells and resuspend the cells into 1 ml of fresh Jurkat growth medium. Add 90 μ l of the cells to each well of the anti-CD3 antibody-coated 96-well plate. Dilute anti-CD28 agonist antibody using Jurkat growth medium, and add 10 μ l of diluted anti-CD28 antibody to the anti-CD3 antibody stimulated wells.
5. Incubate at 37°C with 5% CO₂ overnight.
6. Prepare the ONE-Step™ Luciferase reagent per recommended protocol. Add 100 μ l of ONE-Step™ Luciferase Assay reagent per well. Incubate at room temperature for ~15 to 30 minutes and measure luminescence using a luminometer.

Important Notes:

1. To generate the IL-2 promoter luciferase reporter stable cell line, remove the growth medium 48 hours after transduction and replace it with fresh growth medium containing the appropriate amount of puromycin for antibiotic selection of transduced cells.
2. The following Lenti Reporter Controls are also available from BPS Bioscience to meet your experimental needs:
 - 1) Reporter Negative Control Lentivirus (BPS Bioscience, #79578): Ready-to-transduce lentiviral particles expressing firefly luciferase under the control of a minimal promoter. The negative control is important to establish the specificity of any treatments and to determine the background reporter activity.
 - 2) Renilla Luciferase (Rluc) Lentivirus (BPS Bioscience, #79565): Ready-to-transduce lentiviral particles expressing Renilla luciferase under the CMV promoter. The RLuc

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lentivirus can serve as an internal control to overcome sample-to-sample variability when performing dual-luciferase reporter assays.

- 3) Firefly Luciferase (Fluc) Lentivirus (BPS Bioscience, #79692-G, #79692-H, #79692-P): Ready-to-transduce lentiviral particles expressing firefly luciferase under the CMV promoter. The Fluc lentivirus can serve as a positive control for transduction optimization studies.

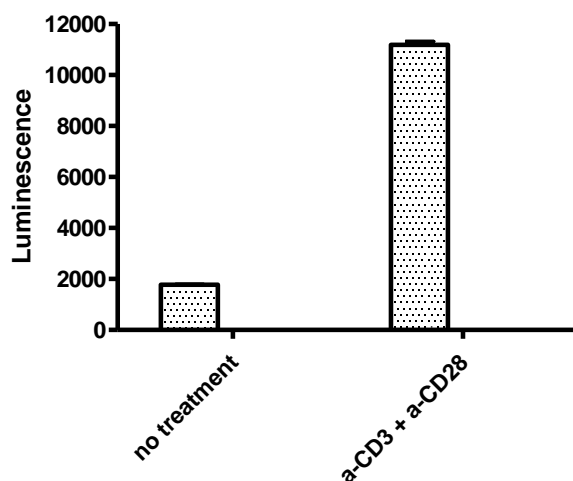


Figure 2. IL-2 promoter luciferase reporter activity stimulated by anti-CD3 and anti-CD28 agonist antibodies in Jurkat cells. Appropriate 50,000 Jurkat cells were transduced with 100,000 TU IL-2 promoter luciferase reporter lentivirus. After 48 hours of transduction, medium was changed to fresh Jurkat growth medium. Cells were stimulated with anti-CD3 agonist antibody (precoated on a 96-well plate) and anti-CD28 agonist antibody (2 µg/ml) for overnight. The noncoated wells were performed in parallel as controls. The results are shown as the raw luminescence reading.

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Related Products

<u>Product</u>	<u>Cat. #</u>	<u>Size</u>
NF- κ B Luciferase Reporter Lentivirus	79564	500 μ l x2
CRE/CREB Luciferase Reporter Lentivirus	79580	500 μ l x2
NFAT Luciferase Reporter Lentivirus	79579	500 μ l x2
STAT3 Luciferase Reporter Lentivirus	79744	500 μ l x2
STAT5 Luciferase Reporter Lentivirus	79745	500 μ l x2
TCF/LEF Luciferase Reporter Lentivirus	79787	500 μ l x2
ISRE Luciferase Reporter Lentivirus	79824	500 μ l x2
IL-8 Promoter Luciferase Reporter Lentivirus	79827	500 μ l x2
Reporter Negative Control Lentivirus	79578	500 μ l x2
Renilla Luciferase (Rluc) Lentivirus	79565	500 μ l x2
Firefly Luciferase (Fluc) Lentivirus (G418)	79692-G	500 μ l x2
Firefly Luciferase (Fluc) Lentivirus (Hygromycin)	79692-H	500 μ l x2
Firefly Luciferase (Fluc) Lentivirus (Puromycin)	79692-P	500 μ l x2
GFP Reporter Lentivirus	79703	1 ml x 2
IL-2 Luciferase Reporter Jurkat Cell Line	60481	2 vials
Anti-CD3 Agonist Antibody	71274-1	50 μ g
Anti-CD28 Agonist Antibody	100182	50 μ g
ONE-Step™ Luciferase Assay System	60690-1	10 ml
ONE-Step™ Luciferase Assay System	60690-2	100 ml
Dual Luciferase (Firefly-Renilla) Assay System	60683	10 ml

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