

Produktinformation



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for the Science of Tomorrow[™]

Picro-Sirius Red Stain Kit (For Collagen)

CLSG80499

Summary and Explanation:

The Picro-Sirius Red Stain Kit (For Collagen) is intended for use in the histological visualization of collagen I and III fibers in addition to muscle in tissue sections. The PSR stain may be viewed using standard light microscopy or polarized light resulting in birefringence of the collagen fibers to distinguish between type I and type III.

Light Microscopy:

Collagen: Red Muscle Fibers: Yellow Cytoplasm: Yellow

Polarized Light Microscopy:

Type I (Thick fibers) Yellow-Orange Birefringence Type III (Thin fibers) Green Birefringence

Control Tissue:

Tissues fixed in 10% formalin are suitable for use prior to paraffin embedding. Consult references (Kiernan, 1981: Sheehan & Hrapchak, 1980) for further details on specimen preparation.

- 1. Cut sections, usually 3 to 5 µm and pick the sections up on glass slides.
- 2. Bake the slides for at least 30 minutes at approximately 70°C.
- 3. Allow to cool.

Recommended Positive Control:

- 1. Lung
- 2. Uterus
- 3. Muscle
- 4. Kidney

Reagents Provided:

Kit Contents	Volume	Storage
Picro-Sirius Red Solution	250 mL	15-30°C
Acetic Acid Solution	250 mL X 2	15-30°C

Visit our website for your local distributor.



In CANADA: Toll Free: 1-800-268-5058

4410 Paletta Court, Burlington, ON L7L 5R2 ph: (289) 288-0001, fax: (289) 288-0020 e-mail: general@cedarlanelabs.com

In the USA: Toll Free: 1-800-721-1644

An ISO 9001:2000 and ISO 13485:2003 registered company.

1210 Turrentine Street, Burlington, NC 27215 ph: (336) 513-5135, fax: (336) 513-5138 e-mail: service@cedarlanelabs.com

Storage and Handling:

Do not use product after the expiration date printed on vial. If reagents are stored under conditions other than those specified here, they must be verified by the user. Diluted reagents should be used promptly.

Staining Procedure:

- 1. Deparaffinize sections if necessary and hydrate to distilled water.
- 2. Apply adequate Picro-Sirius Red Solution to completely cover tissue section and incubate for 60 minutes.
- 3. Rinse slide quickly in two changes of Acetic Acid Solution (0.5%).
- 4. Rinse slide using absolute alcohol.
- 5. Dehydrate in 2 changes of absolute alcohol, clear, and mount in synthetic resin.

Limitations of the Procedure:

- 1. Histological staining is a multiple step diagnostic process that requires specialized training in the selection of the appropriate reagents, tissue selections, fixation, processing, preparation of the slide, and interpretation of the staining results.
- 2. Tissue staining is dependent on the handling and processing of the tissue prior to staining.
- 3. Improper fixation, freezing, thawing, washing, drying, heating, sectioning, or contamination with other tissues or fluids may produce artifacts or false negative results.
- 4. The clinical interpretation of any positive staining, or its absence, must be evaluated within the context of clinical history, morphology and other histopathological criteria. It is the responsibility of a qualified pathologist to be familiar with the special stain and methods used to produce the slide.
- 5. Staining must be performed in a certified licensed laboratory under the supervision of a pathologist who is responsible for reviewing the stained slides and assuring the adequacy of positive and negative controls.

References:

- I. Puchtler H., Waldrop F.S., Valentine L.S. Polarization microscopic studies of connective tissue stained with picro-sirius red FBA. Beitr Path. 1973; 150, pages 174-187.
- II. Junqueira L.C.U., Bignolas G., Brentani R.R. Picro-sirius staining plus polarization microscopy, a specific method for collagen detection in tissue sections. Histochemistry J. 1979, 11, pages 447-455.
- III. Whittaker P. Polarized light microscopy in biomedical research. Microscopy and Analysis 1995; 44, pages 15-17.

Laboratory Reagent For Research Use Only

JK 07/06/18