



# SZABO SCANDIC

Part of Europa Biosite

## Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

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### Lieferung & Zahlungsart

siehe unsere [Liefer- und Versandbedingungen](#)

### Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

### SZABO-SCANDIC HandelsgmbH

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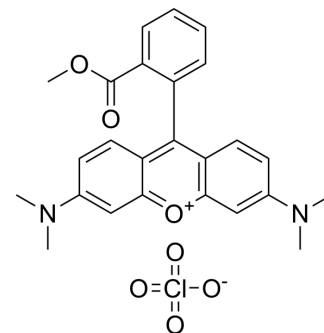
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## TMRM Perchlorate

<b>Cat. No.:</b>	HY-D0984A
<b>CAS No.:</b>	115532-50-8
<b>Molecular Formula:</b>	C <sub>25</sub> H <sub>25</sub> ClN <sub>2</sub> O <sub>7</sub>
<b>Molecular Weight:</b>	500.93
<b>Target:</b>	Fluorescent Dye
<b>Pathway:</b>	Others
<b>Storage:</b>	4°C, sealed storage, away from moisture and light * In solvent : -80°C, 2 years; -20°C, 1 year (sealed storage, away from moisture and light)



### SOLVENT & SOLUBILITY

#### In Vitro

DMSO : 41.67 mg/mL (83.19 mM; Need ultrasonic)

Concentration	Solvent	Mass		
		1 mg	5 mg	10 mg
Preparing Stock Solutions	1 mM	1.9963 mL	9.9814 mL	19.9629 mL
	5 mM	0.3993 mL	1.9963 mL	3.9926 mL
	10 mM	0.1996 mL	0.9981 mL	1.9963 mL

Please refer to the solubility information to select the appropriate solvent.

### BIOLOGICAL ACTIVITY

#### Description

Rhodamine dyes are membrane-permeable cationic fluorescent probes that specifically recognize mitochondrial membrane potentials, thereby attaching to mitochondria and producing bright fluorescence, and at certain concentrations, rhodamine dyes have low toxicity to cells, so they are commonly used to detect mitochondria in animal cells, plant cells, and microorganisms<sup>[1]</sup>.

#### In Vitro

1. Preparation of TMRM working solution
  - 1.1 Preparation of the stock solution  
Dissolve 1 mg TMRM in 525 µL DMSO to obtain 5 mM of stock solution.
  - 1.2 Preparation of TMRM working solution  
Dilute the stock solution in serum-free cell culture medium or PBS to obtain 1-20 µM of working solution.  
Note: Please adjust the concentration of TMRM working solution according to the actual situation.
2. Cell staining
  - 2.1 Suspension cells (6-well plate)
    - a. Centrifuge at 1000 g at 4°C for 3-5 minutes and then discard the supernatant. Wash twice with PBS, 5 minutes each time. The cell density is 1×10<sup>6</sup>/mL.
    - b. Add 1 mL of working solution, and then incubate at room temperature for 5-30 minutes.

c. Centrifuge at 400 g at 4°C for 3-4 minutes and then discard the supernatant.  
d. Wash twice with PBS, 5 minutes each time.  
e. Resuspend cells with serum-free cell culture medium or PBS. Observation by fluorescence microscopy or flow cytometry.

2.2 Adherent cells

a. Culture adherent cells on sterile coverslips.  
b. Remove the coverslip from the medium and aspirate excess medium.  
c. Add 100 µL of working solution, gently shake it to completely cover the cells, and then incubate at room temperature for 30-60 minutes.  
d. Wash twice with medium, 5 minutes each time. Observation by fluorescence microscopy or flow cytometry.

Note: If detection by flow cytometry, cells need to be resuspended before staining.  
MCE has not independently confirmed the accuracy of these methods. They are for reference only.

## PROTOCOL

### Cell Assay <sup>[1]</sup>

Cultures are exposed to Millipore-filtered solutions (0.22 µm) containing TMRM Perchlorate for 1 hr at 37°C (except the experiment involving different durations of exposure to TMRM Perchlorate). After treatment, solutions are removed and growth media reapplied under sterile conditions, and cultures are post-incubated for 18 hours at 37°C (except for the experiment involving analysis at different time points after exposure). Cells are then stained with 2 mg/mL bisbenzimidazole for 20 min at room temperature. Coverslips are subsequently washed in saline and imaged using 2P microscopy. Apoptotic cells are identified as brightly fluorescent nuclei under UV excitation indicating DNA fragmentation. Cell survivability is calculated as the percentage of live, unstained cells (±SD) in five microscopic fields per treatment<sup>[1]</sup>.

MCE has not independently confirmed the accuracy of these methods. They are for reference only.

## CUSTOMER VALIDATION

- Adv Mater. 2023 Mar 29;e2211609.
- Adv Sci (Weinh). 2023 Jan 15;e2203869.
- Autophagy. 2021 Nov;17(11):3592-3606.
- J Thromb Haemost. 2021 Aug 19.
- Basic Res Cardiol. 2024 Jan 2.

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## REFERENCES

- [1]. Crowley LC, et al. Measuring Mitochondrial Transmembrane Potential by TMRE Staining. Cold Spring Harb Protoc. 2016 Dec 1;2016(12):pdb.prot087361.
- [2]. Chowdhury SR, et al. Simultaneous evaluation of substrate-dependent oxygen consumption rates and mitochondrial membrane potential by TMRM and safranin in cortical mitochondria. Biosci Rep. 2015 Dec 8;36(1):e00286.
- [3]. Monteith A, et al. Imaging of mitochondrial and non-mitochondrial responses in cultured rat hippocampal neurons exposed to micromolar concentrations of TMRM. PLoS One. 2013;8(3):e58059.

**Caution: Product has not been fully validated for medical applications. For research use only.**

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