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Monosan[®] Plus HRP Polymer Kit

REF / Cat. No.: **MON-APP103 60 tests, 6 ml**
MON-APP104 1000 tests, 100 ml

Instructions for use

Intended use

Monosan[®] Plus HRP Polymer Kit is designed for the qualitative detection of antigens in fixed paraffin-embedded tissue sections, in frozen tissue sections, and in cytological samples. It is developed for use in combination with mono- and polyclonal primary antibodies and sera obtained from mice and rabbits.

The kit can be used for examining tissues fixed in different solutions, e.g. formalin (neutrally buffered), B5, Bouin, ethanol, or HOPE. Monosan[®] Plus HRP Polymer Kit is for research use only, not for drug, diagnostic or other use.

Summary and Explanation

The purpose of the immunohistochemical staining is to make tissue and cell antigens visible.

Monosan[®] Plus HRP Polymer Kit is a highly sensitive detection kit intended for use in immunohistochemistry and immunocytochemistry. The enzyme polymer in this kit consists of several molecules of secondary antibodies covalently bound to several molecules of horse radish peroxidase (HRP).

Visualisation occurs via an enzyme-substrate reaction in the presence of a colorising reagent which permits microscopical analysis.

The test system is suitable for the detection of mono- and polyclonal primary antibodies and sera obtained from mice and rabbits. In contrast to other detection techniques, which often use the streptavidin-biotin system the Monosan[®] Plus HRP Polymer Kit avoids the problem of background staining caused by endogenous biotin in the tissue.

Principle of the method

Paraffin-embedded tissue sections are first deparaffinised and rehydrated.

Endogenous peroxidase activity in the tissue may cause non-specific staining. This enzyme activity can be blocked by incubation with 3% H₂O₂-solution (peroxide block).

Background staining caused by unspecific binding of the primary antibody or the secondary antibody in the HRP-polymer is minimized by incubation with a protein blocking solution ("Blocking Solution", provided with the kit). This step can be omitted if the primary antibodies are diluted in an appropriate buffer.

The next step is incubation with the specific primary antibody. After washing, the enhancement reagent ("PostBlock") is applied and incubated. A second washing is followed by the application of the HRP-polymer. Any excess of unbound HRP-polymer is thoroughly washed away after incubation. The addition of the chromogenic substrate starts the enzymatic reaction of the peroxidase which leads to colour precipitation where the primary antibody is bound. The colour can be observed with a light microscope.

The chromogen used determines the colour. The chromogen AEC leads to the formation of a red-brown product of reaction at the place of the target antigen. The chromogen DAB forms a dark brown precipitate.

Reagents provided

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6 ml	Blocking Solution	Reagent 1	(ready-to-use)
6 ml	PostBlock	Reagent 2	(ready-to-use)
6 ml	HRP-Polymer (Mouse/Rabbit)	Reagent 3	(ready-to-use)

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100 ml	Blocking Solution	Reagent 1	(ready-to-use)
100 ml	PostBlock	Reagent 2	(ready-to-use)
100 ml	HRP-Polymer (Mouse/Rabbit)	Reagent 3	(ready-to-use)

Substrate systems recommended (if not included in the kit):

<i>AEC Single Solution</i>	<i>Cat. No. MON-APP169</i>	<i>80 tests</i>
	<i>Cat. No. MON-APP170</i>	<i>1250 tests</i>
<i>AEC Substrate Kit</i>	<i>Cat. No. MON-APP171</i>	<i>500 tests</i>
	<i>Cat. No. MON-APP172</i>	<i>5000 tests</i>
<i>DAB Substrate Kit</i>	<i>Cat. No. MON-APP176</i>	<i>500 tests</i>
	<i>Cat. No. MON-APP177</i>	<i>5000 tests</i>
<i>DAB High Contrast Kit</i>	<i>Cat. No. MON-APP174</i>	<i>500 tests</i>
	<i>Cat. No. MON-APP175</i>	<i>5000 tests</i>

Materials required but not supplied

Positive und negative control tissue
Xylene or suitable substitutes
Ethanol, distilled H₂O
3% H₂O₂ solution (Cat. No. MON-APP146)
Reagents for enzyme digestion or heat pre-treatment
Wash buffer PBS or TBS (Cat. No. MON-APP166)
PAP Pen (Cat. No. MON-APP190)
Primary antibody (user-defined)
Primary antibody diluent (Cat. No. MON-APP149)
Negative control reagent
Chromogenic substrate
Counter stain solution
Mounting medium
Cover slips

Storage and handling

The solutions should be stored at 2-8°C without further dilution. Please store the reagents in a dark place and do not freeze them. Under these conditions the solutions are stable up to the expiry date. They should not be used after the expiry date.

A positive and a negative control have to be carried out in parallel to the test material. If you observe unusual staining or other deviations from the expected results which could possibly be caused by the kit reagents, please contact Monosans' technical support or your local distributor.

Precautions

Use by qualified personnel only.

Wear protective clothing to avoid eye, skin or mucous membrane contact with the reagents. In case of a reagent coming into contact with a sensitive area, wash the area with large amounts of water.

ProClin 300 and sodium azide (NaN₃), used for stabilization, are not considered hazardous materials in the concentrations used. Sodium azide deposits in drainage pipes made of lead or copper can result in the formation of highly explosive metallic azides. To avoid such deposits in drainage pipes, sodium azide should be discarded in a large volume of running water. Material safety data sheets (MSDS) for the pure substances are available upon request.

Microbial contamination of the reagents must be avoided, since otherwise non-specific staining might appear.

Reagent preparation

- Reagents should be at room temperature when used.
- Deparaffinise and rehydrate paraffin-embedded tissue sections.
- Pre-treatment (optional) with HIER (*Heat Induced Epitope Retrieval*) or enzymatic digestion.
- Tissue sections have to be completely covered with the different reagents in order to avoid drying out.

Staining procedure

- | | |
|---|------------|
| 1. Peroxide blocking (3 % H ₂ O ₂ solution) | 10 min. |
| 2. Washing with wash buffer | 1 x 2 min. |
| 3. Blocking Solution (protein block, Reagent 1) (<i>This step is optional.</i>) | 5 min. |
| 4. Washing with wash buffer | 1 x 2 min. |
| 5. Primary antibody (optimally diluted) or negative control reagent | 30-60 min. |
| 6. Washing with wash buffer | 3 x 5 min. |
| 7. PostBlock (Reagent 2, yellow) | 20 min. |
| 8. Washing with wash buffer | 3 x 5 min. |
| 9. HRP-polymer (Reagent 3, red) | 30 min. |
| 10. Washing with wash buffer | 3 x 2 min. |
| 11. AEC or DAB (<i>Controlling the colour intensity via light microscope is recommended.</i>) | 5-15 min. |
| 12. Stopping the reaction with distilled H ₂ O when the desired colour intensity is attained | |
| 13. Counterstaining and blueing | |
| 14. Mounting: aqueous with AEC, permanent with DAB | |

Quality control

We recommend carrying out a positive and a negative control with every staining run. The positive control permits the validation of appropriate processing of the sample. If the negative control has a positive result, this points to unspecific staining.

Expected results

During the reaction of the substrate with horse radish peroxidase in the presence of a chromogen, a coloured precipitate is formed at the location of the bound primary antibody. This reaction only takes place if the target antigen is existent in the tissue. The chromogen used determines the colour of the precipitate. The analysis is carried out using a light microscope.

Limitations of the procedure

Immunohistochemistry is a complex method in which histological as well as immunological detection methods are combined. Tissue processing and handling prior to immunostaining, for example variations in fixation and embedding or the inherent nature of the tissue can cause inconsistent results (Nadji and Morales, 1983).

Endogenous peroxidase or pseudoperoxidase activity may cause non-specific staining. The enzyme activity is blocked by incubation with hydrogen peroxide solution. Tissues containing Hepatitis B Surface Antigen (HBsAg) may give false positive results with HRP (horse radish peroxidase) detection systems (Omata et al, 1980). Inadequate counterstaining and mounting can influence the interpretation of the results. The colour intensity of the reaction product can decrease with time, especially when exposed to light.

Overexposure with the protein blocking solution ("Blocking Solution") can result in decreasing signal intensity. Therefore, we recommend washing away the Blocking Solution instead of just draining it away as in other procedures.

Monosan[®] guarantees that the product will meet all requirements described from its shipping date until its expiry date, as long as the product is correctly stored and utilized. No additional guarantees can be given. Under no circumstances shall Monosan[®] be liable for any damages arising out of the use of the reagent provided.

Troubleshooting

If you observe unusual staining or other deviations from the expected results which could possibly be caused by the reagents, please read these instructions carefully, contact Monosans' technical support or your local distributor.

No staining on an actually positive control slide:

1. Reagents were not used in the proper order.
2. Chromogenic substrate solution was too old.
3. Bleaching because chromogen and mounting medium are incompatible.
4. The antigen/epitope in the tissue was insufficiently accessible to the primary antibody. Try a pre-treatment such as heat pre-treatment or enzyme digestion. If you used a pre-treatment it should be extended.
5. Primary antibody not from mouse or rabbit, but from a different species.
6. The antigen/epitope was not stable in the fixation and/or pre-treatment procedure used. Try another fixation or pre-treatment.

Weak staining:

1. Inadequate fixation or overfixation.
2. Incomplete deparaffinisation.
3. The antigen/epitope in the tissue was insufficiently accessible to the primary antibody. If you used heat pre-treatment or enzyme digestion it should be extended.
4. Excessive incubation with Blocking Solution or insufficient washing after this step.
5. Too much wash buffer remains on the slides after washing, diluting the reagents applied in the next step.
6. Incubation times were too short or primary antibody concentration too low.
7. Chromogenic substrate solution was too old.

Nonspecific background staining or overstaining:

1. Incomplete deparaffinisation.
2. Excessive tissue adhesive on slides.
3. Insufficient washing especially after the incubation with the enzyme polymer or the chromogenic substrate solution. These washings are critical.
4. Tissue was allowed to (partially) dry out with reagents on.
5. Unspecific binding of the primary antibody. Please use the Blocking Solution provided with this kit or dilute the primary antibody in appropriate diluents.
6. Incubation time of the primary antibody was too long or primary antibody concentration too high.
7. Incubation time of the chromogenic substrate solution was too long or reaction temperature too high (e.g. if temperature in the laboratory is high).
8. The substrate is metabolised by endogenous horse radish peroxidase in the tissue. Maybe the hydrogen peroxide solution used for blocking was inactivated.

Performance characteristics










Monosan[®] has conducted studies to evaluate the performance of the kit reagents. The product has been found to be suitable for the intended use.

Bibliography

Elias JM "Immunohistopathology – A practical Approach to Diagnosis" ASCP Press 2003
 Nadjji M and Morales AR Ann N.Y. Acad Sci 420:134-9, 1983
 Omata M et al. Am J Clin Pathol 73(5): 626-32, 1980

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Explanation of the symbols on the product label:

	Bestellnummer Catalog Number Reference du catalogue		Chargenbezeichnung Batch Code Code du lot		Reizend Irritant Irritant
	Gesundheitsschädlich Harmful Nocif		Giftig Toxic Toxique	 Hersteller / Manufacturer / Fabricant Monosan@ Frontstraat 2c 5405 PB Uden The Netherlands Tel: (+31) 413 251115 Fax: (+31) 413 266605 info@monosan.com www.monosan.com	
	Verwendbar bis Use By Utiliser jusque				
	Gebrauchsanweisung beachten Consult Instructions for use Consulter les instructions d'utilisation		Lagerungstemperatur Temperature Limitation Limites de température		