SZABO SCANDIC

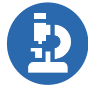
Part of Europa Biosite

Produktinformation

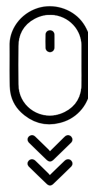
Forschungsprodukte & Biochemikalien

Zellkultur & Verbrauchsmaterial

Diagnostik & molekulare Diagnostik

 Laborgeräte & Service

Weitere Information auf den folgenden Seiten!
See the following pages for more information!



Lieferung & Zahlungsart

siehe unsere Liefer- und Versandbedingungen

Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

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SANTA CRUZ BIOTECHNOLOGY, INC.

ImmunoCruz® goat LSAB Staining System: sc-2053

The Power to Question

PRODUCT

The ImmunoCruz® goat LSAB Staining System includes 15 ml each peroxidase block, serum block (5% normal mouse serum, two 15 ml vials provided), mouse anti-goat IgG-Biotin secondary antibody and Avidin D-HRP reagent. Each of these reagents is pre-diluted and ready to use for immunohistochemical staining of formalin-fixed, paraffin-embedded tissue sections. Also included are 1 ml of 50x peroxidase substrate, 1 ml of 50x DAB chromogen and 3 ml of 15x substrate buffer. Number of slides: 150.

Goat primary antibody and negative control (normal goat IgG) are provided by the researcher. Goat primary antibody should be diluted to 0.5–5 µg/ml, as determined by titration, in serum block (two 15 ml vials provided).

PROCEDURE

ImmunoCruz® LSAB Staining Systems utilize an Avidin D-HRP complex for staining of formalin-fixed, paraffin-embedded tissue sections.

- Clean glass slides with 95% ethanol, treat with subbing solution and air dry. Alternatively, use pre-treated slides.
- Cut 4–6 micron thick tissue sections using microtome and apply to slides. Deparaffinize as follows: 3x xylenes for 5 minutes each, 2x 100% ethanol for 10 minutes each and 2x 95% ethanol for 10 minutes each. Wash in deionized H_2O for 1 minute on stir plate. Aspirate excess liquid from slides.
- Optional: To unmask antigens by heat treatment, place slides in container and cover with 10 mM sodium citrate buffer, pH 6.0. Heat at 95° C for 5 minutes. Top off with fresh buffer and heat at 95° C for 5 minutes (optimal incubation time may vary for each tissue type). Allow slides to cool in buffer for 20 minutes. Wash in deionized H_2O three times for 2 minutes each on stir plate. Aspirate excess liquid from slides.
- All subsequent steps are carried out at room temperature in a humidified chamber. Allow all ImmunoCruz® LSAB Staining System reagents to reach room temperature prior to use. Tissue sections should not be allowed to dry out at any time during the procedure.
- Optional: To quench endogenous peroxidase activity, incubate specimens for 5 minutes in 1–3 drops peroxidase block (white cap). Rinse with PBS and transfer to a PBS wash for 2 minutes on stir plate. Aspirate excess liquid from slides.
- Incubate specimens for 20 minutes in 1–3 drops of serum block (blue cap). Aspirate serum from slides.
- Dilute goat primary antibody to 0.5–5 µg/ml as determined by titration. Antibody should be diluted in mouse serum block (provided). Add diluted goat primary antibody in sufficient volume to cover the tissue.
- Incubate for 2 hours at room temperature, or overnight at 4° C. Rinse with PBS, then wash in PBS twice for 2 minutes each on stir plate. Aspirate excess liquid from slides.
- Incubate specimens for 1.5 hours in 1–3 drops of biotinylated mouse anti-goat IgG secondary antibody (green cap). Rinse with PBS, then wash in PBS twice for 2 minutes each on stir plate. Aspirate excess liquid from slides.
- Incubate specimens for 30 minutes in 1–3 drops of Avidin D-HRP complex (purple cap). Rinse with PBS, then wash in PBS twice for 2 minutes each on stir plate. Aspirate excess liquid from slides.
- During the above incubation step, prepare HRP substrate in the substrate mixing bottle (yellow cap) as follows (sufficient for 15-20 slides): remove tip from mixing bottle and combine 1.7 ml deionized H_2O, 3 drops 15x substrate buffer (orange cap), 1 drop 50x DAB chromogen (yellow cap) and 1 drop 15x peroxidase substrate (yellow cap).
- Add 1–3 drops of HRP substrate to each slide. Develop until light brown staining is visible, usually 30 seconds – 10 minutes, although up to 20 minutes may be required. The section may be checked for staining by rinsing with deionized H_2O and viewing under a microscope. If necessary, add additional HRP substrate and continue to incubate. Rinse with deionized H_2O and transfer to deionized H_2O wash for 2 minutes on stir plate.
- Optional: Counterstain slides in Gill's formulation #2 hematoxylin for 5–10 seconds. Immediately wash with several changes of deionized H_2O.
- Optional: Destain with acid alcohol and bluing reagent. Wash with tap water.
- Dehydrate sections as follows: 2x 95% ethanol for 10 seconds each, 2x 100% ethanol for 10 seconds each and 3x xylenes for 10 seconds each. Wipe off excess xylene from edges of slide.
- Immediately add 1–2 drops of permanent mounting medium and cover with glass coverslip. Observe by light microscopy.

STORAGE

Store at 4° C. **DO NOT FREEZE.** Stable for one year from the date of shipment.

RESEARCH USE

For research use only; not for use in diagnostic procedures.