



# SZABO SCANDIC

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- Expressversand

### SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

[mail@szabo-scandic.com](mailto:mail@szabo-scandic.com)

[www.szabo-scandic.com](http://www.szabo-scandic.com)

[linkedin.com/company/szaboscandic](https://www.linkedin.com/company/szaboscandic) 

# TECHNICALLY *Speaking*

Place your order with CEDARLANE® or your local distributor.  
Please contact CEDARLANE® for lot specific information.

## STEINER

CLSG80173

Lot:

### Kit Components included:

100ml.	OXIDIZER SOLUTION	100ml.	10% ZINC FORMALIN
100ml.	0.1% SILVER NITRATE	125ml.	2.5% GUM MASTIC
1.5gm.	HYDROQUINONE	5ml.	1% SILVER NITRATE

**PRINCIPLE:** This kit demonstrates Fungi, Helicobacter pylori, Legionella pneumophila, and Spirochetes.

### Steiner Stain Kit Procedure

**SPECIMEN:** Any well fixed paraffin embedded tissue cut at 3 to 4 microns.

**QUALITY CONTROL:** Recommended Control Slide: Fungus, Helicobacter pylori, Legionnaires, Spirochete.

### SOLUTION:

#### REDUCING SOLUTION: Prepare at Step #10

1% Hydroquinone (Make Fresh) 25ml  
2.5% Gum Mastic 15ml  
1% Silver Nitrate 6 drops

Prepare by adding 0.25 gm **Hydroquinone** to 25 ml Distilled water and mix until completely dissolved. Add 15ml of **2.5% Gum Mastic**, mix thoroughly, and filter solution through medium filter paper. Add 6 drops of **1% Silver Nitrate** and mix thoroughly.

### PROCEDURE:

1. Preheat water bath to 70°C.
2. Deparaffinize slide using Xylene or a Xylene Substitute and hydrate through alcohols to Tap water.
3. Place slide in **OXIDIZER SOLUTION** for 20 minutes.
4. Rinse slide through 2 changes of Distilled water.
5. Place slide in **10% ZINC FORMALIN** for 5 minutes.  
(At this time place 20 ml of **0.1% SILVER NITRATE** in the water bath.)
6. Rinse slide through 2 changes of Distilled water.
7. Place slide in **0.1% SILVER NITRATE** for 5 minutes at 70°C; discard solution after this step.

*Continued overleaf...*

For more information or to place an order please contact...

**CEDARLANE®**  
**LABORATORIES LIMITED**



**toll free: 1-800-268-5058**  
**in North America**

phone: (905) 878-8891 • fax: (905) 878-7800

5516 - 8th Line, R.R.#2, Hornby, Ontario, CANADA L0P 1E0

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8. Rinse slide through 2 changes of Distilled water.
9. Dehydrate slide through 1 change of fresh Absolute Alcohol.
12. Place slide in **2.5% GUM MASTIC** for 3 minutes. (*At this time, prepare **Reducing Solution** and place in the water bath.*)
13. Air dry slide for 1 minute or until the gum mastic is completely dry.
14. Place slide in **Reducing Solution** at 70° C for 10 to 15 minutes or until section is tan to brown; discard **Reducing Solution** after this step.
15. Dehydrate slide through 3 changes of fresh Absolute Alcohol.
16. Clear slide through 3 changes of fresh Xylene or Xylene Substitute.
17. Coverslip slide immediately using a permanent mounting media.

**RESULTS:**

Cat Scratch Fever, Legionella pneumophila,	<b>DARK BROWN, GRAY, OR</b>
Spirochetes, Helicobacter pylori, and Fungi:	<b>BLACK</b>
Background:	<b>YELLOW, TAN, OR BROWN</b>

**REFERENCE:** Elias JM, Green C: Modified Steiner method for the demonstration of Spirochetes in tissue, Am J Clin Pathol; 1979, 71: 109 - 111.

**STEINER STAIN KIT - MICROWAVE PROCEDURE**

**PRINCIPLE:** This kit demonstrates Fungi, Helicobacter pylori, Legionella pneumophila and Spirochetes.

**SPECIMEN:** Any well fixed paraffin embedded tissue cut at 3 to 4 microns.

**QUALITY CONTROL:** Recommended Control Slide: Fungus, Helicobacter pylori, Legonnairs, Spirochete.

**SOLUTION:**

**REDUCING SOLUTION: Prepare at Step #10**

<b>1% Hydroquinone</b> (Make Fresh)	25ml
<b>2.5% GUM MASTIC</b>	15ml
<b>1% SILVER NITRATE</b>	6 drops

Prepare by adding 0.25gm **HYDROQUINONE** to 25ml Distilled water and mix until completely dissolved. Add 15ml of **2.5% GUM MASTIC**, mix thoroughly, and filter solution through medium filter paper. Add 6 drops of **1% SLVER NITRATE** and mix thoroughly.

**PROCEDURE:**

1. Deparaffinize slide using Xylene or a Xylene Substitute and hydrate through alcohols to Tap water.
2. Place slide in **OXIDIZER SOLUTION** for 20 minutes.
3. Rinse through two changes of Distilled water.
4. Place slide in **10% ZINC FORMALIN** for 5 minutes.
5. Rinse slide through two changes of Distilled water and hold in Distilled water until Step #7.
6. In a loosely capped Slide Jar (*available from A.M.T.S.*) heat 20 ml of **0.1% SILVER NITRATE** in a microwave oven for 10 seconds at full power. Repeat as needed until solution is hot, but do not allow solution to boil. Remove Slide Jar from microwave, tighten Slide Jar cap and agitate to equalize solution temperature.
7. Place slide into the hot **0.1% SILVER NITRATE**, agitate thoroughly and incubate for 2 minutes.
8. Rinse slide through 2 changes of Distilled water.
9. Dehydrate slide through 2 changes of fresh Absolute Alcohol.
10. Place slide in **2.5% GUM MASTIC** for 3 minutes. (*During this step, prepare **Reducing Solution**.*)
11. Air dry slide for 1 minute or until the gum mastic is completely dry.
12. In a loosely capped Slide Jar heat 20 ml of **Reducing Solution** in a microwave oven for 25 seconds at full power. Tightly cap Slide Jar, invert several times and place slide into the **Reducing Solution**. Loosely cap Slide Jar, return Slide Jar to the microwave and heat at full power until the **Reducing Solution** is hot, but do not allow solution to boil. Incubate slide for 3 minutes, and then heat again at full power until the **Reducing Solution** is hot. Incubate slide for 2 to 3 minutes; section color should be tan to light brown.
13. Rinse slide quickly in Distilled water.
14. Dehydrate slide through 3 changes of fresh Absolute Alcohol.
15. Clear slide through 3 changes of fresh Xylene or Xylene Substitute.
16. Coverslip slide immediately using a permanent mounting media.

*Continued overleaf...*

**RESULTS:**

Cat Scratch Fever, Legionella pneumophila,  
Spirochetes, Helicobacter pylori, and Fungi:  
Background:

**DARK BROWN, GRAY, OR BLACK**

**YELLOW, TAN, OR BROWN**

**REFERENCE:** Elias JM, Green C: Modified Steiner method for the demonstration of Spirochetes in tissue, Am J Clin Pathol; 1979, 71: 109 - 111.

**For Research Use Only. Not for Diagnostic or Therapeutic Work.**

JV 04/11/06