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### Methylated & Non-methylated pUC19 DNA Set

Standards for DNA methylation analysis workflows

#### Highlights

- · Ideal positive and negative controls for methylation detection assays.
- · In situ spike-in controls for monitoring bisulfite conversion efficiency in Next-Generation Sequencing experiments.
- Provided pUC19MN primer pair allows for convenient assay quality control.

Catalog Numbers: D5017



Scan with your smart-phone camera to view the online protocol/video.







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#### **Product Contents**

Methylated & Non-methylated pUC19 DNA Set	D5017	Storage Temp.
Methylated pUC19 DNA	20 ng/20 µl	-20°C
Non-methylated pUC19 DNA	20 ng/20 µl	-20°C
pUC19MN Primers	20 µl	-20°C

### **Specifications**

#### Methylated pUC19 DNA

- Source pUC19 plasmid purified from Dam<sup>-</sup>, Dcm<sup>-</sup> E. coli [enzymatically methylated by M.Sssl Methyltransferase (EC 2.1.1.37)].
- Concentration 1 ng/µl in TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.0).

#### Non-methylated pUC19 DNA

- Source pUC19 plasmid purified from Dam<sup>-</sup>, Dcm<sup>-</sup> E. coli.
- Concentration 1 ng/µl in TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.0).

#### pUC19MN Primers

- Concentration 20 µM each primer in TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 8.0).
- Primer sequences –

pUC19MN Forward Primer:

5' – GGTTATAGTTGTTTTTTGTGTGAAATTGTTATT – 3'

pUC19MN Reverse Primer:

5' - CTAACCTTTTACTCACATATTCTTTCCTAC - 3'

#### **Product Description**

The Methylated & Non-methylated pUC19 DNA Set consists of two control DNAs (methylated and non-methylated) along with a set of specifically designed primers that can be used in conjunction with the EZ DNA Methylation<sup>™</sup>, EZ DNA Methylation-Gold<sup>™</sup>, EZ DNA Methylation-Direct<sup>™</sup>, and EZ DNA Methylation-Lightning<sup>™</sup> kits from Zymo Research to assess the efficiency of bisulfite-mediated conversion of DNA. These plasmids can be used in conjunction with genomic DNA to provide internal controls to assess bisulfite conversion efficiency or to produce known mixtures of methylated and non-methylated DNA for assay calibration.

The **Non-Methylated pUC19 DNA** is pUC19 DNA that was isolated from a methylation-negative strain of *E. coli* (Dam<sup>-</sup>, Dcm<sup>-</sup>) and can be used as a negative control for DNA methylation analysis. The **Methylated pUC19 DNA** is pUC19 DNA that has been isolated from the same strain and has been enzymatically methylated at all cytosine positions comprising CG dinucleotides by M.SssI methyltransferase<sup>1</sup> (EC 2.1.1.37; Figure 1) and can be used as a positive control for DNA methylation analysis.

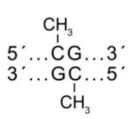


Figure 1. M.SssI methyltransferase methylates all cytosine residues in the double stranded CpG context.

The primer set herein has been designed to amplify a fragment of the supplied pUC19 DNA following bisulfite treatment. The methylated cytosines, comprising CG dinucleotides in the **Methylated pUC19 DNA** remain unconverted following bisulfite treatment, whereas nonmethylated cytosines are converted into uracil and detected as thymine after PCR. The supplied pUC19 DNA has been linearized at position 2177 using Scal endonuclease.

<sup>&</sup>lt;sup>1</sup>Nur et al. J. Bacteriol. 164: 19-24 (1985).

#### **Recommended Usage**

The **Methylated and Non-methylated pUC19 DNA Set** can be used in a variety of methylation analysis applications including bisulfite and methylation-specific PCR, *in situ* spike-in controls, methylation sensitive high resolution melt analysis, methylation arrays, methylated DNA immunoprecipitation (MeDIP), library preparation, and more.

#### Protocol

For best results, it's important to ensure the DNA is completely homogenous and fully in solution before quantification and usage. The following steps are recommended before quantification and usage<sup>1</sup>:

- 1. Bring the standards to room temperature.
- 2. Mix the standards well, briefly spin down for 5-10 seconds.
- 3. Repeat Step 2 twice, for three times total.
- 4. Proceed with quantification or usage.

<u>**Bisulfite Conversion</u></u>: For most applications 5-50 pg of plasmid may be used as an internal spike-in control for bisulfite conversion reactions containing 250 ng to 2 \mug of genomic DNA. Refer to the kit specifications for setup of the bisulfite conversion reaction.</u>** 

Bisulfite PCR Setup: The following is designed for a 25 µl reaction.

Component	Volume	Final Concentration
pUC19MN Primers <sup>2</sup>	Variable	0.2 to 0.8 µM each
Bisulfite-converted DNA <sup>3</sup>	1 µl	Up to 0.25 pg/µl
10 mM dNTP mix	0.5 µl	0.2 mM each dNTP
Standard PCR Buffer	Variable	1X
MgCl <sub>2</sub> or MgSO <sub>4</sub>	Variable	1-4 mM, if needed
ZymoTaq™ DNA Polymerase <sup>4</sup>	Variable	1-2 units
Nuclease Free Water	Bring reaction to 25 µl	N/A

#### Recommended Thermocycler Conditions:

- A. 95 °C, 10 minutes
- B. 95 °C, 30 seconds
- C. 57 °C, 30 to 60 seconds
- D. 72 °C, 60 seconds
- E. Repeat steps B through D an additional 29 to 39 times

depending on the polymerase used.

- F. 72 °C, 7 minutes
- G. 4 °C

<sup>&</sup>lt;sup>1</sup>Standards are quantified using NanoDrop<sup>®</sup> measurements. If using other methods, variation may be observed. <sup>2</sup>Alternatively, you may substitute primers of your choice.

<sup>&</sup>lt;sup>3</sup>Remember to bisulfite-treat the DNA prior to performing PCR.

<sup>&</sup>lt;sup>4</sup>We recommend using **ZymoTag™ DNA Polymerase** or other hot-start DNA polymerases for amplification of bisulfite-treated DNA.

#### Appendix

#### pUC19MN Bisulfite PCR

The expected PCR amplicon for the Methylated & Non-methylated pUC19 DNA is 362 bp, corresponding to nucleotide positions 464 to 825 of the pUC19 sequence, including the regions (italicized) that hybridize to the primers.

<u>Original sequence of pUC19 for bisulfite treatment and PCR amplification (sense strand 5' to 3').</u> The cytosines (underlined) in the CpG dinucleotide context (bold capital letters) are methylated enzymatically by M.SssI methyltransferase:

5′ -	ggtcatagct	gtttcctgtg	tgaaattgtt	<i>atc<mark>C</mark>G</i> ctcac
aattccacac	aacata <b><u>C</u>G</b> ag	c <b>CG</b> gaagcat	aaagtgtaaa	gcctggggtg
cctaatgagt	gagctaactc	acattaattg	<b><u>C</u>G</b> ttg <b>CG</b> ctc	actgcc <b>CG</b> ct
ttccagt <b><u>C</u>G</b> g	gaaacctgt <b>C</b>	$\mathbf{G} \texttt{tgccagctg}$	cattaatgaa	t <b>CG</b> gccaa <b>CG</b>
<b><u>C</u>GCC</b> gggaga	gg <b>CG</b> gtttg <u>C</u>	<b>G</b> tattggg <b>C</b> G	ctcttc <b><u>C</u>G</b> ct	tcct <b>CG</b> ctca
ctgact <b><u>C</u>G</b> ct	g <u>C</u> Gct <u>C</u> Ggt <u>C</u>	Gtt <u>C</u> Ggctg <u>C</u>	<b>G</b> g <b>C</b> Gag <b>C</b> Ggt	atcagctcac
tcaaagg <b>CG</b> g	taata <b>CG</b> gtt	atccacagaa	tcaggggata	a <b>C</b> Gcaggaaa
gaacatgtga	gcaaaaggcc	ag - 3'		

#### Expected sequence of the above PCR amplicon following bisulfite treatment:

<u>Methylated pUC19 DNA:</u> Below is the expected sequence for the Methylated pUC19 DNA after bisulfite conversion and PCR (sense strand). Methylated cytosines in the CpG dinucleotide context remain unconverted following bisulfite treatment, whereas non-methylated cytosines, or cytosines not in the CpG context, are converted to uracil and detected as thymine after PCR.

5 <b>′</b> –	ggttatagtt	gttttttgtg	tgaaattgtt	<i>att<b>C</b>G</i> tttat
aattttatat	aatata <b><u>C</u>G</b> ag	t <b>CG</b> gaagtat	aaagtgtaaa	gtttggggtg
tttaatgagt	gagttaattt	atattaattg	<u>C</u> Gttg <u>C</u> Gttt	attgtt <b>CG</b> tt
ttttagt <b>CG</b> g	gaaatttgt <b>C</b>	<b>G</b> tgttagttg	tattaatgaa	t <b>CG</b> gttaa <b>CG</b>
<b><u>C</u>G<u>C</u>G</b> gggaga	gg <u>C</u> Ggtttg <u>C</u>	<b>G</b> tattggg <b><u>C</u>G</b>	tttttt <b>CG</b> tt	tttt <b><u>C</u>G</b> ttta
ttgatt <b>CG</b> tt	g <b>CG</b> tt <b>CG</b> gt <u>C</u>	GttCGgttgC	<b>G</b> g <b>CG</b> ag <b>C</b> Ggt	attagtttat
ttaaagg <b><u>C</u>G</b> g	taata <b>CG</b> gtt	atttatagaa	ttaggggata	a <b>C</b> Gtaggaaa
gaatatgtga	gtaaaaggtt	ag - 3'		

<u>Non-methylated pUC19 DNA:</u> Below is the expected sequence for the nonmethylated pUC19 DNA after bisulfite conversion and PCR (sense strand). During treatment with sodium bisulfite, non-methylated cytosines are converted to uracil, which are later detected as thymine after PCR.

```
5' - ggttatagtt gttttttgtg tgaaattgtt att<u>T</u>Gtttat
aatttatat aatata<u>T</u>Gag t<u>T</u>Ggaagtat aaagtgtaaa gtttggggtg
tttaatgagt gagttaattt atattaattg <u>T</u>Gttg<u>T</u>Gttt attgtt<u>T</u>Gtt
ttttagt<u>T</u>Gg gaaatttgt<u>T</u> Gtgttagttg tattaatgaa t<u>T</u>Ggttaa<u>T</u>G
<u>T</u>GTGgggagag gg<u>T</u>Ggtttg<u>T</u> Gtattggg<u>T</u>G tttttt<u>T</u>Gtt tttt<u>T</u>Gttt
ttgatt<u>T</u>Gtt g<u>T</u>Gtt<u>T</u>Ggt<u>T</u> Gtt<u>T</u>Ggttg<u>T</u> Gg<u>T</u>Gag<u>T</u>Ggt attagtttat
ttaaagg<u>T</u>Gg taata<u>T</u>Ggtt atttatagaa ttagggata a<u>T</u>Gtaggaaa
gaatatgtga gtaaaaggtt ag - 3'
```

## **Ordering Information**

Product Description	Catalog No.	Size
Human Methylated & Non-methylated DNA Set	D5014	5 µg/20 µl
Human HCT116 DKO Non-Methylated DNA	D5014-1	5 µg/20 µl
Human HCT116 DKO Methylated DNA	D5014-2	5 µg/20 µl
Bisulfite-Converted Universal Methylated Human DNA Standard	D5015	1 µg/50 µl
Zymo <i>Taq</i> ™ qPCR Premix	E2054 E2055	50 Rxns. 200 Rxns.
Zymo <i>Taq</i> ™ Premix	E2003 E2004	50 Rxns 200 Rxns.
EZ DNA Methylation-Lightning™ Kit	D5030 D5031	50 Rxns 200 Rxns.
EZ DNA Methylation-Direct™ Kit	D5020 D5021	50 Rxns 200 Rxns.
EZ DNA Methylation™ Kit	D5001 D5002	50 Rxns 200 Rxns.
EZ DNA Methylation-Gold™ Kit	D5005 D5006	50 Rxns 200 Rxns.

### Notes


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