

## Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

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siehe unsere Liefer- und Versandbedingungen

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- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

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# Instructions For Use ABC-IFU

Rev. Date: June 27, 2019

**Revision: 3** 

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P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 - Fax (435) 755-0015 - www.scytek.com

## SensiTek Anti-Mouse

**Description:** Polyclonal secondary antibody conjugated to biotin for 3-step immunohistochemistry protocols.

Formulated to provide optimal staining with an incubation for 15-20 minutes. May be used with

automated systems, reagent jars, and manual dropping/pipetting.

Target: Mouse Species of Origin: Goat

Antigen Specificity: Anti-Mouse IgG+IgM (H+L)
Preadsorbed Against: Human, Bovine, Horse

Uses/Limitations: Not to be taken internally.

For In-Vitro Diagnostic use only.

Histological applications.

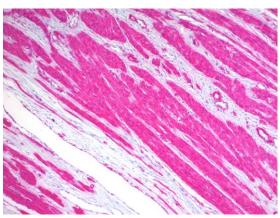
Do not use if reagent becomes cloudy. Do not use past expiration date. Use caution when handling reagent.

Non-Sterile.

Control Tissue: Any well-fixed tissue.

### **Availability/Contents:**

<u>volume</u>
8 ml
15 ml
125 ml
500 ml
1000 ml



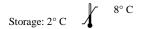
Smooth Muscle Actin demonstrated with Permanent red and SensiTek Anti-Mouse within an IHC protocol.

**Storage:** Store at 2-8°C. Product is stable for 18 months from date of manufacture.

#### Procedure:

Allow reagents to come to room temperature before use.

- 1. Deparaffinize and rehydrate tissue section.
- If needed, incubate slide in hydrogen peroxide for 10-15 minutes to reduce nonspecific background staining due to endogenous peroxidase.
- 3. Wash 2 times in buffer.
- 4. If required, incubate tissue in digestive enzyme.
- 5. Wash 4 times in buffer.









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- 6. Place slide in protein block (ScyTek's "Superblock") and incubate 5-10 minutes at room temperature to block nonspecific background staining. Note: Do not exceed 10 minutes or there may be a reduction in desired stain.
- 7. Wash 1 time in buffer.
- 8. Apply primary antibody and incubate according to manufacturer's protocol.
- 9. Wash 4 times in buffer.
- 10. Place slide in biotinylated anti-mouse, and incubate for 15-20 minutes at room temperature.
- 11. Wash 4 times in buffer.
- 12. Place slide in enzyme label, and incubate per instructions
- 13. Rinse 4 times in buffer.
- 14. Place slide in appropriate chromogenic substrate and incubate until desired reaction is achieved.
- 15. Counterstain and coverslip.

### **Troubleshooting Guide**

#### Overstaining:

- 1. Concentration of the primary antibody was too high or the incubation time was too long.
- 2. Temperature during incubation was too high.
- 3. Incubation time with link antibody or streptavidin/enzyme label was too long.

### **Nonspecific Background Staining:**

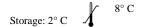
- 1. Rinsing between steps was inadequate.
- 2. Tissue was allowed to dry with reagents on.
- 3. Folds in tissue trapped reagents.
- 4. Tissue contains endogenous enzyme.
- 5. Tissue contains endogenous biotin.
- 6. Antigen migrated in tissue.
- 7. Excessive tissue adhesive on slides.
- 8. Inadequate blocking with protein block.

### Weak Staining:

- 1. Primary antibody concentration was too low or incubation time was too short.
- 2. Reagents are past their expiration date.
- Reagent is reaching the end of its useful life.
- 4. Counterstain or mounting media were incompatible and dissolved the chromogen reaction product.
- 5. Room temperature was excessively cool.
- 6. The primary antibody does not recognize an antigen that survives fixation and embedding in high enough amounts.
- 7. Excessive incubation with protein block (Super Block or normal serum).

### No Staining:

- 1. Steps were inadvertently left out.
- 2. There is no relevant antigen in the tissue.
- 3. The primary antibody is not of mouse origin.
- 4. Chromogenic substrate does not match enzyme label.
- 5. One or more components have been inactivated.











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#### References:

- 1. Kawasaki, N., Matsuo, Y., Yoshino, T., Yanai, H., Oka, T., Teramoto, N., ... Akagi, T. (1996). Metastatic Potential of Lymphoma/Leukemia Cell Lines in SCID Mice Is Closely Related to Expression of CD44. *Japanese Journal of Cancer Research*, 87(10), 1070–1077. https://doi.org/10.1111/j.1349-7006.1996.tb03112.x
- 2. Manabe, M., Mizoguchi, M., Niwa, M., Bertolino, A. P., Ishidoh, K., Kominami, E., & Ogawa, H. (1996). Assembly of Hair Keratins in Transfected Epithelial Cells. *Biochemical and Biophysical Research Communications*, 229(3), 965–973. https://doi.org/10.1006/bbrc.1996.1909
- 3. Manabe, M., Yaguchi, H., Butt, K. I., O'guin, W. M., Loomis, C. A., Sung, T.-T., & Ogawa, H. (1996). Trichohyalin Expression in Skin Tumors: Retrieval of Trichohyalin Antigenicity in Tissues by Microwave Irradiation. *International Journal of Dermatology*, *35*(5), 325–329. <a href="https://doi.org/10.1111/j.1365-4362.1996.tb03632.x">https://doi.org/10.1111/j.1365-4362.1996.tb03632.x</a>

