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# Q-Plex™ NHP

Version 1.1  
For Research Use Only  
Not For Use in Diagnostic Procedures

Symbol	Explanation
	Catalog Number
	Lot Number
	Use By YYYY-MM-DD
	Temperature Limitation
	Manufacturer
	Keep Away from Sunlight

# TABLE OF CONTENTS

Name and Intended Use	2
Principle of the assay	3
Sample Collection and Storage	4
Warnings and Precautions	5
Kit Contents & Storage	6
Materials Required but not Supplied	7
Assay Preparation	8
Assay Procedure	9
Analyzing a Q-Plex™ Image	12
Performance Characteristics	13
Calibrator Curves	13
Typical Data	14
Sensitivity	15
Precision	15
Linearity/Parallelism	16
Specificity	16
Appendix A: Plate Washing Methods	17
Automated Wash Method	17
Multichannel Pipette Wash Method	18
Abbreviated Protocol	19
Preparation	19
Running the Assay	19
Plate Diagram	20



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# NAME AND INTENDED USE

## Q-Plex™ Non-Human Primate

The Q-Plex Non-Human Primate (NHP) is a chemiluminescent Enzyme-Linked Immunosorbent Assay (ELISA) intended for quantitative concurrent measurement of different proteins in serum, plasma, or cell culture supernatant samples. The kit accompanying this manual will contain a subset of the following analytes.

Ang-2	sIL-2Ra	IL-15	PDGF
CRP	IL-4	IL-17a	sTfR
CXCL-1	IL-5	IL-18	RANTES
Ferritin	IL-6	IP-10	TIMP-1
GCSF	sIL-6R	MCP-1	TIMP-2
GMCSF	IL-8	MCP-3	TNF $\alpha$
IFN $\gamma$	IL-10	MIF	sTNFRII
IL-1 $\beta$	IL-12p40	MIG	VEGF
sIL-1Ra	IL-12p70	MIP-1 $\alpha$	
IL-2	IL-13	MIP-1 $\beta$	

# PRINCIPLE OF THE ASSAY

This multiplex assay is based on the microplate sandwich enzyme immunoassay technique. Samples or calibrators are pipetted into wells of a 96-well plate arrayed with immobilized, analyte-specific antibodies that capture the protein of interest. After washing away any unbound protein, a mixture that contains biotinylated analyte-specific antibodies is added. The biotinylated antibodies complete the sandwich for each specific arrayed analyte. After washing away unbound biotinylated antibody, streptavidin-horseradish peroxidase (SHRP) is added. Following an additional wash, the amount of SHRP bound to each location of the array is proportional to the amount of protein initially captured.

The amount of conjugated enzyme on each location of the array is measured with the addition of a chemiluminescent substrate.

# SAMPLE COLLECTION AND STORAGE

The sample collection and storage conditions are intended as general guidelines. The Clinical and Laboratory Standards Institute (CLSI GP44-A4) recommends the following storage conditions for samples: Samples should be stored at room temperature no longer than 8 hours. If the assay will not be completed within 8 hours, the samples should be refrigerated at +2°C to +8°C. If the assay will not be completed within 48 hours, or if the samples will be stored beyond 48 hours, samples should be frozen at -20°C or lower. Samples should not be repeatedly frozen and thawed. Frozen samples must be mixed well after thawing and prior to testing. Do not use bacterially contaminated samples. It is the responsibility of the individual laboratory to use all available references and/or its own studies to determine its own specific stability criteria. Assay has shown sensitivity to Bilirubin at concentrations above 0.05 mg/mL, Intralipid at concentrations above 1.25 mg/mL, and Hemoglobin at concentrations above 5 mg/mL. It is not recommended to run samples containing higher levels than specified value due to interference with assay performance.

**Cell Culture Supernatants** – Remove particulates by centrifugation and assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Serum** – Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Plasma** – Collect plasma using EDTA, Citrate, or Heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

# WARNINGS AND PRECAUTIONS

1. Read all instructions before beginning test.
2. For research use only. Not for use in diagnostic procedures.
3. The kit should not be used beyond the expiration date on the kit label.
4. If running multiple kits, a calibration curve must be included for each 96-well plate. The curve from one plate cannot be used to calculate sample values from other plates.
5. Do not mix or substitute reagents with those from other kits or lots.
6. Pre-wet pipette tips 3 times by drawing up the liquid into the pipette and then dispensing back into the original vessel prior to the addition of samples, calibrators, or external controls to the microplate.
7. Load all calibrators, controls, and samples into the microplate within 10 minutes of each other.
8. Be exact with incubation times, particularly the streptavidin-horseradish peroxidase (SHRP) incubation.
9. Be exact when mixing Substrate A and B+ and mix thoroughly.
10. Do not allow the substrate or SHRP to be exposed to UV light, as this may degrade these reagents.
11. Do not allow the plate to dry out between steps.
12. All products are carefully validated; however due to the variability encountered in biological sample matrices, the possibility of interference or sample matrix effects cannot be excluded.



# KIT CONTENTS & STORAGE

**Unopened Kit** - ❄ Store at 2-8°C. Do not use past kit expiration date. Do not re-use.

Part/Description	Storage of opened/reconstituted material
<b>Q-Plex™ Array Microplate</b> 1 arrayed and blocked 96-well polystyrene micro-titer plate	❄ 2-8°C until kit expiration
<b>Wash Buffer Concentrate (20X)</b> Liquid, 50 mL/vial of a concentrated solution of buffered surfactant	
<b>Sample Diluent</b> Liquid, 10 mL/vial of a buffered protein solution with heterophilic antibody and rheumatoid factor blockers and preservatives	
<b>Detection Mix</b> Lyophilized, biotinylated antibodies in a buffered protein base with preservatives	
<b>Calibrator</b> Lyophilized, recombinant antigens in a buffered protein base	Discard unused reconstituted calibrator.
<b>Streptavidin-HRP</b> Liquid, 6 mL/vial of streptavidin-conjugated horseradish peroxidase	☀ Do not expose to UV light. ❄ 2-8°C until kit expiration
<b>Substrate A</b> Liquid, 3.5 mL/vial	☀ Do not expose to UV light. ❄ Store mixed substrate solution at room temperature (20-25°C) for up to 1 week. ❄ Store unmixed solutions at 2-8°C until kit expiration.
<b>Substrate B+</b> Liquid, 3.5 mL/vial	
<b>Plate Seals (3)</b> Adhesive strips	Non-perishable

# MATERIALS REQUIRED BUT NOT SUPPLIED

In addition to the kit contents listed, the following materials are required to run this assay:

1. Multichannel pipette (20-200  $\mu\text{L}$ ) and/or single channel pipettes (20-1000  $\mu\text{L}$ ) with appropriate tips
2. 10 mL serological pipette
3. Polypropylene tubes or polypropylene 96-well plate(s) for sample and calibrator preparation
  - a. Example: Nunc® MicroWell™ 96-Well Plates, Polypropylene, 249944; Eppendorf® LoBind Protein or Genomic Microcentrifuge Tubes, 022431102
4. Q-View™ Imager and Q-View™ Software
5. Microplate shaker capable of 500 RPM
6. Deionized water
7. Microplate washer (An optional multichannel pipette protocol is provided in Appendix A)
8. 1 L graduated cylinder for the preparation of wash buffer.

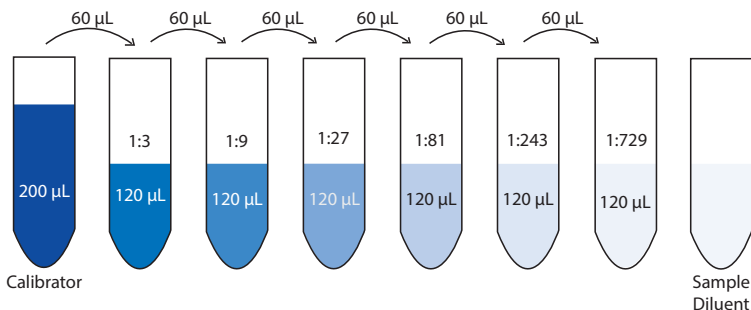
# ASSAY PREPARATION

1. Install Q-View Software on any computers that will be used for analysis or operating a Q-View Imager.
2. Set up the imager. For imager-specific instructions, see [www.quansysbio.com/manuals](http://www.quansysbio.com/manuals).
3. Set the microplate shaker to 500 RPM.
4. Allow all kit components to come to room temperature (20-25°C) prior to use.
5. Prepare Wash Buffer: Place 50 mL of Wash Buffer Concentrate (20X) into 950 mL deionized water and mix thoroughly.
6. Prepare Calibrator: Reconstitute lyophilized Calibrator using Sample Diluent with the volume specified on the In-Kit Certificate of Analysis accompanying the kit. Allow to sit for 5 minutes. Mix thoroughly. Use prepared Calibrator within 30 minutes.
7. Prepare Detection Mix: Reconstitute lyophilized Detection Mix using 6 mL of deionized water. Allow to sit for 5 minutes. Mix thoroughly.
8. Prepare chemiluminescent substrate: Combine 3 mL of Substrate A with 3 mL of Substrate B+ and mix gently. **Do not expose to UV light. Store at room temperature (20-25°C) after mixing.**

# ASSAY PROCEDURE

**Allow all reagents to equilibrate to room temperature (20-25°C) before use and prepare as directed in the previous sections. It is recommended that all calibrators, samples, and controls be assayed in duplicate.**

- Using the previously prepared Calibrator (see "Assay Preparation" step 6), prepare an 8-point calibration curve (7 points plus 1 blank) in either polypropylene tubes or a polypropylene 96-well plate (see "Materials Required but Not Supplied" for example tubes and plates).
  - Pipette 200  $\mu\text{L}$  of prepared Calibrator into the first tube or well.
  - Pipette 120  $\mu\text{L}$  of Sample Diluent into the other 7 tubes or wells.
  - Transfer 60  $\mu\text{L}$  of undiluted prepared Calibrator from the first tube or well into the second, mix thoroughly, and repeat the transfer from tube to tube or well to well for 5 more points, leaving the last tube or well without any prepared Calibrator. This process is diagrammed below. The undiluted prepared Calibrator serves as the high point of the calibration curve. The Sample Diluent serves as the blank.



- Prepare samples by diluting 1:2 (1 part sample to 1 part Sample Diluent) in either polypropylene tubes or a polypropylene 96-well plate in a volume sufficient to provide 50  $\mu\text{L}$  of diluted sample per well.

**Note:** If you anticipate that your analyte concentrations will be greater than the high point of the calibration curves as reported on the In-Kit Certificate of Analysis, use sample diluent to dilute your samples further.

3. Add 50  $\mu\text{L}$  per well of the prepared calibration curve and samples to the Q-Plex™ Array Microplate. Load all samples and calibration curve to the plate within 10 minutes.
4. Cover the plate with a plate seal and incubate on a plate shaker set to 500 RPM for 120 minutes at room temperature (20-25°C).
5. Wash the plate 3 times (see Appendix A).
6. Add 50  $\mu\text{L}$  per well of Detection Mix, cover with a new plate seal, and return to the plate shaker set to 500 RPM for 90 minutes at room temperature (20-25°C).
7. Wash the plate 3 times (see Appendix A).
8. Add 50  $\mu\text{L}$  per well of Streptavidin-HRP, cover with a new plate seal, and return to the plate shaker set to 500 RPM for 20 minutes at room temperature (20-25°C).
9. Wash the plate 6 times (see Appendix A).
10. Add 50  $\mu\text{L}$  per well of previously prepared substrate (see “Assay Preparation” step 8). Image plate immediately. Wait no longer than 5 minutes to commence imaging.

**Note:** If imaging cannot commence immediately, protect the plate from drying for up to 15 minutes by dispensing 100  $\mu\text{L}$  of wash buffer into each well of the plate prior to adding the mixed substrate. When ready to image, remove the wash buffer from the plate and add the substrate.

11. Place the plate in the Q-View Imager.
12. Open Q-View Software, create or open a project, and click Acquire Image.
13. For imager specific recommended exposure times, see [www.quansysbio.com/manuals](http://www.quansysbio.com/manuals).
14. Click the Capture Image(s) button. Users may perform Well Assignment while images are being captured.

*Note:* Details about these imaging steps are available in the Q-View Software Manual viewable at [www.quansysbio.com/manuals](http://www.quansysbio.com/manuals) or within Q-View Software under **Support > Manual**.

15. Dispose of all used and unused materials in accordance with local regulations.

# ANALYZING A Q-PLEX™ IMAGE

The following summarizes a general workflow for analyzing a Q-Plex image in Q-View Software. Each of these steps is described in greater detail in the Q-View Software and Imager manuals, viewable at [www.quansysbio.com/support](http://www.quansysbio.com/support) or within Q-View Software under **Support > Manual**.

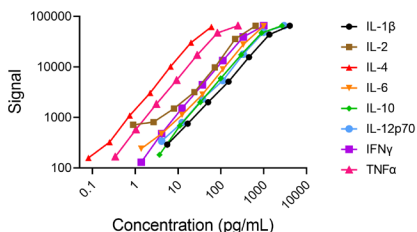
1. Acquire or import an image into Q-View Software as previously described.
2. Enter the **Software Product Code** (found on the In-Kit Certificate of Analysis) into the **Product Code** field of the software.
3. **Image Processing:** Align the plate overlay as follows:
  - a. Set the overlay: If using the Auto-Set Plate Overlay feature, this will occur automatically. Otherwise, go to **Overlay Options > Set Plate Overlay**
  - b. To visualize bright or dim spots, optimize the display using **Image Options > Adjust Gamma** (does not affect the data).
  - c. Optimize overlay alignment: Go to **Overlay Options > Adjust Plate Overlay** to pivot the overlay, **Adjust Well** and **Adjust Spot** to move individual wells and spots, then **Auto-Adjust Spots** to automatically snap each circle of the overlay to the nearest spot of the image beneath.
4. **Well Assignment:** Label wells as samples, calibrators, or negatives (blank), and specify their dilution factors. Use Templates to quickly assign layouts that are repeated often or export the layout as a .csv file.
5. **Data Analysis:** Once Image Processing and Well Assignment are complete, select Data Analysis. Click Perform Analysis to generate charts, concentrations, and statistics based on optimal regression settings provided by the product definition.

# PERFORMANCE CHARACTERISTICS

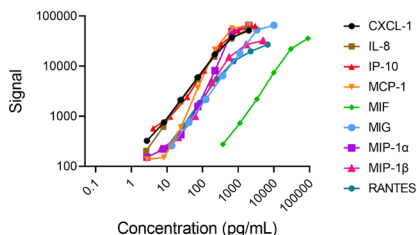
## Calibrator Curves

Expected signal for calibrator curves on Q-Plex assays vary depending on the imager model used and, to a lesser extent, between kit lots. Using a Q-Plex kit on a Q-Plex imager without negative subtraction turned on, the minimum signal observed will be around 100 and the maximum can be up to 65535. Below are examples of calibrator curves demonstrating the types of curves you may see on your kit. If you have concerns about your assay performance, please contact support at 1-888-QUANSYS (1-888-782-6797) or at [support@quansysbio.com](mailto:support@quansysbio.com).

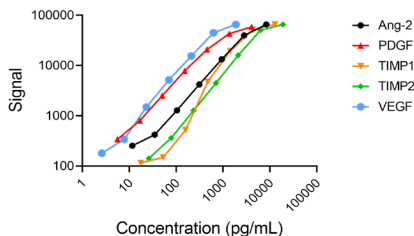
Typical Calibrator Curves NHP Cytokine Screen



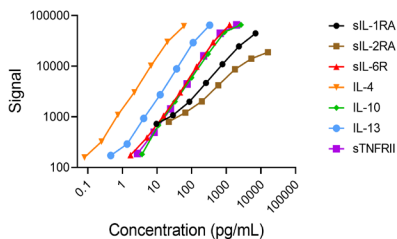
Typical Calibration Curves NHP Chemokine Screen



Typical Calibration Curves NHP Cancer Panel 1



Typical Calibration Curves NHP Anti-Inflammatory Panel 1





## Typical Data

Representative data from one lot of kits are shown below to demonstrate typical calibrator curve performance. See below for explanation of data calculations.

Analyte	Units	ULOQ	LLOQ	LLD	Intra-Assay Precision (%CV)	Inter-Assay Precision (%CV)	Linearity Range	Linearity Average
<b>Ang-2</b>	pg/mL	8458	11.60	5.80	7%	9%	69-115%	94%
<b>CRP</b>	mg/L	0.70	0.0018	0.00089	6%	5%	92-113%	103%
<b>CXCL-1</b>	pg/mL	1767	2.29	0.33	4%	8%	73-119%	94%
<b>Ferritin</b>	µg/L	21	0.029	0.015	8%	7%	82-98%	87%
<b>GCSF</b>	pg/mL	4419	6.06	3.03	7%	14%	84-113%	100%
<b>GMCSF</b>	pg/mL	5974	8.53	5.29	8%	8%	93-178%	111%
<b>IFN<math>\gamma</math></b>	pg/mL	1000	1.22	0.52	5%	8%	101-145%	118%
<b>IL-1<math>\beta</math></b>	pg/mL	4075	5.26	1.03	7%	9%	110-124%	116%*
<b>sIL-1Ra</b>	pg/mL	6506	8.62	4.99	6%	9%	81-122%	103%
<b>IL-2</b>	pg/mL	558	0.8	1.04	9%	7%*	108-134%	118%
<b>sIL-2Ra</b>	pg/mL	15921	21.94	2.94	12%	13%	85-136%	111%
<b>IL-4</b>	pg/mL	42	0.11	0.05	12%	16%*	95-120%	109%
<b>IL-5</b>	pg/mL	334	0.47	0.073	9%	17%	74-125%	98%
<b>IL-6</b>	pg/mL	986	1.6	0.044	6%	16%	98-125%	107%
<b>sIL-6R</b>	pg/mL	1106	1.35	0.1	5%	6%	95-131%	109%
<b>IL-8</b>	pg/mL	1921	2.61	0.47	6%	12%	74-131%	99%*
<b>IL-10</b>	pg/mL	2679	3.42	0.47	4%	13%	92-120%	102%
<b>IL-12p40</b>	pg/mL	2525	3.17	0.22	8%	14%	66-121%	92%
<b>IL-12p70</b>	pg/mL	2480	3.98	0.65	12%	16%	98-112%	106%
<b>IL-13</b>	pg/mL	333	0.44	0.25	8%	11%*	84-119%	95%
<b>IL-15</b>	pg/mL	3549	4.81	2.53	7%	18%*	88-109%	97%
<b>IL-17a</b>	pg/mL	5590	8.15	2.55	13%	19%	110-121%	117%
<b>IL-18</b>	pg/mL	235	0.32	0.16	4%	13%	89-114%	103%
<b>IP-10</b>	pg/mL	952	3.62	1.24	4%	20%	73-121%	93%
<b>MCP-1</b>	pg/mL	1882	6.98	4.7	4%	12%	65-123%	82%*
<b>MCP-3</b>	pg/mL	101	0.12	0.12	6%	16%*	76-132%	98%*
<b>MIF</b>	pg/mL	278574	361.25	39.93	4%	20%	67-161%	113%
<b>MIG</b>	pg/mL	12372	13.85	2.84	5%	16%	88-114%	104%

Analyte	Units	ULOQ	LLOQ	LLD	Intra-Assay Precision (%CV)	Inter-Assay Precision (%CV)	Linearity Range	Linearity Average
<b>MIP-1<math>\alpha</math></b>	pg/mL	1782	2.86	0.90	4%	11%	82-160%	120%*
<b>MIP-1<math>\beta</math></b>	pg/mL	5163	5.92	3.81	6%	8%	99-130%	110%
<b>PDGF</b>	pg/mL	4099	5.62	2.81	12%	11%	72-118%	93%
<b>RANTES</b>	pg/mL	6956	8.5	2.53	12%	17%	90-153%	113%*
<b>sTfR</b>	mg/L	3.18	0.0044	0.0022	11%	10%	90-112%	101%
<b>TIMP-1</b>	pg/mL	12844	17.62	8.81	11%	12%	70-130%	110%
<b>TIMP-2</b>	pg/mL	19136	26.25	13.12	7%	10%	71-132%	107%
<b>TNF<math>\alpha</math></b>	pg/mL	250	0.29	0.019	3%	11%	73-112%	98%
<b>sTNFR<math>\text{II}</math></b>	pg/mL	1886	2.52	0.33	3%	9%	91-139%	115%*
<b>VEGF</b>	pg/mL	1892	2.60	1.30	6%	8%	62-115%	81%

\*Samples out of range from the ULOQ and LLOQ were excluded from this assay analysis

## Sensitivity

The upper limit of quantitation (ULOQ) is the highest concentration at which the calculated recovery of the analyte remains within 80-120% of the known value.

The lower limit of quantitation (LLOQ) is the lowest concentration at which the calculated recovery of the analyte remains within 75-125% of the known value and the CV of the calculated concentration is <25%.

The lower limit of detection (LLD) is the concentration that is two standard deviations above the average signal of twenty blank (zero) replicates.

## Precision

Intra-Assay Precision – 3 samples containing and/or spiked with various concentrations of each analyte were tested 20 times on 1 plate to assess intra-assay precision. The average %CV of all samples is reported.

Inter-Assay Precision – 3 samples containing and/or spiked with various concentrations of each analyte were tested on at least 10 individual assays by at least 3 users. The average %CV of all samples is reported.

## Linearity/Parallelism

Samples containing and/or spiked with various concentrations of each analyte were diluted with sample diluent at dilutions of 1:2, 1:4, 1:8, 1:16 and assayed. The highest, lowest, and average dilution recovery are reported.

## Specificity

Specificity of the array was assessed by testing each recombinant antigen at the concentration of the highest calibrator in the multiplexed assay. The IL-12p40 assay has less than 5% cross-reactivity with IL-12p70. The cross-reactivity of all other antigens on each assay was less than 1%.

# APPENDIX A: PLATE WASHING METHODS

## Automated Wash Method

1. Use a program that will aspirate and dispense 300-400  $\mu\text{L}$  wash buffer.



2. Ensure that the plate washer has a program that will not scratch the bottom of the microplate but will prevent plate drying. This is critical to prevent damage to the capture antibody arrays. The simplest method to avoid plate drying is to leave a small, uniform amount (1-3  $\mu\text{L}$ ) of wash in the well after the final aspiration and add the next reagent to the plate as quickly as possible.
3. Leaving a uniform amount of wash in the wells can be accomplished with an automated washer by slightly increasing the aspiration height of the washer head.

For example:

<b>Process</b>	<b>Distance</b>	<b>Steps on a Biotek ELX-405</b>
Aspiration Height	3.81 mm	30
Aspiration Position	1.28 mm from center	-28
Dispense Height	15.24 mm	120

No soak or shake cycles are needed during wash steps

4. Connect the prepared wash buffer to your automatic plate washer.
5. Run 1-2 priming cycles to make sure that the wash buffer is running through the plate washer. When the buffer has run through the machine, the waste will be foamy.
6. In a spare microtiter plate, dispense 100  $\mu$ L wash buffer, ensure that all pins dispense uniformly, then aspirate and ensure that all pins aspirate uniformly. This will ensure that all pins are functioning.
7. Prime the plate washer 1 time before the first wash step. When running the assay, perform the wash 3 or 6 times according to the protocol.

## Multichannel Pipette Wash Method

1. Just prior to washing, pour the prepared wash buffer into a trough or tray.
2. After each incubation, flick the solution out of the plate over a waste container before starting the wash protocol.
3. Using a multichannel pipette, dispense 400  $\mu$ L of wash buffer into each of the wells used in the test.
4. Aggressively flick the wash buffer out over a waste container.
5. This washes the plate 1 time. When the assay procedure calls for 3 or 6 washes, repeat steps 3-4 accordingly.
6. Tap the plate upside down on a paper towel to remove any residual wash.
7. Proceed immediately to dispense the next solution so drying does not occur.

# ABBREVIATED PROTOCOL

## Preparation

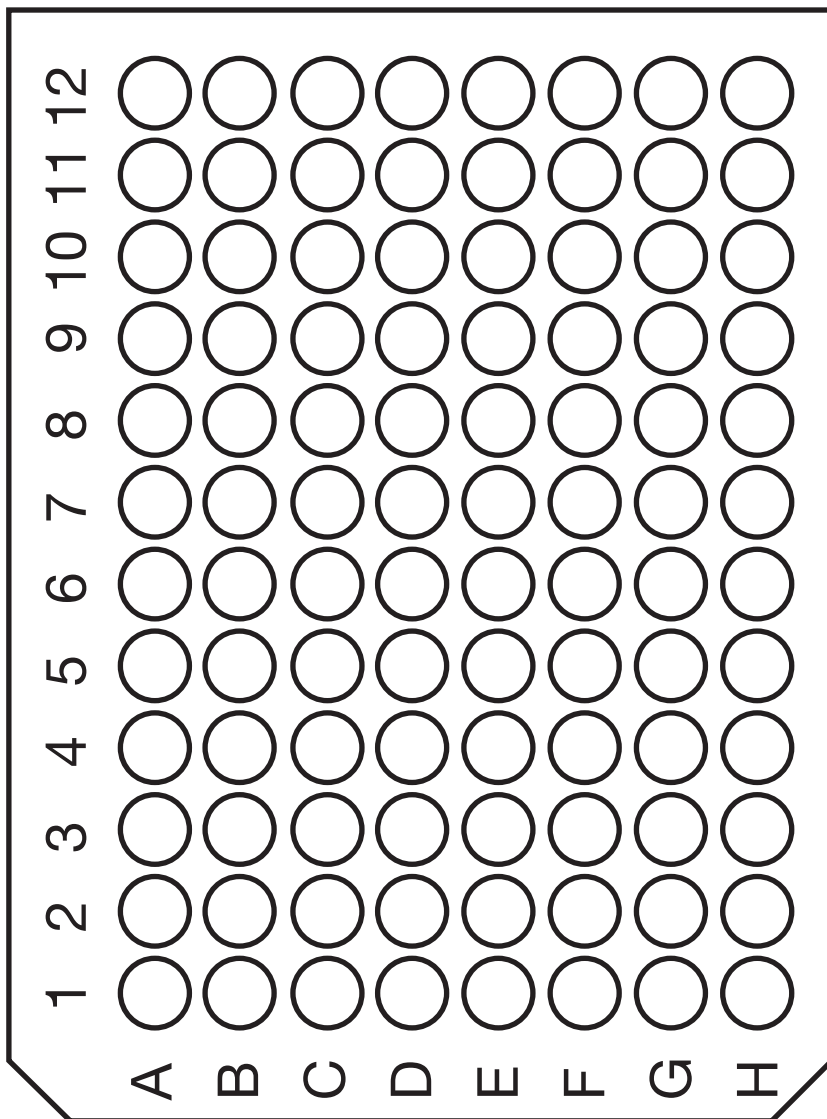
1. Install Q-View Software (page 8).
2. Set up the imager (page 8).
3. Set up microplate washer (page 17) and shaker (page 8).
4. Reconstitute and prepare reagents (page 8).

## Running the Assay

5. Prepare the calibration curve using the prepared Calibrator and Sample Diluent according to the In-Kit Certificate of Analysis (page 9).
6. Prepare the samples with Sample Diluent (page 9).
7. Load the calibration curve and samples onto the plate. Shake for 120 minutes (500 RPM) at room temperature (page 10).
8. Wash the plate 3 times, add the Detection Mix, and shake for 90 minutes (500 RPM) at room temperature (page 10).
9. Wash the plate 3 times, add the Streptavidin-HRP, and shake for 20 minutes (500 RPM) at room temperature (page 10).
10. Wash the plate 6 times and add the prepared Substrate (page 10).
11. Capture and analyze image of the plate (page 11).

We take great care to ensure that customers have success using our products and services. If you have further questions about running the assay, data analysis, troubleshooting, or our products or services, please contact us at [1-888-QUANSYS \(1-888-782-6797\)](tel:1-888-QUANSYS) or at [support@quansysbio.com](mailto:support@quansysbio.com).

# PLATE DIAGRAM







# Q-Plex™ ARRAY

For technical support, email: [support@quansysbio.com](mailto:support@quansysbio.com)

109217MB

365 North 600 West, Logan, Utah 84321  
Tel: 1-888-782-6797 | Fax: (435)750-6869 | [www.quansysbio.com](http://www.quansysbio.com)