



SZABO SCANDIC

Part of Europa Biosite

Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

Weitere Information auf den folgenden Seiten!
See the following pages for more information!



Lieferung & Zahlungsart

siehe unsere [Liefer- und Versandbedingungen](#)

Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

mail@szabo-scandic.com

www.szabo-scandic.com

[linkedin.com/company/szaboscandic](https://www.linkedin.com/company/szaboscandic) 

Technically
Speaking

CEDARLANE[®] 
www.cedarlanelabs.com

Conveniently Delivering You Today's Innovations
for the Science of Tomorrow™

Rabbit Anti-Mouse/Rat Asialo GM1 Polyclonal Antibody

CL8955

LOT: KQF6371

DESCRIPTION:

Cedarlane's Anti-Asialo GM1 polyclonal antibody reacts with mouse and rat Natural Killer (NK) cells. It also exhibits slight reactivity with mouse monocytes (liver cells which contain no NK cells; bone marrow; fetal liver cells; spleen cells of nude mice), macrophages, and fetal thymocytes (12 days old; ratio of existence decreased gradually until there were none in newborn mice). Cedarlane's anti-Asialo GM1 antiserum has been shown to eliminate NK activity in cells of various strains of mice and rats.

Applications include: flow cytometry, immunohistochemistry (frozen sections and formalin-fixed paraffin embedded sections), immunoprecipitation and NK cell depletion.

SIZE: 1 ml

RECONSTITUTION:

Distilled water is recommended (1 ml). Since the material is lyophilized with salts, use of other solvents such as PBS or MEM may increase the salt concentration.

STORAGE/STABILITY:

Store at 4°C before reconstitution: stable for 2 years when refrigerated.

After reconstitution: stable for 2-3 months when refrigerated, 2-3 days at room temperature. **DO NOT FREEZE.**

PREPARATION:

Rabbits were immunized with asialo GM1 purified from bovine brain tissue, methylated bovine serum albumin and complete Freund's adjuvant. The gammaglobulin fraction of serum was obtained by 50% ammonium sulfate precipitation followed by dialysis with phosphate buffered saline (pH 7.2).

SPECIFICATIONS:

Ig Classes: IgG, IgA, and IgM

Antibody Titre: Approximately 1:1000 by immunoflocculation test.

Total Protein Concentration: 40 mg/ml

Albumin Concentration: 8 mg/ml

Note: Since this antibody is not a purified product, the protein concentration is not equal to the concentration of Asialo-GM1-specific antibody, even after correcting for the albumin content. Both the total protein and albumin concentrations will vary from lot to lot.

Visit our website for your local distributor.

CEDARLANE[®] 

www.cedarlanelabs.com

An ISO 9001:2000 and ISO 13485:2003
registered company.

In CANADA: Toll Free: 1-800-268-5058

4410 Paletta Court, Burlington, ON L7L 5R2 ph: (289) 288-0001, fax: (289) 288-0020
e-mail: general@cedarlanelabs.com

In the USA: Toll Free: 1-800-721-1644

1210 Turrentine Street, Burlington, NC 27215 ph: (336) 513-5135, fax: (336) 513-5138
e-mail: service@cedarlanelabs.com

Injections:

Mouse - intravenously: 10-50 μ l (approximately 20 μ l)*. The exact dosage should be decided from titration data enclosed with package (please see following page) and the nature of the study. The first injection may be effective for 4 days with a gradual diminution. Therefore, 3-4 injections are necessary for a 2 week study.

(incubation)				
Days	<u>0</u>	<u>5</u>	<u>10</u>	<u>14</u>
Injection	1st	2nd	3rd	4th

* 50 injections can be made using 20 μ l doses.

Rat - intravenously: 50 - 250 μ l (4 or 5 times the usual mouse dose is required). Health conditions and weight of rats should be taken into consideration. It is recommended that the researcher assay NK activity to determine the proper dosage.

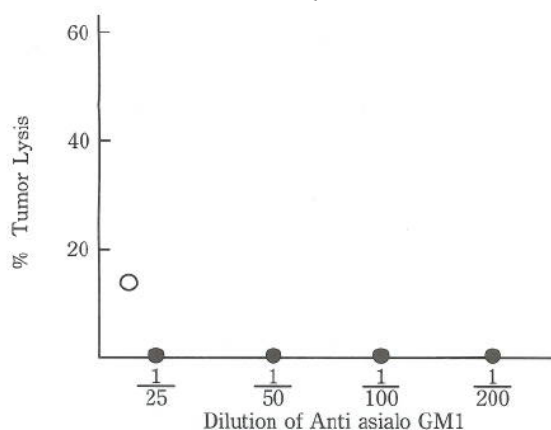
Mouse and rat - intraperitoneally: Dosage should be equal to or greater than the i.v. dosage.

Each package comes with *in vivo* and *in vitro* titration data for the lot.

ANTI-ASIALO GM1
Product code No. CL8955
Lot No. KQF6371

Titration of Anti-Asialo GM1 *in vitro*

Lot No. KQF6371



Spleen cells of BALB/c were treated with Anti-Asialo GM1 and Cedarlane Low-Tox[®] Guinea Pig Complement. Remaining NK activities were tested *in vitro* by using YAC-1 cells as target (●—●). Effector/target ratio was 50:1. O represents NK activities of BALB/c spleen cells treated with complement

Titration of Anti-Asialo GM1 *in vivo*

Amount of Anti-Asialo GM1 i.v. injected into BALB/c* mice	% Lysis against YAC-1 cells by spleen cells taken 3 days after a 1-shot injection (Effector/target 50:1)
10 μ l	2.8
25 μ l	1.8
50 μ l	1.4
100 μ l	2.5
Normal rabbit serum injected	% Lysis against YAC-1 cells
100 μ l	7.6

• Note: BALB/c mice were injected with 100 μ g of polyinosinic-polycytidylic acid sodium salt (0.2 ml of 500 μ g/ml Poly I:C) and maintained for 18 hours before next procedure.

Procedure For Measurement of Anti-NK Cell Activity

IN VITRO:

1. Preparation of target cells:
 - Suspend 5×10^6 cells of YAC-1 in RPMI 1640 containing 10% FCS.
2. Preparation of effector cells:
 - Inject 0.2 ml of polyinosinic- polycytidylic acid sodium salt solution (500 µg per ml of poly I:C in RPMI 1640) into BALB/c mice.
 - Remove spleens from mice on the following day (after approximately 10 hours of treatment) and prepare spleen cell suspension as follows:
 - Centrifuge at 1000 rpm for 10 minutes and discard supernatant.
 - Add 0.83% NH_4Cl to the tube to hemolyze the precipitated spleen cells.
 - Centrifuge at 1000 rpm for 10 minutes and discard supernatant. Add 10 ml RPMI 1640 to wash cells. Repeat washing process using same procedure.
 - Adjust the cell number to 2.5×10^7 cells per ml.
3. Treatment of effector cells:
 - Dilute the target cell suspension with RPMI 1640 to ratios of 1:50, 1:100, and 1:200.
 - Place 0.5 ml of effector cell suspension into centrifugation tubes. Centrifuge at 1000 rpm for 10 minutes and discard supernatant.
 - Add 0.5 ml of the diluted target cell suspension to each tube above and mix well.
 - Incubate tubes at 37°C for 30 minutes in 5% CO_2 .
 - Centrifuge at 1000 rpm for 10 minutes and discard supernatant.
 - Prepare control by adding 0.5 ml of RPMI 1640 to effector cells. Mix well to make suspension.
 - Make a 1:10 dilution of Cedarlane Low-Tox[®] Guinea Pig Complement with RPMI 1640 and add the diluted complement to effector cell suspensions. Mix well.
 - Incubate at 37°C for 30 minutes with occasional stirring.
 - Centrifuge at 1000 rpm for 10 minutes and discard supernatant.
 - Add 1 ml of inactivated FCS (10% FCS in RPMI 1640) and mix well.
4. Measurement of activity (n=2):
 - Place 100 µl of the target cell suspension into each well of microplate.
 - Place 100 µl each of the untreated samples, the diluted samples, and the samples of effector cells with complement into each well.
 - For measurement of spontaneous Cr release, add 100 µl of RPMI 1640 containing inactivated 10% FCS to well.
 - For measurement of maximum Cr release, centrifuge 0.5 ml of target cell suspension at 1000 rpm for 5 minutes. Discard supernatant and add 1.0 ml of sterile water.
 - Cover the microplate and tubes. Incubate at 37°C for 10 hours in 5% CO_2 .
 - Assay the radioactivity of 100 µl of each reaction mixture using an autogamma counter.
 - Calculate the % Lysis using the following equation:

$$\% \text{ Lysis} = \frac{\text{Experimental Cr release} - \text{Spontaneous Cr release}}{\text{Maximum Cr release} - \text{Spontaneous Cr release}} \times 100$$

IN VIVO:

1. Preparation of target cells:
 - Suspend 5×10^6 cells of YAC-1 in RPMI 1640 containing 10% FCS.
2. Preparation of effector cells:
 - Dilute the sample with RPMI 1640 to ratios of 1:2, 1:4, and 1:8
 - Inject BALB/c mice with diluted samples in 0.2 ml doses (n=3).
 - Dilute rabbit serum to 1:2 with RPMI 1640 and inject into BALB/c mice in 0.2 ml doses.
 - After 3 days of treatment, remove spleens and prepare suspensions using RPMI 1640 as follows:
 - Centrifuge at 1000 rpm for 10 minutes and discard supernatant.
 - Add 0.83% NH_4Cl to the tubes to hemolyze the precipitated spleen cells.
 - Centrifuge at 1000 rpm for 10 minutes and discard supernatant.
 - Add 10 ml of RPMI 1640 to the tubes to wash the precipitated cells. Repeat washing process using same procedure.
 - Centrifuge at 1000 rpm for 10 minutes and collect the cells. Add RPMI 1640 containing inactivated 10% FCS to the tubes to adjust the cell number to 2.5×10^7 cells per ml.
3. Measurement of activity:
 - Follow same procedure as outlined in section #4 under "*IN VITRO*".

Indirect immunoperoxidase method

FROZEN SECTION (6 μm)

1. Washed with PBS buffer (pH 7.2) for 10 min. (X3).
Added to PBS containing 0.05% gluteraldehyde.
2. Fixed for 5 min. at 4°C.
3. Washed with PBS buffer for 5 min.
Added to 100% methanol containing 0.5% H_2O_2 *1
*1: for removal of endogenous peroxidase activity
4. Incubated for 30 min.
5. Added to Anti normal goat serum *2.
*2: for avoiding non-specific antigen-antibody reaction.
Added to anti-asialo GM1, Rabbit (CL8955) as first antibody.
6. Incubate for one hour at room temperature, or overnight at 4°C.
7. Washed with PBS buffer for 5 min. (X3).
Added to anti-rabbit peroxidase goat IgG as second antibody.
8. Incubated for 30 min.-3 hr.
9. Washed with PBS buffer (X3).
Added to 100 ml of Karnowsky substrate solution *3
*3: Karnowsky substrate solution:

0.05M Tris-HCl buffer, pH 7.6	100 ml
Diaminobenzidine tetrachloride	0.25 mg
5% Hydrogen peroxide	0.1 ml
10. Incubated at RT for 3-30 min.
11. Transferred to distilled water (to stop)
Added to 1% OsO_4 (Osmic Acid Solution 1%) for 2-3 sec
Stained with methyl green.
Dehydrated with dilution series of ethanol.
Treated with xylene.
Embedded with balsam.

Control:

1. First antibodies were omitted.
2. First antibodies were replaced with non-immune sera of the same species as the specific antiserum.
3. DAB reaction only.
mounted with Hematoxylin-Eosin staining

REFERENCES:

1. Godney, E.K. and C.J. Gauntt: *Journal of Immunology*. 139, 913 (1987).
2. Habu, S., H. Fukui, M. Shimamura, M. Kasai, Y. Nagai, K. Okamura, and N. Tamaoki: *Journal of Immunology*. 127,34 (1981).
3. Kasai, M., M. Iwamori, Y. Nagai, K. Okamura, and T. Tada: *Eur. Journal of Immunology*. 10, 175 (1980).
4. Okamura, K. and Y. Ochali: *Metabolism*. 17, 47 (1980). (In Japanese)
5. Yao, L et al. *Blood*, VOL. 93, No 5 (March 1), 1999; pp1612-1621

IHC REFERENCES:

5. Salazar-Matcher, Orange and Biron: *J. Exp. Med.* 187 (1998) 1-14.
6. Nishiyama, Fuchimoto and Orita: *Jpn. J. Cancer Res.* 80 (1989) 366-372.
7. Basse, P. Hokland, Gundersen and M. Hokland: *APMIS*. 100 (1992) 202-208.
8. Wiltrout, Santoni, Peterson, Knott, Overton, Herberman and Holden: *Journal of Leukocyte Biology*. 37 (1985) 597-614.
9. Yeo, Eun-Jin, et al. *J Natl Cancer Inst.* 2003; 95 (7):516-25.
10. Keilbaugh, Sue A et al. *Gut*. 2005; 54(5):623-9.
11. Rosato, Antonio et al. *J Immunol*. 2003; 171(10):5172-9.

For Research Use Only
® is a Registered Trademark of Cedarlane