



# SZABO SCANDIC

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# Instructions For Use

## AFT-IFU

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## Alcian Blue (pH 1.0) Stain Kit

### Description and Principle

The Alcian Blue (pH 1.0) Stain Kit is intended for use in the histological visualization of strongly sulfated mucosubstances.

Alcian Blue, a copper phthalocyanine dye, binds acid mucosubstances. When used in a pH 1.0 acid solution Alcian Blue selectively stains sulfated acid mucosubstances. Acid mucins that are carboxylated only are protonated and will not be stained.

### Expected Results

Strongly Sulfated Mucosubstances:	Blue
Nuclei:	Red
Background:	Pink

### Kit Contents

Kit Contents	Storage
1. Alcian Blue Solution (pH 1.0)	18-25°C
2. Hydrochloric Acid Solution (1N)	18-25°C
3. Nuclear Fast Red (Enhanced Stability)	18-25°C

### Suggested Controls (not provided)

Tissue known to be positive for sulfated mucins. e.g. deep mucosa of colon

### Uses/Limitations

For In-Vitro Diagnostic use only.

Do not use if reagents become cloudy or precipitate

Do not use past expiration date.

Use caution when handling reagents.

Non-Sterile

Intended for FFPE sections cut at 5-10µm.

This procedure has not been optimized for frozen sections.

Frozen sections may require protocol modification.

### Storage

Store kit and all components at room temperature (18-25°C).

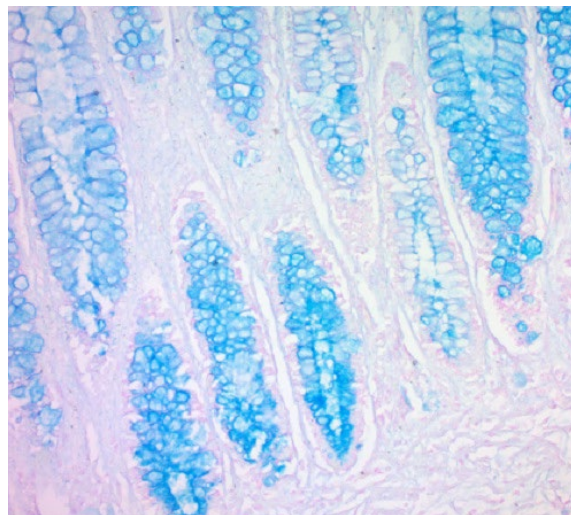
### Safety and Precautions

Please see current Safety Data Sheets (SDS) for this product and components GHS classification, pictograms, and full hazard/precautionary statements.

### Procedure notes:

1. Maintaining proper pH is critical to preventing false-positive mucin staining. 'Working Rinse Solution' should fall within pH 1.0 ± 0.15 and is used both before and after the Alcian Blue to control pH. Rinsing with deionized water or any other rinse may affect pH and cause non-specific staining. We recommend running at least one slide without the counterstain to compare intensity with slides that have been counterstained. Below procedure is written for slides laying horizontally that are stained by applying a small amount of solution.

2. In tissues with low-to-moderately sulfated mucins, such as human stomach, the addition of Methanol and Sodium Chloride (neither provided) may be need to be added to the post rinsing solution (step 5) to prevent nonspecific staining for accurate relative quantification<sup>6</sup>. First rinse in 0.1N HCl (Working Rinse Solution) containing 10% (v/v) Methanol three times then rinse three times with 0.1N HCl containing 0.5M NaCl.



Sulfated Mucins stained with Alcian Blue Solution (pH 1.0) on Mouse Intestine. Magnification 200X

### Procedure

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Make up sufficient amount of 'Working Rinse Solution' (0.1N HCl) by mixing the following:

**1 part** of Hydrochloric Acid Solution (1N)  
**9 parts** Deionized water

**Notes:** -An example mixture would be 10mls Hydrochloric Acid Solution + 90mls Deionized Water  
-We suggest making at least 10mls per slide. A smaller amount is required for step 3 and a larger amount used in step 5 for rinsing.

3. Apply a small amount (<2ml/slide) of 'Working Rinse Solution' to tissue for 30 seconds to adjust pH in preparation for staining. Save remaining 'Working Rinse Solution' for step 5.
4. Drain slide and without rinsing, stain tissue section with Alcian Blue Solution (pH 1.0) solution for 30 minutes
5. Quickly and thoroughly rinse excess stain off slide using remaining 'Working Rinse Solution'.
6. Carefully blot and allow slide to air dry.
7. If preferred, counter stain in Nuclear Fast Red (Enhanced Stability) for 1-2 minutes with occasional agitation. Rinse very briefly in deionized water and allow to completely air dry again.
8. Clear, and mount in synthetic resin.

## References

1. Cowper, M.; Frazier, T.; Wu, X.; Curley, J.L.; Ma, M.H.; Mohiuddin, O.A.; Dietrich, M.; McCarthy, M.; Bukowska, J.; Gimble, J.M. Human Platelet Lysate as a Functional Substitute for Fetal Bovine Serum in the Culture of Human Adipose Derived Stromal/Stem Cells. *Cells* 2019, 8, 724. <https://doi.org/10.3390/cells8070724>
2. Sheenan, D.C.; Hrapchak, B.B. *Theory and Practice of Histotechnology*, 2nd Edition. Battelle Press, Columbus, OH. Pages 172-173.
3. Churukian, C.J., 1989, *Manual of Special Stains Laboratory*, 4th Edition. University of Rochester, Rochester, New York. Pages 55-56.
4. Carson, F.L., 1996, *Histotechnology; A Self-Instructional Text*, 2nd Edition. ASCP Press, Chicago, IL. Pages 117-121.
3. Lillie, R.D. 1977, H.J. *Conn's Biological Stains*, 9th Edition. Williams & Wilkins, Baltimore. Pages 452-455.
6. János Tamás Padra, Sara K Lindén, Optimization of Alcian blue pH 1.0 histo-staining protocols to match mass spectrometric quantification of sulfomucins and circumvent false positive results due to sialomucins. *Glycobiology*, Volume 32, Issue 1, January 2022. Pages 6–10, <https://doi.org/10.1093/glycob/cwab091>



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