



# SZABO SCANDIC

Part of Europa Biosite

## Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

Weitere Information auf den folgenden Seiten!  
See the following pages for more information!



### Lieferung & Zahlungsart

siehe unsere [Liefer- und Versandbedingungen](#)

### Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

### SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

[mail@szabo-scandic.com](mailto:mail@szabo-scandic.com)

[www.szabo-scandic.com](http://www.szabo-scandic.com)

[linkedin.com/company/szaboscandic](https://www.linkedin.com/company/szaboscandic) 

## Datasheet for 100-4159

## Hexokinase Antibody

### Overview

<b>Description:</b>	Anti-Hexokinase (Yeast) (RABBIT) Antibody - 100-4159
<b>Item No.:</b>	100-4159
<b>Size:</b>	2 mL
<b>Applications:</b>	WB
<b>Reactivity:</b>	Hexokinase (Yeast)
<b>Host Species:</b>	Rabbit

### Product Details

<b>Background:</b>	Hexokinase is the main glucose phosphorylating enzyme. It may play a regulatory role in both induction and repression of gene expression by glucose. In yeast there are three glucose-phosphorylating isoenzymes, designated hexokinase I, II and glucokinase. This protein is involved in the pathway hexose metabolism, which is part of Carbohydrate metabolism.
<b>Synonyms:</b>	rabbit anti-Hexokinase Antibody, DKFZp686M1669 antibody, Hexokinase 2 antibody, Hexokinase 2 muscle antibody, Hexokinase type II antibody
<b>Host Species:</b>	Rabbit
<b>Clonality:</b>	Polyclonal
<b>Format:</b>	Antiserum

### Target Details

<b>Gene Name:</b>	HXK2
<b>Reactivity:</b>	Hexokinase (Yeast)
<b>Immunogen Type:</b>	Native Protein
<b>Immunogen:</b>	Hexokinase [Yeast]
<b>Purity/Specificity:</b>	This product was prepared from monospecific antiserum by a delipidation and defibrination. Assay by immunoelectrophoresis resulted in a single precipitin arc against anti-rabbit serum, purified and partially purified Hexokinase [Yeast]. Cross reactivity against Hexokinase from other tissues and species may occur but have not been specifically determined.

<b>Relevant Links:</b>	<ul style="list-style-type: none"><li>• <a href="#">UniProtKB - P04806</a></li><li>• <a href="#">NCBI - CAA96973.1</a></li><li>• <a href="#">UniProtKB - P04807</a></li><li>• <a href="#">GeneID - 852639</a></li></ul>
------------------------	---

## Application Details

<b>Suggested Applications:</b>	WB (Based on references)
<b>Application Note:</b>	Anti-Hexokinase is suitable for use in ELISA, western blot, and immunohistochemistry. Specific conditions for reactivity should be optimized by the end user.
<b>Assay Dilutions:</b>	All assays should be optimized by the user. Recommended dilutions (if any) may be listed below.
<b>ELISA:</b>	1:5,000 - 1:25,000
<b>IHC:</b>	User Optimized
<b>WB:</b>	1:500 - 1:3,000

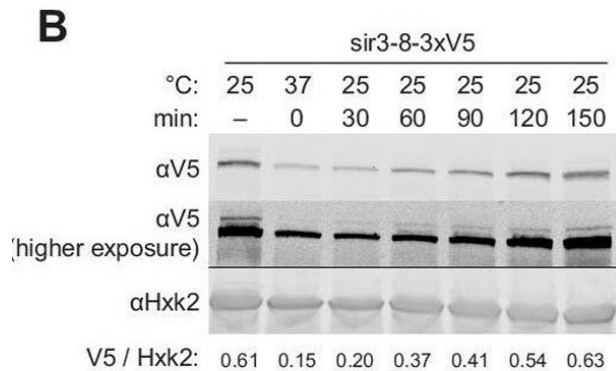
## Formulation

<b>Physical State:</b>	Lyophilized
<b>Concentration:</b>	80 mg/mL by Refractometry
<b>Buffer:</b>	0.02 M Potassium Phosphate, 0.15 M Sodium Chloride, pH 7.2
<b>Preservative:</b>	0.01% (w/v) Sodium Azide
<b>Stabilizer:</b>	None
<b>Reconstitution Volume:</b>	2.0 mL
<b>Reconstitution Buffer:</b>	Restore with deionized water (or equivalent)

## Shipping & Handling

<b>Shipping Condition:</b>	Ambient
<b>Storage Condition:</b>	Store vial at 4° C prior to restoration. For extended storage aliquot contents and freeze at -20° C or below. Avoid cycles of freezing and thawing. Centrifuge product if not completely clear after standing at room temperature. This product is stable for several weeks at 4° C as an undiluted liquid. Dilute only prior to immediate use.
<b>Expiration:</b>	Expiration date is one (1) year from date of receipt.

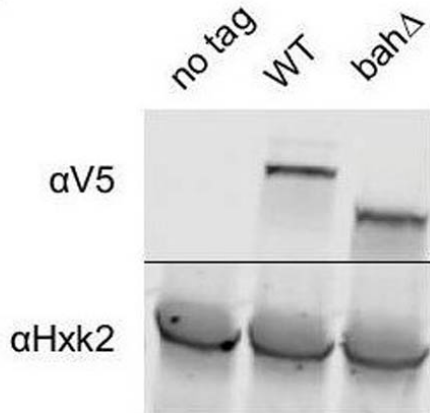
## Images



### Western Blot

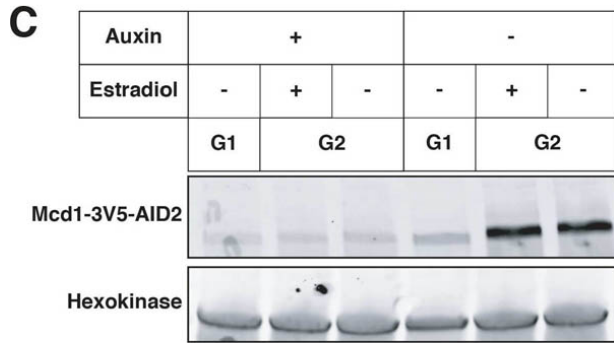
Repression of HML and HMR preceded heterochromatin maturation. (A) Schematic of temperature-shift time course with sir3-8-M.ECOGII. (B) Protein immunoblotting in a strain expressing sir3-8-3xV5 (JRY13467) constitutively at 25°C (first lane), constitutively at 37°C (second lane), and at 30, 60, 90, 120, and 150 min after a shift to 25°C. Top row is 3xV5-tagged sir3-8 protein, the middle row is the same as the top row but at a higher exposure, and the bottom row is the loading control Hxk2. The unedited blot is in . (C) Aggregate methylation results at HML (top) and HMR (bottom) from long-read Nanopore sequencing of strains expressing sir3-8-M.ECOGII (JRY13114) grown constitutively at 25 or 37°C. Plots are as described in Figure 1D. (D) Aggregate methylation results at HML (top) and HMR (bottom) from long-read Nanopore sequencing of a strain expressing sir3-8-M.ECOGII (JRY13134) grown constitutively at 25°C (dotted gray line) and collected at 0, 15, 45, and 90 min after a temperature switch from 37 to 25°C. (E) RT-qPCR of HML±2 (left) and HMRA1 (right) mRNA in strains expressing SIR3-M.ECOGII (black, JRY13027, JRY12840), sir3,Δ::M.ECOGII (green, JRY13029, JRY13030), or sir3-8-M.ECOGII (purple, JRY13114, JRY13134) collected at 0, 30, 60, 90, 120, and 150 min after a temperature switch from 37 to 25°C. Points are the average of three biological replicates and bars mark one standard deviation. DIP-seq of sir3-8-M.ECOGII (JRY13114). Shown are 10 kb regions centered at HML (left) and HMR (right). Cells were grown constitutively at either 25 or 37°C. Input results are plotted but not visible due to the strong DIP-seq signals. Nanopore sequencing over temperature switch time course (biological replicate). Aggregate methylation results at HML (top) and HMR (bottom) from long-read Nanopore sequencing of a strain expressing sir3-8-M.ECOGII (JRY13114) grown constitutively at 25°C (dotted gray line) and collected at 0, 15, 45, and 90 min after a temperature switch from 37 to 25°C. Figure provided by CiteAb. Source: Elife, PMID: 35073254.

**B**



**Western Blot**

Nucleosome binding was required for spread, but not recruitment, of Sir3 to regions of heterochromatin. (A) Schematic of Sir3 protein domains. (B) Protein immunoblotting in strains expressing Sir3 (no tag, JRY11699), Sir3-3xV5 (JRY12601), and sir3-bah,Δ3xV5 (JRY13621). Top row are 3xV5-tagged Sir3 proteins, and bottom row is the loading control Hxk2. The unedited blot is in Figure 2, source data 1. (C) RT-qPCR of HML±2 and HMRa1 mRNA, normalized to ACT1 mRNA, in strains expressing SIR3-M.ECOGII (JRY12840, JRY13027), sir3,Δ::M.ECOGII (JRY13029, JRY13030), and sir3-bah,Δ-M.ECOGII (JRY13438, JRY13439). Data are the average of three biological replicates, and bars mark one standard deviation. (D) Aggregate methylation results at HML (top) and HMR (bottom) from long-read Nanopore sequencing of sir3,Δ::M.ECOGII (JRY13029, JRY13030), SIR3-M.ECOGII (JRY12840, JRY13027), and sir3-bah,Δ-M.ECOGII (JRY13438). Plots are as described in Figure 1D. (E) Single-read plots from long-read Nanopore sequencing of sir3-bah,Δ-M.ECOGII (JRY13438) at HML (top) and HMR (bottom). Plots are as described in Figure 1E. (F) Aggregate methylation results at four representative telomeres (1 L, 2 L, 4 R, and 11 R) from long-read Nanopore sequencing of the same strains as D. Shown are 15 kb windows of each telomere. Plots are as described in Figure 1D. (G) Single-read plots from long-read Nanopore sequencing of SIR3-M.ECOGII (JRY13027) and sir3-bah,Δ-M.ECOGII (JRY13438) at two representative telomeres (1 L and 2 L). Shown are 10 kb windows of each telomere. Figure 2, source data 1. Uncropped protein immunoblot of Sir3 mutants. Uncropped protein immunoblot of Sir3 mutants. DIP-seq of SIR3-M.ECOGII (top row, JRY13027), sir3,Δ::M.ECOGII (middle row, JRY13030), and sir3-bah,Δ-M.ECOGII (bottom row, JRY13438). Input results are plotted but not visible due to the strong DIP-seq signals. (A) Shown are 10 kb regions centered at HML (left) and HMR (right). (B) Shown are 10 kb regions at two representative telomeres (1 L and 2 L). Methylation by Sir3,ΔM.EcoGII and sir3-bah,Δ-M.EcoGII at all 32 telomeres. Aggregate methylation results from long-read Nanopore sequencing of the same strains as Figure 2D. Shown are 15 kb windows of each telomere. Plots are as described in Figure 1D. Highlighted in yellow are windows shorter than 15 kb due to discrepancies between the S288C and W303 genomes (see Ellahi et al., 2015). Figure provided by CiteAb. Source: Elife, PMID: 35073254.



### Western Incubation Box

Effects of cohesin depletion and tT(AGU)C deletion on silencing establishment. (A) SIR3-EBD strains (JRY12269, JRY12270) and SIR3-EBD strains with seamless deletion of tT(AGU)C (JRY12267; JRY12268) were arrested in G1 with  $\text{CE}\pm$  factor, then split, with half the culture receiving estradiol and half receiving ethanol. Samples were collected after 3 hr for RT-qPCR, with each sample normalized to its own pre-estradiol value. (B) Cells with MCD1-AID (JRY12560, JRY12561) were arrested in G1 with  $\text{CE}\pm$  factor, then split, with half receiving auxin and the other half receiving DMSO (solvent control). After 30 min, each culture was further split, with half receiving estradiol and the other half ethanol. All cultures were released to G2/M by addition of protease and nocodazole. Cells were collected after 3 hr for RT-qPCR, with each sample normalized to its own pre-estradiol value. (C) Immunoblot analysis showing Mcd1-AID depletion for experiment described in (B). Figure provided by CiteAb. Source: Elife, PMID: 32687055.

## References

- Ben-Menachem RH et al. Mitochondrial-derived vesicles retain membrane potential and contain a functional ATP synthase. *EMBO Rep.* (2023)
- Saxton DS et al. Distinct silencer states generate epigenetic states of heterochromatin. *Mol Cell.* (2022)
- Brothers, M et al. Distinguishing between recruitment and spread of silent chromatin structures in *Saccharomyces cerevisiae*. *ELife* (2022)
- Otto GM et al. Programmed cortical ER collapse drives selective ER degradation and inheritance in yeast meiosis. *J Cell Biol.* (2021)
- Farris D et al. A novel allele of SIR2 reveals a heritable intermediate state of gene silencing. *Genetics.* (2021)
- Goodnight D et al. S-phase-independent silencing establishment in *Saccharomyces cerevisiae*. *Elife.* (2020)
- Eisenberg AR et al. Translation initiation site profiling reveals widespread synthesis of non-AUG-initiated protein isoforms in yeast. *Cell Syst.* (2020)
- Muller R et al. CiBER-seq dissects genetic networks by quantitative CRISPRi profiling of expression phenotypes. *Science.* (2020)
- Lim G et al. Phosphoregulation of Rad51/Rad52 by CDK1 functions as a molecular switch for cell cycle-specific activation of homologous recombination. *Sci Adv.* (2020)
- Sawyer EM et al. Developmental regulation of an organelle tether coordinates mitochondrial remodeling in meiosis. *J Cell Biol.* (2019)
- King GA et al. Meiotic cellular rejuvenation is coupled to nuclear remodeling in budding yeast. *Elife.* (2019)
- Brothers M et al. Mutations in the PCNA DNA Polymerase Clamp of *Saccharomyces cerevisiae* Reveal Complexities of the Cell Cycle and Ploidy on Heterochromatin Assembly. *Genetics.* (2019)
- Brace JL et al. A cell separation checkpoint that enforces the proper order of late cytokinetic events. *J Cell Biol.* (2019)
- Lam, MHY et al. A Comprehensive Membrane Interactome Mapping of Sho1p Reveals Fps1p as a Novel Key Player in the Regulation of the HOG Pathway in *S. cerevisiae*. *Journal of Molecular Biology* (2015)
- Thanabalu T et al. Verprolin function in endocytosis and actin organization: Roles of the Las17p (yeast WASP)-binding domain and a novel C-terminal actin-binding domain. *FEBS J.* (2007)

## Disclaimer

This product is for research use only and is not intended for therapeutic or diagnostic applications. Please contact a technical service representative for more information. All products of animal origin manufactured by Rockland Immunochemicals are derived from starting materials of North American origin. Collection was performed in United States Department of Agriculture (USDA) inspected facilities and all materials have been inspected and certified to be free of disease and suitable for exportation. All properties listed are typical characteristics and are not specifications. All suggestions and data are offered in good faith but without guarantee as conditions and methods of use of our products are beyond our control. All claims must be made within 30 days following the date of delivery. The prospective user must determine the suitability of our materials before adopting them on a commercial scale. Suggested uses of our products are not recommendations to use our products in violation of any patent or as a license under any patent of Rockland Immunochemicals, Inc. If you require a commercial license to use this material and do not have one, then return this material, unopened to: Rockland Inc., P.O. BOX 5199, Limerick, Pennsylvania, USA.