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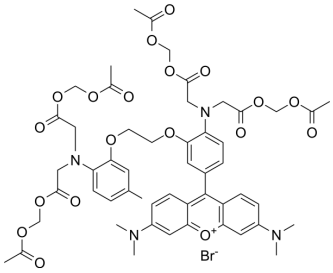
Rhod-2 AM

Cat. No.:	HY-D0989
CAS No.:	145037-81-6
Molecular Formula:	$C_{52}H_{59}BrN_4O_{19}$
Molecular Weight:	1123.94
Target:	Fluorescent Dye
Pathway:	Others
Storage:	-20°C, sealed storage, away from moisture and light * In solvent : -80°C, 6 months; -20°C, 1 month (sealed storage, away from moisture and light)



BIOLOGICAL ACTIVITY

Description

Rhod-2 is a high-affinity visible light excitation wavelength Ca^{2+} fluorescent probe, Rhod-2, AM is an acetyl methyl ester derivative of Rhod-2, which has cell membrane permeability and can easily enter cells with simple culture. Once it enters the cell, it is sheared by its lactesterase to produce Rhod-2 without membrane permeability, which remains in the cell to perform the corresponding physiological functions. Maximum excitation/emission wavelength: 549/578 nm[1].

In Vitro

1.Preparation of Rhod-2 AM working solution
1.1Preparation of the stock solution
Dissolve Rhod-2 AM in DMSO to obtain 5 mM of stock solution.
1.2Preparation of Rhod-2 AM working solution
Dilute the stock solution in serum-free cell culture medium or PBS to obtain 5-10 μM of working solution.
Note: Please adjust the concentration of Rhod-2 AM working solution according to the actual situation.
2.Cell staining (6-well plate)
2.1Suspension cells
a.Centrifuge at 1000 g at 4℃ for 3-5 minutes and then discard the supernatant. Wash twice with PBS, 5 minutes each time.The cell density is 1×10^6/mL.
b.Add 1 mL of working solution, and then incubate at room temperature for 5-30 minutes.
c.Centrifuge at 400 g at 4℃ for 3-4 minutes and then discard the supernatant.
d.Wash twice with PBS, 5 minutes each time.
e.Resuspend cells with serum-free cell culture medium or PBS. Observation by fluorescence microscopy or flow cytometry.
2.2 Adherent cells
a.Culture adherent cells on sterile coverslips.
b.Remove the coverslip from the medium and aspirate excess medium.
c.Add 100 μL of working solution, gently shake it to completely cover the cells,and then incubate at room temperature for 5-30 minutes.
d.Wash twice with medium, 5 minutes each time.Observation by fluorescence microscopy or flow cytometry.
MCE has not independently confirmed the accuracy of these methods. They are for reference only.

PROTOCOL

Cell Assay [1]

For flow cytofluorometry, cells are harvested, pelleted, and resuspended in ice-cold PBS containing 10 mM glucose, 10% fetal bovine serum (FBS), and 10 μM Rhod-2 AM (Rhod2-AM). Mitochondrial calcium levels are determined by the flow

cytofluorometry analysis of aliquots of 4×10^5 cells. For fluorescence microscopy, IMR5 cells are grown on polylysine-coated (10 μg/mL) slides and stained with 7.5 μM Rhod-2 AM in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% FBS for 2 h before poliovirus (PV) infection. Cells are fixed by incubation for 15 min at 4°C in 4% paraformaldehyde. Cells are washed in PBS, and images are acquired with Zeiss Apotome and Axiovision software[1].

MCE has not independently confirmed the accuracy of these methods. They are for reference only.

CUSTOMER VALIDATION

- Environ Sci Technol. 2017 Dec 5;51(23):13938-13948.
- Cell Death Discov. 2021 Feb 10;7(1):31.
- Life Sci. 2024 Feb 14:122505.
- J Agric Food Chem. 2023 May 5.
- Clin Sci. 2023 Dec 7:CS20231039.

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REFERENCES

[1]. Brisac C, et al. Calcium flux between the endoplasmic reticulum and mitochondrion contributes to poliovirus-induced apoptosis. J Virol. 2010 Dec;84(23):12226-35.

[2]. Brisac C, et al. Calcium flux between the endoplasmic reticulum and mitochondrion contributes to poliovirus-induced apoptosis. J Virol. 2010 Dec;84(23):12226-35.

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