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- Trockeneiszuschlag
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- Expressversand

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**(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)**

## Annexin V-AF488 / 7-AAD Apoptosis Detection Kit

**Catalog No:** E-CK-A238

**Sizes:** 20 Tests / 50 Tests / 100 Tests / 200 Tests

Cat.	Products	20 Tests	50 Tests	100 Tests	200 Tests	Storage
E-CK-A111	Annexin V-AF488 Reagent	100 µL	250 µL	500 µL	1 mL	2~8 °C
E-CK-A151	Annexin V Binding Buffer (10 ×)	1.4 mL×2	5.5 mL	11 mL	11 mL×2	2~8 °C
E-CK-A162	7-AAD Viability Staining Solution	100 µL	250 µL	500 µL	1 mL	-20 °C
<b>Manual</b>				<b>One Copy</b>		

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Phone: 240-252-7368(USA) Fax: 240-252-7376(USA)

Email: [techsupport@elabscience.com](mailto:techsupport@elabscience.com)

Website: [www.elabscience.com](http://www.elabscience.com)

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

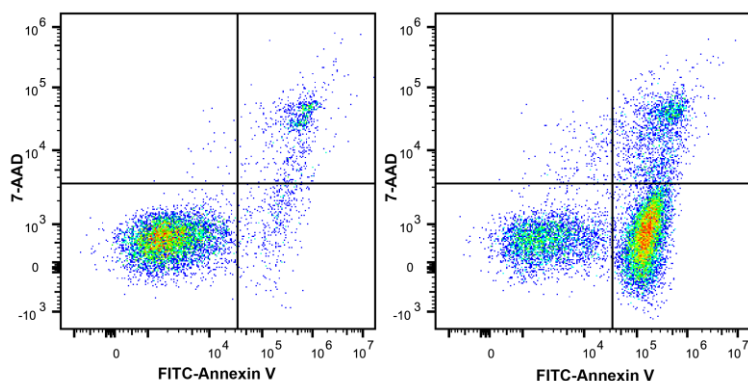
## Introduction

Elabscience® Annexin V-AF488/7-AAD Cell Apoptosis Detection kit is developed to identify apoptotic and necrotic cells.

Annexin V is a member of the annexin family, which binds to phosphatidylserine (PS) in a calcium-dependent manner. Annexin V-AF488, the AF488-conjugated format, binds specifically to the PS on the outer leaflet of apoptotic cell membrane and can be detected with flow cytometry or fluorescence microscopy.

7-AAD (7-amino-actinomycin D) has a high DNA binding constant and is efficiently excluded by intact cells. It is useful for DNA analysis and dead cell discrimination during flow cytometric analysis. Due to the loss of integrity of membrane, 7-AAD can enter late apoptotic or necrotic cells to stain DNA. Cells at different apoptotic stages can be distinguished by using Annexin V and 7-AAD.

*Jurkat cells were treated with 5  $\mu$ M Camptothecin and detected with this kit.*



Jurkat cells were cultured with (Right) or without (Left) 5  $\mu$ M Camptothecin for 4 h. Annexin V-AF488 single-positive cells were early apoptotic cells, Annexin V-AF488 and 7-AAD double-positive cells were necrotic or late apoptotic cells, and 7-AAD single-positive cells were naked nuclei.

## Instructions

The Annexin V Binding Buffer (10  $\times$ ) [E-CK-A151] is a 10  $\times$  concentrated solution. Dilute with DI water to 1  $\times$  working solution before use.

For example: Take 1 mL Annexin V Binding Buffer (10  $\times$ ), dilute with DI water to 10 mL.

## Staining Procedure

### One-step process

1. Induce apoptosis of suspension cells with reagents of interest. Collect cell cultures, centrifuge at 300 g for 5 min and discard the supernatant. Add PBS to wash the cells and resuspend the cells gently followed by the cell counting.

**Tip:** This product is only validated in suspension cells. Good cell viability is the key to the experiment. When the adherent cells are used for apoptotic detection, treatments like digestion may increase the ratio of necrotic or apoptotic cells and cause uncontrollable effects on the experimental results. Please be aware!

2. Split the cell suspension into tubes, 1~5  $\times 10^5$  cells for each, centrifuge at 300 g for 5 min and discard the supernatant. Add PBS to wash the cells and discard the supernatant. Add 500  $\mu$ L of 1  $\times$  Annexin V Binding Buffer to resuspend the cells.
3. Add 5  $\mu$ L of Annexin V-AF488 and 5  $\mu$ L of 7-AAD Viability Staining Solution to each tube.

4. Gently vortex the cells and incubate at room temperature for 15~20 min in the dark.
5. Analyze the cells immediately with proper machine settings. Otherwise, place the cells on ice in the dark and analyze within 1 h.

### Two-step process

1. Induce apoptosis of suspension cells with reagents of interest. Collect cell cultures, centrifuge at 300 g for 5 min and discard the supernatant. Add PBS to wash the cells and resuspend the cells gently followed by the cell counting.

**Tip: This product is only validated in suspension cells. Good cell viability is the key to the experiment. When the adherent cells are used for apoptotic detection, treatments like digestion may increase the ratio of necrotic or apoptotic cells and cause uncontrollable effects on the experimental results. Please be aware !**

2. Split the cell suspension into tubes,  $1\sim5 \times 10^5$  cells for each, centrifuge at 300 g for 5 min and discard the supernatant. Add PBS to wash the cells and discard the supernatant. Add 100  $\mu\text{L}$  of 1  $\times$  Annexin V Binding Buffer to resuspend the cells.
3. Add 2.5  $\mu\text{L}$  of Annexin V-AF488 and 2.5  $\mu\text{L}$  of 7-AAD Viability Staining Solution to each tube. (Attributed to the higher resolution of two-step protocol, half the amount of the reagents can still guarantee a result of matched quality as in the one-step protocol. It's also recommended that users titrate the reagents for optimal performance in specific models.)
4. Gently vortex the cells and incubate at room temperature for 15~20 min in the dark.
5. Add 400  $\mu\text{L}$  of 1  $\times$  Annexin V Binding Buffer to the tube, and mix gently.
6. Analyze the cells immediately with proper machine settings. Otherwise, place the cells on ice in the dark and analyze within 1 h.

### Storage

Annexin V-AF488 and 7-AAD Viability Staining Solution should be stored in dark.

7-AAD Viability Staining Solution should be split into small tubes and stored at  $-20\text{ }^\circ\text{C}$ .

Other reagents should be stored at  $2\sim8\text{ }^\circ\text{C}$ .

### Cautions

1. For maximal assay performance, this kit should be used within 12 months. Avoid freeze / thaw cycles.
2. For FCM analysis, please set untreated cells stained with both Annexin V-AF488 and 7-AAD as negative control. As for compensation controls, please use drug-treated cells stained with either Annexin V-AF488 or 7-AAD.
3. Annexin V-AF488 can be detected in FITC channel while 7-AAD can be detected in PerCP/Cy5.5 channel.
4. Detect apoptosis as soon as possible after staining to avoid the increase number of apoptosis or necrosis.
5. Avoid extended exposure of the samples to direct light to protect the fluorophores from quenching.
6. For your safety and health, please wear the lab coat and disposable gloves before the experiments.