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Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

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(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)

Immuno Fluorescence Staining Kit (Anti-Rabbit IgG-AF594)

Catalog No: E-IR-R327

Size: 50 T / 100 T / 200 T

Cat.	Product Name	50 T	100 T	200 T	Storage
E-AB-1060	Goat Anti-Rabbit IgG (H+L) (AF594 conjugated)	120 µL	200 µL	200 µL×2	-20 ℃
E-IR-R110	Normal Goat Serum (Ready-to-Use)	5 mL	10 mL	20 mL	2~8 ℃
E-IR-R103	DAPI Reagent	5 mL	10 mL	20 mL	-20 ℃
E-IR-R119	Anti-Fluorescence Quenching Agent	5 mL	10 mL	20 mL	2~8 ℃
Manual				One copy	

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Phone: 240-252-7368(USA) 240-252-7376(USA)

Email: techsupport@elabscience.com

Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

Introduction

Immuno Fluorescence Staining Kits are developed for immunofluorescence detection of cell or tissue sections. When there is an appropriate antibody to detect specific target protein, fluorescence can be detected by immunofluorescence staining kit.

Immuno Fluorescence Staining Kit (Anti-Rabbit IgG-AF594) contains Goat Anti-Rabbit IgG(H+L)(AF594 conjugated), this secondary antibody can detect primary antibody from rabbit with bright red fluorescence.

The kit contains anti fluorescence quenching sealing solution, which can make the fluorescence more lasting.

Experimental Procedure

1. Preparation of Immunofluorescence Staining

A. Preparation of Fixation Solution

It is recommended to use 4% Paraformaldehyde as the fixation solution (E-IR-R113), or use ethanol, methanol or other types of fixative according to specific primary antibody or sample.

B. Preparation of TBST Working Buffer

It is recommended to use TBST as washing Buffer. Use Elabscience ® 10×TBST (E-BC-R335) and dilute to 1 ×TBST Working Buffer with deionized water at ratio of 1:9.

C. Preparation of Antibody Dilution Solution

It is recommended to use Elabscience ® Antibody Dilution Buffer (E-IR-R106) or PBS as primary antibody dilution Buffer.

D. Dilute Primary Antibody

Dilute the primary antibody according to the manual of primary antibody.

E. Dilute the Secondary Antibody

It is recommended to use Elabscience ® Antibody Dilution Buffer (E-IR-R106) or PBS as secondary antibody dilution Buffer. Dilute the secondary antibody with antibody dilution Buffer at the dilution of 1:50. The dilution ratio can be increased or decreased appropriately according to the intensity of fluorescence.

2. Immuno Fluorescence Staining for Adherent Cells

A. Immerse a clean cover glass into 70% ethanol for 5 min or a longer time, dry it in the sterile super clean table or wash it with cell culture grade PBS or 0.9% NaCl solution for 3 times, then wash it with cell culture solution. Place the cover glass into six hole plate, seed the cells on the glass overnight to make it about 50% - 80% full.

B. Treat the cells according to the specific experimental purpose, then discard the culture solution, add 1 ml fixation solution, and incubate for 10 min or a longer time (or overnight at 4 °C).

C. Discard the fixation solution, wash the glass with TBST working Buffer for 3~5 min, 3 times. Discard the liquid.

D. Blocking. Add 100 µL Normal Goat Serum (Ready-to-Use) (E-IR-R110) to each glass and incubate for 30 min.

E. Discard the blocking Buffer, add 100 µL diluted primary antibody and incubate at 37 °C in wet box for

60 min (Or incubate at 4 °C overnight).

- F. Discard the primary antibody, wash the glass with TBST Working Buffer for 3~5 min, 3~5 times.
- G. Discard the liquid. Add 100 µL diluted secondary antibody and incubate at 37°C in wet box for 60 min.
- H. Wash the glass with TBST Working Buffer for 3~5 min, 3~5 times, avoid light during the washing.
- I. Nuclear staining. Add DAPI Reagent (E-IR-R103) and incubate in wet box for 5 min, wash the glass with TBST Working Buffer for 5 min, wash for 4 times to remove the redundant DAPI Reagent.
- J. Add one drop of Anti-Fluorescence Quenching Agent (E-IR-R119) on a slice glass, cover the glass with cells, avoid air bubbles. Make the cell contact with the Anti-Fluorescence Quenching Agent, do not reverse it.
- K. Observe the result by fluorescence microscope.

3. Immuno Fluorescence Staining for Suspension Cells

- A. Collect the cells by centrifugation into a 1.5 ml centrifuge tube. Scatter the cells gently after discarding the supernatant.
- B. Add 0.5 ml fixation solution to re-suspend the cells gently, and incubate for 10 min or a longer time (or overnight at 4 °C).
- C. Centrifuge the cells and discard the fixation solution, wash the cells with TBST Working Buffer for 3~5 min, 3 times.
- D. Discard the TBST working Buffer and left about 50 µL liquid for the last time, re-suspend the cells, add the cells to a clean slide glass, make sure that the cells are uniform distributed.
- E. Slightly dry the cells to make the cells are attached to the slide and not easy to flow with the liquid. If the conditions permit, the cells can be attached to the slide by centrifugation using a suitable centrifuge.
- F. Blocking. Add 100 µL Normal Goat Serum (Ready-to-Use) (E-IR-R110) to each glass and incubate for 30 min.
- G. Discard the blocking Buffer, add 100 µL diluted primary antibody and incubate at 37°C in wet box for 60 min (Or incubate at 4 °C overnight).
- H. Discard the primary antibody, wash the glass with TBST working Buffer for 3~5 min, 3~5 times.
- I. Discard the liquid. Add 100 µL diluted secondary antibody and incubate at 37 °C in wet box for 60 min.
- J. Wash the glass with TBST Working Buffer for 3~5 min, 3~5 times, avoid light during the washing.
- K. Nuclear staining. Add DAPI Reagent (E-IR-R103) and incubate in wet box for 5 min, wash the glass with TBST Working Buffer for 5 min, wash for 4 times to remove the redundant DAPI Reagent.
- L. Add one drop of Anti-Fluorescence Quenching Agent (E-IR-R119) on a slice glass, cover the glass with cells, avoid air bubbles. Make the cell contact with the Anti-Fluorescence Quenching Agent, do not reverse it.
- M. Observe the result by fluorescence microscope.

4. Immuno Fluorescence Staining for Tissue Slice

- A. For paraffin section, dewaxing, hydration and antigen repair should be completed first. For frozen sections, follow the steps below.
- B. Incubate with fixation solution for 10 min or a longer time (or overnight at 4 °C).

- C. Discard the fixation solution, wash the slice with TBST Working Buffer for 3~5 min, 3 times. Discard the liquid.
- D. Blocking. Add 100 μ L Normal Goat Serum (Ready-to-Use)(E-IR-R110) to each glass and incubate for 30 min.
- E. Discard the blocking Buffer, add 100 μ L diluted primary antibody and incubate at 37°C in wet box for 60 min (Or incubate at 4 °C overnight).
- F. Discard the primary antibody, wash the glass with TBST Working Buffer for 3~5 min, 3~5 times.
- G. Discard the liquid. Add 100 μ L diluted secondary antibody and incubate at 37°C in wet box for 60 min.
- H. Wash the glass with TBST Working Buffer for 3~5 min, 3~5 times, avoid light during the washing.
- I. Nuclear staining. Add DAPI Reagent (E-IR-R103) and incubate in wet box for 5 min, wash the glass with TBST Working Buffer for 5 min, wash for 4 times to remove the redundant DAPI Reagent.
- J. Add one drop of Anti-Fluorescence Quenching Agent (E-IR-R119) on a slide glass, cover the glass with cells, avoid air bubbles. Make the cell contact with the Anti-Fluorescence Quenching Agent, do not reverse it.
- K. Observe the result by fluorescence microscope.

Storage

Store at 2~8/-20 °C, Refer to the label. Valid for 12 months. Secondary antibody (E-AB-1060) and DAPI Reagent (E-IR-R103) should be stored at -20 °C and avoid light.

Notice

1. This result should be observed by fluorescence microscope.
2. Anti-Fluorescence Quenching Reagent can slow down the quenching, but avoid light and especially shorten the result observation time is still needed.
3. If it can't be observed in time, please store the slice at 4 °C and avoid light and observe in one week.
4. If the fluorescence is too weak, increase the primary antibody concentration properly. If the fluorescence is still weak, increase the secondary antibody concentration appropriately.
5. The reagents for immunofluorescence staining and the cover glass and slides should be prepared in advance.
6. For your safety and health, please wear the lab coat and disposable gloves before the experiments.