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# Data Sheet (Cat.No.T36957)



#### **BODIPY 493/503**

#### **Chemical Properties**

CAS No.: 121207-31-6

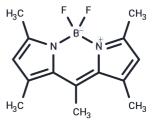
Formula: C14H17BF2N2

Molecular Weight: 262.11

Appearance: no data available

Storage: keep away from direct sunlight

Powder: -20°C for 3 years | In solvent: -80°C for 1 year



#### **Biological Description**

Description

| 1,0,0         | 493/503 nm. BODIPY 493/503 localizes to polar lipids and can be used to label cellular neutral lipid contents and for live and fixed cell applications.  |  |  |  |
|---------------|--|--|--|--|
| Targets(IC50) | Others   |  |  |  |
| In vitro      | <ul> <li>METHODS: Flow cytometry was used to detect cellular lipid droplets:</li> <li>1. BODIPY 493/503 is dissolved in 5 mM DMSO stock solution and diluted 1:2500 in PBS to a 2 μM working solution prior to use.</li> <li>2. Cultivate cells under culture conditions relevant to the study, e.g. 50,000 A498 cells in 35 mm wells. Overnight incubation of cells with 30 μM oleic acid serves as a positive control for increased neutral lipid content.</li> <li>3. At the time point of interest, prepare a 2 μM BODIPY staining solution in PBS. The</li> </ul>                               |  |  |  |
| Caldell       | volume of staining solution required for each sample corresponds to the volume of medium used to incubate the cells.  4. Rapidly rinse the cells with 3 mL of PBS to remove the medium/serum. Incubate in BODIPY Staining Solution for 15 min at 37°C in the dark.  5. Rapidly rinse the cells with 3 mL of PBS to remove the staining solution. Trypsinize the cells to produce a single-cell suspension. Add 5 mL of PBS and transfer the cell suspension to a 15 mL conical tube.   |  |  |  |
|               | <ul> <li>6. Centrifuge cells at 250×g for 5 min at 4°C. Remove the supernatant, quickly rinse the cell sediment with 3 mL of PBS, and centrifuge again, 250 × g, 5 min, 4°C.</li> <li>7, Remove the supernatant and resuspend the cells in 300 μL of 1× flow cytometry buffer for flow cytometry assay. [1]</li> <li>METHODS: Fluorescent microscopy to detect cellular lipid droplets:</li> <li>1. Dissolve BODIPY 493/503 into 1 mg/mL ethanol stock solution, and add 10 μL of 1 mg/ml BODIPY 492/503 stock solution to 10 mL of 150 mM NaCl to prepare a working solution before use.</li> </ul> |  |  |  |
|               | 2. One or two days before staining, culture the cells on sterile glass coverslips. Plate the cells at 50%-70% fusion to keep them semi-fused during staining.  3. To enhance lipid droplet formation and facilitate detection, supplement cell growth medium with 400 µM acid salt for 6-24 h prior to fixation and lipid droplet staining.  4. Rinse cells twice with 2 mL of PBS. Fix cells by incubating with 2 mL of 3% (w/v) paraformaldehyde for 30 min at room temperature.   |  |  |  |

BODIPY 493/503 (Pyrromethene 546) is a lipophilic fluorescent probe with Ex/Em of

5. Rinse the cells three times with 2 mL PBS. Cells were covered with 1 mL of BODIPY

493/503 Working Solution and incubated for 10 min at room temperature, protected from ambient light.

6. Wash cells three times with 2 mL PBS. Mount coverslips onto slides using 20-40  $\mu$ L of anti-fade mounting medium.

7, Detect BODIPY 493/503 staining of lipid droplets using fluorescence microscopy. [2]

#### **Solubility Information**

Chloroform: Soluble Methanol: Soluble

Ethanol: 0.24 mg/mL (908.38 uM)

DMSO: 2 mg/mL (7.63 mM), Sonication is recommended. (< 1 mg/ml refers to the product slightly soluble or insoluble)

#### **Preparing Stock Solutions**

|       | 1mg       | 5mg       | 10mg       |
|-------|-----------|-----------|------------|
| 1 mM  | 3.8152 mL | 19.076 mL | 38.1519 mL |
| 5 mM  | 0.763 mL  | 3.8152 mL | 7.6304 mL  |
| 10 mM | 0.3815 mL | 1.9076 mL | 3.8152 mL  |
| 50 mM | 0.0763 mL | 0.3815 mL | 0.763 mL   |

Please select the appropriate solvent to prepare the stock solution, according to the solubility of the product in different solvents. Please use it as soon as possible.

#### Reference

Xiong Q, Sun H, Wang Y, et al.Lipid droplet accumulation in Wdr45-deficient cells caused by impairment of chaperone-mediated autophagic degradation of Fasn.Lipids in Health and Disease.2024, 23(1): 91.

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