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Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

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Please contact CEDARLANE® for lot specific information.

### PE-Cy5 Anti-Mouse γδ TCR Monoclonal Antibody

CL7201TC LOT:05020203

#### **DESCRIPTION:**

Cedarlane's anti-mouse  $\gamma\delta$  T cell receptor monoclonal antibody reacts with the surface on all  $\gamma\delta$  TCR bearing cells and does not react with receptors on  $\alpha\beta$  TCR positive cells. It is thought that this clone may be specific for a determinant present on C $\delta$  <sup>7</sup>. The  $\gamma\delta$  T cell receptors are present on murine CD4-CD8- thymocytes, peripheral T cells, intestinal CD8+ intraepithelial lymphocytes and Thy 1+ dendritic epidermal cells in the skin <sup>1</sup>.

Use of this antibody in conjunction with an anti-CD3 monoclonal antibody (Cedarlane's anti-CD3 $\epsilon$  Monoclonal Antibody CL7202F) allows for accurate measurements of the mutually exclusive sub-populations of  $\gamma\delta$  TCR and  $\alpha\beta$  TCR bearing T cells. Cedarlane's anti mouse  $\gamma\delta$  TCR monoclonal antibody has also been used successfully for the characterization of murine intraepithelial lymphocytes.

Applications include: flow cytometry.

#### PRESENTATION:

100 μg PE-Cy5 conjugated Ig buffered in PBS, 0.1% NaN<sub>3</sub> and EIA grade BSA as a stabilizing protein to bring total protein concentration to 4-5 mg/ml.

#### STORAGE/STABILITY:

Store at 4°C. **DO NOT FREEZE**. Avoid prolonged exposure to light. If the reagent is being diluted it is recommended that only the quantity to be used within one week be diluted. Check label for expiry date.

For more information or to place an order please contact...



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#### **SPECIFICATIONS:**

Clone: GL-3

Specificity: Mouse γδ T cell receptor

Ig Class: Hamster IgG

<u>PE-Cy5 Excitation</u>: 488 nm <u>PE-Cy5 Emmission</u>: 667 nm

Notes: PE-Cy5 conjugates require a 650 nm long pass filter in the FL3 channel.

FL2-FL3 compensation will be in the range of 1%.

Antibody Concentration: 0.2 mg/ml

#### **FLOW CYTOMETRY ANALYSIS:**

#### Method:

- 1. Prepare a cell suspension in media A. For cell preparations, deplete the red blood cell population with Lympholyte®-M cell separation medium (CL5030).
- 2. Wash 2 times.
- 3. Resuspend the cells to a concentration of  $2x10^7$  cells/ml in media A. Add 50  $\mu$ l of this suspension to each tube (each tube will then contain  $1 \times 10^6$  cells, representing 1 test).
- 4. To each tube, add  $\sim 1.0 \,\mu\text{g}^*$  of **CL7201TC** per  $10^6$  cells.
- 5. Vortex the tubes to ensure thorough mixing of antibody and cells.
- 6. Incubate the tubes for 30 minutes at 4°C. (It is recommended that the tubes are protected from light, since most flurochromes are light sensitive.)
- 7. Wash 2 times at 4°C.
- 8. Resuspend the cell pellet in 50 μl ice cold media B.
- 9. Transfer to suitable tubes for flow cytometric analysis containing 15 μl of propidium iodide at 0.5 mg/ml in PBS. This stains dead cells by intercalating in DNA.

#### Media:

- A. Phosphate buffered saline (pH 7.2) + 5% normal serum of host species + so-dium azide (100  $\mu$ l of 2M sodium azide in 100 mls).
- B. Phosphate buffered saline (pH 7.2) + 0.5% Bovine serum albumin + sodium azide (100  $\mu$ l of 2M sodium azide in 100 mls).

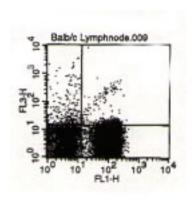
#### Results:

Tissue Distribution by flow Cytometry Analysis:

(Representative Histogram) Mouse Strain: BALB/c

Cell Concentration: 1 x 10<sup>6</sup>cells per test Antibody Concentration used: 10ìl /10<sup>6</sup>

Isotypic Control: Hamster IgG Pe-Cy5 (CLCHM06)



Cell Source: Lymph Node

- N.B. Appropriate control samples should always be included in any labeling studies.
  - \* For optimal results in various applications, it is recommended that each investigator determine dilutions appropriate for individual use.

#### REFERENCES:

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- Cron. R & et al. 1988. A functional subpopulation of peripheral murine T lymphocytes which express a novel T Cell Structure. J. Immunol. 141:1074.
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- 5. Goodman, T & L. Lefrancois. 1988. Expression of the γδ TCR on intestinal CD8<sup>+</sup> intraepithelial lymphocytes. Nature (Lond.) **333**:855.
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- 7. Goodman, T & L. Lefrancois. 1989. Intraepithelial Lymphocytes. J. Exp. Med. Vol. 170:1569-1581.

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