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Description

The HLA-A/B/C Knockout Electroporation Kit is suitable for cell line and primary T cell engineering via electroporation. The kit contains both the Cas9 enzyme (*Streptococcus pyogenes*) and the gRNA targeting HLA-A/B/C (Human Leukocyte Antigens). This kit is sufficient to engineer up to 5 million primary T cells.

Background

HLA (Human Leukocyte Antigens)-A, B, and C, are the three major types of MHC (major histocompatibility complex) class 1 transmembrane proteins. They form a heterodimer with the $\beta 2$ microglobulin protein (encoded by the B2M gene). The MHC class 1 molecules present short polypeptides, usually between 7-11 amino acids long, to the immune system for recognition as either “self” or “non-self”. HLA-C, for instance, is present in all cells and exists as several haplotypes due to the diversity of HLA-C genes. C*08:02 represents one such haplotype. HLA class I present neoantigen-derived peptides to the cell surface, allowing them to be recognized by T cells, via TCR (T cell receptors). Cancer immunotherapy has been taking advantage of that mechanism, by engineering T cells to express TCRs able to recognize specific cancer immunogens. In 2016 the use of HLA-C*08:02-restricted TIL (tumor infiltrating lymphocytes) specifically targeting KRAS (Kirsten rat sarcoma virus) G12D mutation in lung cancer resulted in positive results. A similar approach was pursued in a patient with metastatic pancreatic cancer and resulted in regression of the disease. The study of HLA-C*08:02-restricted TIL expressing TCR against other neoantigens may prove beneficial in cancer therapy. K562 cells are HLA class I and II negative, making them an ideal cellular model to introduce and study specific haplotype responses. HLA mismatching between donor cells and the individual can lead to immune rejection, and one option is the knockout the endogenous HLA, allowing cells to be more widely universally used.

Application

- Knockout of HLA-A/B/C in cells of interest

Supplied Materials

Catalog #	Name	Amount	Storage
	HLA-A/B/C gRNA	10 μ l	-20°C
	Cas9 enzyme (<i>Streptococcus pyogenes</i>)	10 μ l	-20°C

Storage Conditions

Components are shipped in dry ice and stored at -20°C for long term storage. The components maintain their stability and performance after 5 freeze-thaw cycles.

Materials Required but Not Supplied

These materials are not supplied with the kit but may be required for cell culture and cellular assays. BPS Bioscience’s reagents are validated and optimized for use with this kit and are highly recommended for the best results.

Name	Ordering Information
TCellIM™	BPS Bioscience #78753
Normal Human Peripheral Blood Mononuclear Cells, Frozen	BPS Bioscience #79059
Human Interleukin-2 Recombinant	BPS Bioscience #90184
EasySep™ Human CD4+ T Cell Isolation Kit	Stemcell technologies #17952
EasySep™ Human CD8+ T Cell Isolation Kit	Stemcell technologies #17953
Human CD3/CD28/CD2 T cell Activator	Stemcell technologies #10970
PE-labeled anti-human HLA-A,B,C Antibody	BioLegend #311405
Neon NxT Electroporation System (or other electroporation system)	NEON1SK

Media Required for T cell Culture

T Cell Medium: TCellIM™ (BPS Bioscience #78753) supplemented with 10 ng/ml Human Interleukin-2 Recombinant (BPS Bioscience #90184).

Assay Protocol

- The following protocol was used to knock out the HLA-A/B/C in CD4⁺ and CD8⁺ primary T cells using the HLA-A/B/C Knockout Electroporation kit, and it is a general guideline only.
- The assay conditions (concentration of gRNAs and Cas9 enzyme, and time of assay) should be optimized according to the cell type, donor, and the assay requirements.

Day 0:

1. Isolate CD4⁺ T cells and CD8⁺ T cells from previously frozen human PBMC by negative selection, according to the manufacturer's instructions.
2. Mix CD4⁺ T cells and CD8⁺ T cells at a 1:1 ratio.
3. Culture cells in T Cell Medium at 1 x 10⁶ cells/ml, at 37°C in 5% CO₂ overnight.

Day 1:

1. Activate T cells with the Human CD3/CD28/CD2 T cell Activator reagent, following the vendor's recommendation and incubate at 37°C in 5% CO₂ for 24 - 48 hours.

Day 3:

1. Prepare the Cas9 RNP Complex in Resuspension Buffer R (provided with Neon NxT Electroporation System) as shown in the table below.

Note: Always include extra volume to avoid pipetting errors and the creation of bubbles.

Component	Amount
HLA-A/B/C gRNA	0.5 μ l
Cas9 enzyme	0.5 μ l
Resuspension Buffer R	4 μ l
Total	5 μl

2. Incubate the mixture at Room Temperature (RT) for 15 minutes.
3. Spin down the cells at 300 x g for 5 minutes, remove the medium and wash the cells with PBS.
4. Spin down the cells again, and resuspend the cell pellet in Resuspension Buffer R at a density of 4×10^7 cells/ml.
5. For each reaction, add 5 μ l of the cell suspension to 5 μ l of the Cas9 RNP Complex Resuspension in Buffer R (from step 1).
6. Using the 10 μ l Neon™ tip, pipet 10 μ l of cells/Cas9 RNP complexes, and electroporate using an optimized protocol for primary T cells (1,600V, 10 ms, 3 pulses).



Note: Avoid creating bubbles, which can affect the electroporation efficiency.

7. After electroporation, immediately transfer the contents of the Neon tip into one well of the 6-well culture plate containing 2 ml of prewarmed T Cell Medium.
8. Incubate the cells at 37°C for 2-3 days.

Day 5-6:

1. Analyze the HLA-A/B/C knockout efficiency by flow cytometry, or other method of interest.

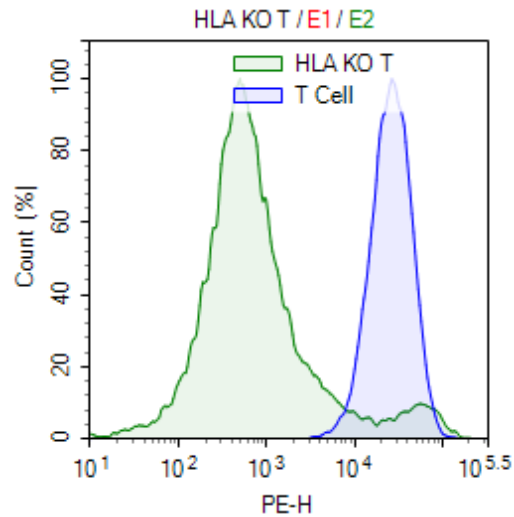


Figure 1: HLA-A/B/C knockout in CD4⁺ and CD8⁺ T cells by flow cytometry.

Approximately 250,000 CD4⁺ and CD8⁺ activated T cells were mixed with Cas9 RNP Complexes were electroporated using the Neon NxT Electroporation System. 72 hours after electroporation, the HLA-A/B/C expression was analyzed by flow cytometry using PE-labeled anti-human HLA-A, B, C Antibody (BioLegend #311405). Non-electroporated T cells were used as control (blue). T cells where TCR was knockout are shown in green. The y axis represents the % of cells, while the x axis represents the fluorophore intensity.

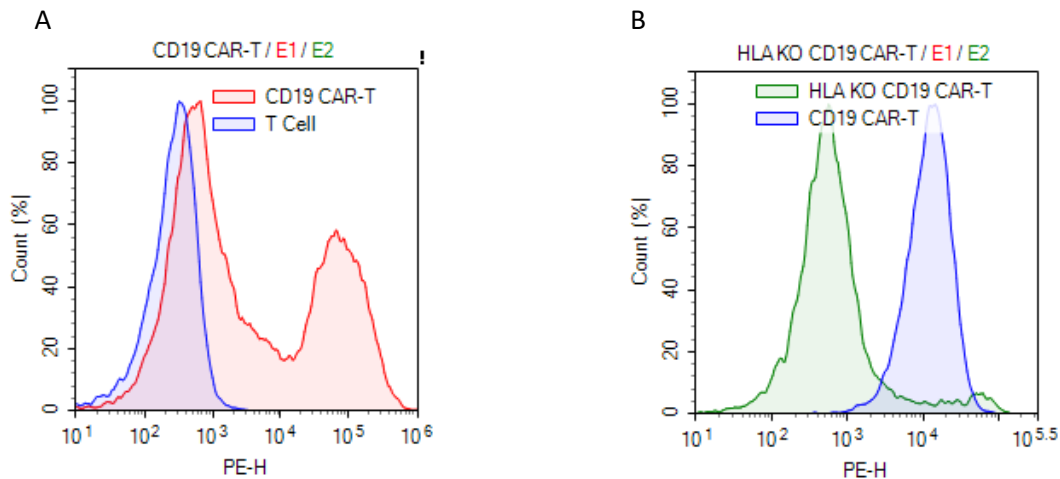


Figure 2: HLA-A/B/C Knockout in CD19 CAR T cells.

CD4⁺ and CD8⁺ activated T cells were transduced with Anti-CD19 CAR Lentivirus (CD19 ScFv-CD8-4-1BB-CD3 ζ ; SIN Vector) (#78601). 72 hours after transduction, approximately 250,000 transduced cells were mixed with Cas9 RNP complexes and electroporated using the Neon NxT Electroporation System. 72 hours after electroporation, CD19 CAR expression was analyzed by flow cytometry with FITC-Labeled Monoclonal Anti-FMC63 Antibody, Mouse IgG1 (AcroBiosystems #FM3-FY45G0) (panel A), and the HLA-A/B/C expression was analyzed using a PE-labeled anti-human HLA-A, B, C Antibody (BioLegend #311405) (panel B).

Gene Target	gRNA Sequence
HLA-A/B/C	CGGCTACTACAACCAGAGCG

Figure 3: gRNA sequence of the gRNA included in the HLA-A/B/C Knockout Electroporation Kit.

References

Leidner R., et al., 2022 *N Engl J Med* 386:2112-2119.

Tran E., et al., 2016 *N Engl J Med* 375:2255-2262.

Troubleshooting Guide

Visit bpsbioscience.com/cell-line-faq for detailed troubleshooting instructions. For all further questions, please email support@bpsbioscience.com.

Related Products

Products	Catalog #	Size
TCR Knockout Electroporation Kit	82394	1 kit
HLA-C*08:02 K562 Cell Line	78974	2 vials
HLA-C*08:02 Lentivirus	78930	500 µl x 2
HLA-E Lentivirus	78929	500 µl x 2

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