

# Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

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# Lieferung & Zahlungsart

siehe unsere Liefer- und Versandbedingungen

## Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

### SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

mail@szabo-scandic.com

www.szabo-scandic.com

linkedin.com/company/szaboscandic in



### Description

The Chemi-Verse™ CDK2/CyclinA2 Kinase Assay Kit is designed to measure CDK2 (cyclin dependent kinase 2)/CyclinA2 kinase activity for screening and profiling applications using ADP-Glo™ as a detection reagent. The assay kit comes in a convenient 96-well format, with enough purified recombinant CDK2/CyclinA2 (amino acids 174-432) complex, kinase substrate, ATP, and kinase assay buffer for 100 enzyme reactions.

#### **Background**

CDK2 (cyclin dependent kinase 2), also known as cell division protein kinase 2, is a member of the serine/threonine cyclin-dependent protein kinase family, and it is involved in cell cycle. CDK2 is regulated by phosphorylation and can associate with either cyclin E during G1 phase and cyclin A during S phase. Its association with cyclins induces a conformational change that results in a dramatic increase of the kinase activity. Cyclin levels vary during the cell cycle, which allow cyclins to regulate CDK activity in the cell. Dissociation of the complex returns CDK to its basal activity, and CDK is degraded by ubiquitin-mediated proteolysis. CDK2 can phosphorylate several proteins, being part of DNA damage, protein degradation, signal transduction, and other crucial cellular pathways. Lack of regulation in cell cycle can result in cancer. In general, CDK2 is not itself upregulated or hyperactive in cancer, and its abnormal activity comes from dysregulation in its binding partners. The development of inhibitors specific for CDK2 has been difficult, as CDKs have similar active sites, and for instance inhibition of CDK1 can be highly detrimental. The understanding of the mechanisms involved in cell cycle regulation, and its control via the use of small molecule inhibitors alone or in combination therapy will open new therapeutic avenues for the treatment of cancer and neurodegenerative diseases.

#### **Applications**

Study enzyme kinetics and screen small molecule inhibitors for drug discovery and high throughput screening (HTS) applications.

#### **Supplied Materials**

Catalog #	Name	Amount	Storage
40101	CDK2/CyclinA2, GST-Tag*	1 μg	-80°C
79334	5x Kinase Buffer 1	1.5 ml	-20°C
79686	500 μM ATP	50 μΙ	-20°C
	10x CDK Substrate Peptide 1	50 μΙ	-20°C
79696	White 96-well plate	1	Room Temperature

<sup>\*</sup>The concentration of the protein is lot-specific and will be indicated on the tube.

#### **Materials Required but Not Supplied**

Name	Ordering Information
ADP-Glo™ Kinase Assay	Promega #V6930
DTT (Dithiothreitol), 1M, optional	
Microplate reader capable of reading luminescence	
Adjustable micropipettor and sterile tips	
30°C incubator	



#### **Storage Conditions**



This assay kit will perform optimally for up to **6 months** from date of receipt when the materials are stored as directed.

### Safety



This product is for research purposes only and not for human or therapeutic use. This product should be considered hazardous and is harmful by inhalation, in contact with skin, eyes, clothing, and if swallowed. If contact occurs, wash thoroughly.

#### **Assay Principle**

The ADP-Glo™ Kinase Assay (Promega #V6930) quantifies the amount of ADP produced by a kinase upon phosphorylation of a substrate. First, addition of the ADP-Glo™ reagent terminates the reaction and quenches the remaining ATP. Second, the addition of the Kinase Detection reagent converts the produced ADP to ATP. The newly generated ATP is quantified by a luciferase reaction. The luminescent signal correlates with the amount of ADP generated by the kinase and is linear to 1 mM ATP.

#### **Contraindications**

The final concentration of DMSO in the assay should not exceed 1%.

#### **Assay Protocol**

- All samples and controls should be tested in duplicate.
- The assay should include "Blank", "Positive Control" and "Test Inhibitor" conditions.
- We recommend maintaining the diluted protein on ice during use.
- For detailed information on protein handling please refer to Protein FAQs (bpsbioscience.com).
- We recommend using Dinaciclib as internal control. If not running a dose response curve for the control
  inhibitor, we recommend running the control inhibitor at 0.1X, 1X and 10X the IC<sub>50</sub> value shown in the
  validation data below.
- 1. Thaw 5x Kinase Assay Buffer 1, 500 µM ATP, and 10x CDK Substrate Peptide 1.

Optional: If desired, make 5x Kinase Assay Buffer 1 with 10 mM DTT.

2. Prepare 3 ml of **1x Kinase Assay Buffer 1** by mixing 600 μl of **5x Kinase Assay Buffer 1** with 2,400 μl of distilled water.

Note: Three (3 ml) of 1x Kinase Assay Buffer 1 is sufficient for 100 reactions.

- 3. Prepare a **Master Mix** (12.5  $\mu$ l/well): N wells x (6  $\mu$ l of 5x Kinase Assay Buffer 1 + 0.5  $\mu$ l of 500  $\mu$ M ATP + 0.5  $\mu$ l of 10x CDK Substrate Peptide 1 + 5.5  $\mu$ l of distilled water).
- 4. Add 12.5 μl of Master Mix to every well.
- 5. Prepare the **Test Inhibitor** (2.5  $\mu$ l/well): for a titration prepare serial dilutions at concentrations 10-fold higher than the desired final concentrations. The final volume of the reaction is 25  $\mu$ l.



5.1 If the Test Inhibitor is water-soluble: Prepare serial dilutions in 1x Kinase Assay Buffer 1, 10-fold more concentrated than the desired final concentrations.

For the positive and negative controls, use 1x Kinase Assay Buffer 1 (Diluent Solution).

#### OR

5.2 If the Test inhibitor is soluble in DMSO: Prepare the test inhibitor at 100-fold the highest desired concentration in 100% DMSO, then dilute the inhibitor 10-fold in 1x Kinase Assay Buffer 1 to prepare the highest concentration of the 10-fold intermediate dilutions. The concentration of DMSO is now 10%.

Prepare serial dilutions of the Test Inhibitor at 10-fold the desired final concentrations using 10% DMSO in 1x Kinase Assay Buffer 1 to keep the concentration of DMSO constant.

For positive and negative controls, prepare 10% DMSO in 1x Kinase Assay Buffer 1 (vol/vol) so that all wells contain the same amount of DMSO (Diluent Solution).

Note: The final concentration of DMSO should not exceed 1%.

- 6. Add 2.5 μl of Test Inhibitor to each well labeled "Test Inhibitor".
- 7. Add 2.5  $\mu$ l of Diluent Solution to the "Positive Control" and "Blank" wells.
- 8. Add 10 μl of 1x Kinase Assay Buffer 1 to the "Blank" wells.
- 9. Thaw CDK2/CyclinA2 Kinase on ice. Briefly spin the tube to recover its full content.
- 10. Dilute the protein kinase (10  $\mu$ l/well) to 1 ng/ $\mu$ l with 1x Kinase Assay Buffer 1.
- 11. Initiate the reaction by adding 10  $\mu$ l of diluted kinase to the wells designated "Positive Control" and "Test Inhibitor".

Component	Blank	<b>Positive Control</b>	<b>Test Inhibitor</b>
Master Mix	12.5 μl	12.5 μΙ	12.5 μΙ
Test Inhibitor	-	-	2.5 μΙ
Diluent Solution	2.5 μΙ	2.5 μΙ	-
1x Kinase Assay Buffer 1	10 μΙ	-	-
Diluted CDK2/CyclinA2 (1 ng/μl)	-	10 μΙ	10 μΙ
Total	25 µl	25 μΙ	25 µl

- 12. Incubate at 30°C for 45 minutes.
- 13. Thaw the ADP-Glo™ reagent.
- 14. At the end of the 45-minute reaction, add 25 μl of ADP-Glo™ reagent to each well.



- 15. Cover the plate with aluminum foil and incubate at Room Temperature (RT) for 45 minutes.
- 16. Thaw the Kinase Detection Reagent.
- 17. Add 50 μl of Kinase Detection reagent to each well.
- 18. Cover the plate with aluminum foil and incubate at RT for another 45 minutes.
- 19. Immediately read in a luminometer or a microplate reader capable of reading luminescence.
- 20. The "Blank" value is subtracted from all other readings.

#### **Reading Luminescence**

Luminescence is the emission of light resulting from a chemical reaction. The detection of luminescence requires no wavelength selection because the method used is emission photometry and not emission spectrophotometry.

To properly read luminescence, make sure the plate reader is set for LUMINESCENCE mode. Typical integration time is 1 second, delay after plate movement is 100 msec. Do not use a filter when measuring light emission. Typical settings for the Synergy 2 BioTek plate reader: use the "hole" position on the filter wheel; Optics position: Top; Read type: endpoint. Sensitivity may be adjusted based on the luminescence of a control assay without enzyme (typically we set this value as 100).

#### **Example Results**

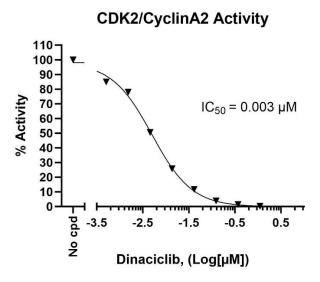


Figure 1: Inhibition of CDK2/CyclinA2 kinase activity by Dinaciclib. CDK2/CyclinA2 kinase activity was measured in the presence of increasing concentrations of Dinaciclib (SelleckChem S2768). The "Blank" value was subtracted from all other values. Results are expressed as the percent of control (kinase activity in the absence of inhibitor, set at 100%).

Data shown is representative. For lot-specific information, please contact BPS Bioscience, Inc. at support@bpsbioscience.com.



### **Troubleshooting Guide**

Visit bpsbioscience.com/assay-kits-faq for detailed troubleshooting instructions. For all further questions, please email support@bpsbioscience.com

#### References

Tadesse S., et al., 2020 Drug Discovery Today 25(2):406-413.

#### **Related Products**

Products	Catalog #	Size
Chemi-Verse™ CDK2/CyclinE1 Kinase Assay Kit	82228	96 reactions
CDK2/CyclinA2, GST-Tag Recombinant	40101	10 μg
CDK2 (no tag)/CyclinA2, His-GST-Tags Recombinant	41101	10 μg
Chemi-Verse™ CDK3/CyclinE1 Kinase Assay Kit	78884	96 reactions
CDK3/CyclinE1, GST-Tag Recombinant	40103	10 μg

Version 012524

