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- Mindermengenzuschlag
- Trockeneiszuschlag
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- Expressversand

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ImmunoCruz[®] rabbit ABC Staining System: sc-2018

PRODUCT

ImmunoCruz[®] rabbit ABC Staining System includes 1.0 ml normal mouse blocking serum, 250 μ l of 100x mouse anti-rabbit IgG-Biotin secondary antibody, 0.5 ml each avidin and biotinylated horseradish peroxidase (AB reagents), 1.0 ml 50x peroxidase substrate, 1.0 ml 50x DAB chromogen and 3.0 ml 15x substrate buffer. Also included are mixing bottles for the preparation of reagent working solutions. One ImmunoCruz[®] ABC Staining System contains sufficient reagent for 200 slides.

Solutions to be provided by the researcher are phosphate buffered saline (PBS) prepared in glass distilled H₂O; 0.1–1% hydrogen peroxide (H₂O₂) diluted in PBS, distilled H₂O or methanol (optional); rabbit primary antibody; any reagents needed to fix and/or deparaffinize specimens on slides; counterstain (optional); and mounting medium.

PREPARATION OF WORKING SOLUTIONS

Use only freshly prepared buffers. Prepare all working solutions in the mixing bottles provided. After preparation, insert the drop dispenser top (supplied in inverted position) into the cap in correct orientation. Place the entire unit on the bottle and twist the cap. The drop dispenser will snap into place. To remove the drop dispenser for refilling, press laterally with thumb until the top snaps off. To prevent evaporation, secure the caps on bottles when not in use. After completion of the staining procedure, working solutions should be discarded and mixing bottles washed with distilled H₂O.

- Blocking serum: In mixing bottle 1 (blue cap), combine 30 μ l normal mouse blocking serum stock with 2 ml PBS.
- Biotinylated secondary antibody: In mixing bottle 2 (green cap), combine 30 μ l normal mouse blocking serum stock, 2 ml PBS and 20 μ l biotinylated mouse anti-rabbit IgG secondary antibody stock.
- AB enzyme reagent: In AB mixing bottle (purple cap), combine 40 μ l reagent A (avidin), 40 μ l reagent B (biotinylated HRP) and 2 ml PBS. Mix and let stand for approximately 30 minutes.
- Peroxidase substrate: In substrate mixing bottle (yellow cap), combine 1.7 ml distilled H₂O, 3 drops 15x substrate buffer, 1 drop 50x DAB chromogen and 1 drop 50x peroxidase substrate. Sufficient for 15–20 slides.
- Refer to the Immunoperoxidase Staining Protocol at www.scbt.com for tissue section preparation and additional technique notes.

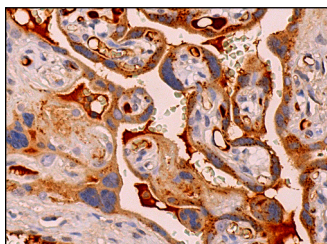
PROCEDURE

All steps are carried out at room temperature in a humidified chamber. Staining dishes or coplin jars may also be used. Apply sufficient volumes of reagents to completely cover the section; 100 μ l is usually adequate, or 1–3 drops of working solutions. Use suction to remove reagents after each step, but avoid drying of specimens between steps.

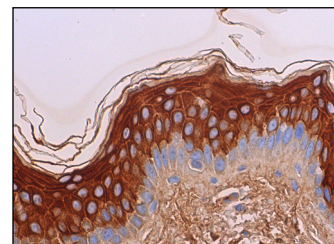
- Optional: After preparation of tissue sections, slides may be incubated for 5–10 minutes in 0.1–1% hydrogen peroxide diluted in PBS, deionized H₂O or methanol to quench endogenous peroxidase activity. Wash in PBS twice for 5 minutes each.

- Optional: Incubate sections for one hour in 1.5% blocking serum in PBS (mixing bottle 1). This step may be omitted if non-specific staining is not a problem. Blot excess blocking serum from slides.
- Incubate sections with rabbit primary antibody for 2 hours at room temperature or overnight at 4° C. Optimal antibody concentration should be determined by titration; recommended range is 0.5–5.0 μ g/ml, diluted in 1.5% mouse blocking serum in PBS (from mixing bottle 1). Wash with three changes of PBS for 5 minutes each.
- Incubate sections for 1.5 hours with biotinylated secondary antibody as prepared in mixing bottle 2. Wash with three changes of PBS for 5 minutes each.
- Incubate sections for 30 minutes with AB enzyme reagent (AB mixing bottle). Wash with three changes of PBS for 5 minutes each.
- Incubate sections in 1–3 drops peroxidase substrate (substrate mixing bottle) for 30 seconds–10 minutes or until desired stain intensity develops. The section may be checked for staining by rinsing with H₂O and viewing under a microscope. If necessary, add additional peroxidase substrate and continue to incubate. Wash sections in deionized H₂O for 5 minutes.
- Optional: Counterstain sections in Gill's formulation #2 hematoxylin for 5–10 seconds. Immediately wash with several changes of deionized H₂O.
- Optional: Destain with acid alcohol and bluing reagent. Wash with tap water.
- For paraffin-embedded tissue sections, dehydrate as follows: 2x 95% ethanol for 10 seconds each, 2x 100% ethanol for 10 seconds each, 3x xylenes for 10 seconds each. Wipe off excess xylenes.
- Immediately add 1–2 drops of permanent mounting medium and cover with a glass coverslip. Observe by light microscopy.

DATA



GDF-15 (FL-308): sc-66904. Immunoperoxidase staining of formalin fixed, paraffin-embedded human placenta tissue showing cytoplasmic staining of trophoblastic cells and decidual cells. Staining system used: ImmunoCruz[®] rabbit ABC Staining System: sc-2018.



PtT1 (H-130): sc-98814. Immunoperoxidase staining of formalin fixed, paraffin-embedded human skin tissue showing cytoplasmic staining of keratinocytes, fibroblasts and Langerhans cells. Staining system used: ImmunoCruz[®] rabbit ABC Staining System: sc-2018.

STORAGE

Store all ABC Staining System components at 2–8° C. ****DO NOT FREEZE.**** Stable for one year from the date of shipment.

RESEARCH USE

For research use only, not for use in diagnostic procedures.