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TECHNICALLY Speaking

Place your order with CEDARLANE® or your local distributor.

Please contact CEDARLANE® for lot specific information.

Purified Anti-*Kudoa thyrsites*
Monoclonal Antibody

CL010AP
LOT: P1021

DESCRIPTION:

Cedarlane's Purified Anti-*Kudoa thyrsites* monoclonal antibody detects the histozoic parasite *Kudoa thyrsites* (Myxozoa: Myxosporea); specifically a carbohydrate epitope on the surface of the myxospores. This parasite infects pen-reared Atlantic salmon (*Salmo salar*) and coho salmon (*Oncorhynchus kisutch)* in British Columbia. However, the parasite is not restricted to salmonids and is able to infect the myocytes of many species of marine fish. Although *K. thyrsites* rarely results in fish morbidity, it has caused significant economic impact to the fish farming industry due to post-mortem myoliquefaction, or 'soft-flesh' syndrome, which lowers the market value of affected fish. There is an inverse correlation between the numbers of *Kudoa* spores and flesh quality in farmed Atlantic salmon (2). The Cedarlane monoclonal antibody has been used in dot-blot assays to detect *Kudoa thrysites* spores in infected fish tissue (3).

Applications: This antibody has also been used in western blotting, indirect immunofluorescence, flow cytometry and ELISA.

PRESENTATION: 250 µg, purified Ig fraction in PBS + 0.1% NaN_3.

STORAGE/STABILITY:

Store at 4°C. For long term storage, aliquot and freeze unused portions at -20°C in volumes appropriate for single usage. Avoid repeated freeze/thaw cycles.

SPECIFICATIONS:

Clone: IPA-2F4

Continued overleaf…

For more information or to place an order please contact...

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website: www.cedarlanelabs.com • e-mail: info@cedarlanelabs.com

Hybridoma Production:

Immunization:	Percoll™-purified *K. thyrsites* spores
Donor:	BALB/c Draining lymph nodes
Fusion Partner:	Mouse myeloma cell line X63-Ag8.6.5.3

Specificity: A carbohydrate epitope on the surface of *Kudoa thyrsites* myxospores

Ig Class: Mouse IgM

Format: Purified IgM in PBS + 0.1% NaN_3

Antibody Concentration: 1 mg/ml

CELL PREPARATION AND IMMUNOFLUORESENCE ANALYSIS:

Example:

1. Prepare somatic and opercular muscle samples (8 samples/fish, 100mg each).
2. Combine all 8 samples and homogenize in 4.8 ml dH_2O.
3. Pipette 300 ul of the cell suspension onto 500ul of 30% Percoll in dH_2O in a microcentrifuge tube.
4. Centrifuge at 15 800 x g for 1 minute at room temperature.
5. Remove all but 20 ul of the supernatant.
6. Resuspend pelleted spores by tapping the tube.
7. Spot 5 ul of the spore suspension onto membrane and dry.
8. Process for immunodetection by enhanced chemiluminescense.

Dilutions:

Use a 1:500 dilution for IFA. Lower or higher dilutions may be necessary for other applications.

NOTE: For optimal results in various applications, it is recommended that each investigator determine dilutions appropriate for individual use.

REFERENCES:

1. Chase, J.C., J.A. Dawson-Coates, J.D. Haddow, M.H. Stewart, L.R.Haines, D.J., Whitaker, M.L. Kent, R.W. Olafson, and T.W. Pearson. 2001. Analysis of *K. thyrsites* (Myxozoa: Myxosporea) spore antigens using monoclonal antibodies. Diseases of Aquatic Organisms 45: 121-129

2. Dawson Coates, J.A., Chase, J.C., Funk, V., Booy, M.H., Haines, L.R., Falkenberg, C.L., Whitaker, D. J., Olafson, R.W., and Pearson, T.W. 2003. The relationship between flesh quality and numbers of *Kudoa thyrsites* plasmodia and spores in farmed Atlantic salmon (*Salmo salar* L.). Journal of Fish Diseases **26**, 1-10.

3. Chase, J.C., M.H. Booy, J.A. Dawson-Coates, J.D. Haddow, L.R. Haines, D.J. Whitaker, R.W. Olafson, and T.W. Pearson. 2003. Immunological detection of *Kudoa thyrsite* spores in muscle tissues of farmed Atlantic Salmon, *Salmo salar L.* Journal of Fish Diseases 26, 427-431

Manufactured by Vancouver Island Antibodies Ltd. and Immuno-Precise Antibodies for Cedarlane Laboratories Limited

FOR RESEARCH USE ONLY

EJ/09/03