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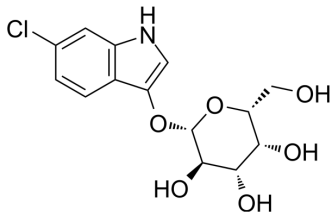
Rose-β-D-Gal

Cat. No.:	HY-W127769
CAS No.:	138182-21-5
Molecular Formula:	$C_{14}H_{16}ClNO_6$
Molecular Weight:	329.73
Target:	Fluorescent Dye
Pathway:	Others
Storage:	Please store the product under the recommended conditions in the Certificate of Analysis.



BIOLOGICAL ACTIVITY

Description	Rose-β-D-Gal is a flurescent dye, is also a β-galactosidase substrate. Rose-β-D-Gal creates a pink/magenta color after the reaction and has been used for detection of β-gal activity[1][2].
In Vitro	Guidelines (Following is our recommended protocol. This protocol only provides a guideline, and should be modified according to your specific needs).Immunostaining of Tissues[1][2]: 1.Prepare 25 mg/mL Rose-β-D-Gal stock solution in Dimethyl Sulfoxide (DMSO) and store at -20 °C. It is vital to protect the stock solution from the light. 2.Prepare rinse buffer for Rose-β-D-Gal staining: 0.1 % Sodium Deoxycholate, 0.2 % IGEPAL CA-630, 2 mM $MgCl_2$ in 0.1 M Na Phosphate buffer (pH 7.3). Prepare 1,000 mL of 0.5 M Na Phosphate buffer (pH 7.3) by mixing with 158 mL of 1 M NaH_2PO_4, 342 mL of 1 M Na $_2HPO_4$, and 500 mL water. 3.Prepare substrate solution for Rose-β-D-Gal fresh, containing 1 mg/mL Rose-β-D-Gal, 5 mM Potassium Ferricyanide, and 5 mM Potassium Ferrocyanide in the rinse buffer. 4.Rehydrate frozen tissue sections in PBS: quickly wash with PBS twice, wash with rinse buffer for 10 min 3 time. 5.Expose to β-galactosidase substrate Rose-β-D-Gal: stain at 37 ⊠ for as long as needed to see stain, up to an overnight time period, keep covered and in the dark during color development. Wash with PBS for 5 min twice. 6.Fix tissue in 4% PFA, unless on intend to continue with in situ hybridizations, and continue to step 4. Note: Rose-β-D-Gal is not very stable in alcohols or organic solvents, so must use water-based counterstains such as Gill's hematoxylin and cover slipping with aqueous mountant. MCE has not independently confirmed the accuracy of these methods. They are for reference only.

REFERENCES

[1]. Ismail J A , et al. Immunohistologic labeling of murine endothelium[J]. 2003, 12(2):0-90.

[2]. Komatsu Y, et al. In situ hybridization methods for mouse whole mounts and tissue sections with and without additional β-galactosidase staining. Methods Mol Biol. 2014;1092:1-15.

Caution: Product has not been fully validated for medical applications. For research use only.

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